Estradiol alters cholecystokinin stimulus-response coupling in rat pancreatic acini

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Blevins, George T., J r., Sandra S. McCullough, Tasha N. Wilbert, Remelle M. Isom, Parimal Chowdhury, and Stephanie T. Miller. Estradiol alters cholecystokinin stimulus-response coupling in rat pancreatic acini. Am. J. Physiol. 275 (Gastrointest. Liver Physiol. 38): G993–G998, 1998.—We have previously demonstrated that altered exocrine pancreatic stimulus-secretion coupling is associated with ovariectomy and chronic estradiol administration. To elucidate possible mechanisms underlying those effects we examined the ability of chronic administration of different doses of estradiol to regulate the CCK signal transduction pathway in isolated rat pancreatic acini. Doses of estradiol ranging from 0.5 to 119 µg/day were administered to ovariectomized rats for 18 days. Ovariectomy was associated with enhanced CCK-stimulated pancreatic amylase release, whereas estradiol dose dependently decreased the magnitude of CCK-stimulated amylase release. Ovariectomy was also associated with enhanced CCK receptor numbers on acinar cell membranes. Estradiol administration was associated with dose-dependent decreases in CCK receptor numbers. Neither ovariectomy nor estradiol administration affected CCK receptor affinity. Moreover, estradiol administration was associated with increased expression of the α subunit of the heterotrimeric G protein Gq11 (Gq11). Recent findings (H. Ohnishi, S. A. Ernst, D. I. Yule, C. W. Baker, and J. A. Williams. J. Biol. Chem. 272: 16056–16061, 1997) demonstrate that Gq11 may exert a tonic inhibitory effect on pancreatic enzyme release. In view of these findings, the increased expression of Gq11 induced by estradiol likely contributes to the inhibition of pancreatic enzyme release. We conclude that the effect of estradiol to decrease pancreatic secretion is mediated through regulation of CCK receptor density and Gq11 expression.

MATERIALS AND METHODS

Materials. The following were purchased: collagenase from Worthington Biochem (Freehold, NJ), synthetic cholecystokinin octapeptide (CCK-8) from Peninsula Laboratories (Belmont, CA), CCK-8 labeled by Bolton-Hunter reagent method (125I-labeled BH-CCK-8, specific activity 2,000 Ci/mmol) from Amersham (Arlington Heights, IL), Eagle's medium amino acid supplement from GIBCO (Grand Island, NY), 21-day timed-release pellets containing vehicle or estradiol from Innovative Research of America (Sarasota, FL), procion yellow dye MX-8G from Polysciences (Warrington, PA), and estradiol double antibody kit from Diagnostic Products (Los Angeles, CA). Unless otherwise noted all other chemicals were purchased from Sigma Chemical (St. Louis, MO).

Animals. The Institutional Animal Care and Use Committee of the University of Arkansas for Medical Sciences approved this study, and the study was performed in accordance with the Institutional Animal Care and Use Committee guidelines.

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with the National Institutes of Health Guide for Care and Use of Laboratory Animals.

Female Sprague-Dawley rats obtained from the Charles River breeding colony were housed in plastic cages with wire covers, in a temperature-controlled room (22 ± 2°C), on a 12:12-h light-dark cycle (lights on at 6 AM and off at 6 PM), and allowed ad libitum access to standard laboratory rat chow. On day 1 of the study, the respective surgery was performed and 21-day timed-release pellets containing vehicle or estradiol were implanted subcutaneously. Ovariectomized rats were administered either vehicle or estradiol at one of three doses (0.5, 24, or 119 µg/day), whereas sham ovariectomized rats were administered vehicle. On day 18 rats fasted for 18 h were killed by decapitation. Blood was removed by exanguination using heparinized tubes and centrifuged at 1,500 g for 10 min, and plasma was saved frozen at −80°C for later determination of plasma estradiol. Pancreata and uteri were removed, and wet weights were measured. Sham ovariectomized rats were utilized at random times during the estrous cycle.

Preparation of isolated pancreatic acini and amylase release. Subsequent to removal the pancreas was trimmed free of fat and lymph nodes, and enzymatic digestion was used to prepare isolated pancreatic acini as described previously (2). Acini were preincubated and resuspended in 10% HEPES-buffered Ringer solution (pH 7.4), supplemented with 11.1 mM glucose, minimum Eagle’s medium amino acid solution, 0.5% bovine serum albumin, and 0.02% soybean trypsin inhibitor. Aliquots (3 ml) of acinar suspension were incubated with various concentrations of CCK-8 for 30 min at 37°C with shaking at 60 cycles/min and 100% O2 gas phase. Duplicate flasks were used to determine stimulated amylase release for each concentration of CCK.

Preparation of enriched acinar cell membranes for [125I]-labeled BH-CCCK-8 binding. Pancreatic acinar cell membranes were prepared from aliquots of acini obtained after the preincubation step of acinar cell isolation. Acini were kept on ice at 4°C during all steps of membrane preparation. Acini were washed in nine volumes of buffer B (0.2 M sucrose, 10 mM HEPES, 0.5 mM phenylmethylsulfonyl fluoride, 1 mM benzamidine, at pH 7.4) and sedimented by centrifugation at 300 g for 5 min. The supernatant was removed by suction and discarded, and the washing and centrifugation steps were repeated once more. The pellet obtained was then diluted with nine volumes of buffer A (1.35 M sucrose, 10 mM HEPES, pH 7.4, 0.5 mM phenylmethylsulfonyl fluoride, 1 mM benzamidine) and homogenized using six 6-s bursts of a sonicator, with a 1-cm tip at a setting of 40. The homogenate was then decanted into 14 × 95 mm centrifuge tubes and buffer B was layered over the top. These tubes were centrifuged at 100,000 g for 120 min. The membrane fraction at the interface between the two layers was collected. These membranes were diluted fivefold with buffer C (20 mM HEPES, 5 mM MgCl2, 1 mM EGTA, 100 mM NaCl, at pH 6.5) and centrifuged at 30,000 g for 35 min. The pellet of enriched membranes was resuspended in buffer C, and a small aliquot was removed for measurement of protein concentration. The remainder was dissolved in gel-loading buffer for immunoblot analysis and stored at −80°C until used.

Immunoblot measurement of Goq expression. One dimensional gel electrophoresis was performed as described by Laemmli (18). Twenty micrograms of protein per sample were loaded in each lane of 12% SDS-PAGE minigels and run at 200 V. After gel electrophoresis, proteins were transferred to nitrocellulose membranes according to the method of Towbin et al. (28) using glycin-Tris-methanol transfer buffer containing 0.1% SDS to enhance the transfer from the gel at 30 V overnight. Nitrocellulose membranes were blocked with Tris-buffered saline (2.42 g Tris base, 8.0 g NaCl, pH 7.6) plus Tween 20, containing 5% bovine serum albumin for 1 h. Membranes were then washed and probed with anti-Goq11 antisemur (Dupont-NE) at a concentration of 1:10,000 for 2 h. Membranes were washed again and then probed for 90 min with donkey anti-rabbit antibody conjugated with horseradish peroxidase. Membranes were then washed five additional times, and bands were visualized using enhanced chemiluminescence following the manufacturer’s procedures, by exposing the membranes to Dupont Reflection Autoradiography Films. For quantitation, film images were scanned using a flat bed scanner and saved as image files. Band densities were quantitated using the Un-Scan-It gel analysis program (Silk Scientific, Orem, UT).

Assays. Amylase activity of incubation media and calibration standards (Sera Chem, Sigma Chemical) were measured using the method of Jung (15), with procion yellow starch as substrate. Protein was determined by the method of Bradford (5). Four milliliters of 1:5 diluted Bio-Rad dye reagent were added to each sample of protein standard or unknown, and bovine serum albumin was used as standard.

Plasma estradiol was measured using a commercial radioimmunoassay kit (Double Antibody kit, Diagnostic Products). [125I]-labeled compounds were counted using a Packard Auto-Gamma 5650 gamma counter at 70% counting efficiency.

Data analysis. All measurements were made in duplicate and are expressed as means ± SE of the average of duplicate determinations. Statistical significance was tested by analysis of variance with the appropriate multiple range test. P < 0.05 was considered statistically significant. Binding data were analyzed using the LIGAND least-squares curve fitting program of Munson and Rodbard (19).
RESULTS

Effects of estradiol administration on plasma estradiol, uterine weight, and pancreatic weight. Plasma estradiol levels of ovariectomized rats treated with either vehicle or 0.5 µg estradiol per day were below the sensitivity of the estradiol radioimmunoassay (Table 1), and for the purpose of statistical analysis the value of radioimmunoassay sensitivity (5 pg/ml) was assigned to these groups. Plasma estradiol of these two groups was significantly less than that of sham ovariectomized rats or ovariectomized rats treated with 24 or 119 µg estradiol per day. Doses of 24 and 119 µg estradiol per day yielded measurable plasma estradiol levels and mean plasma estradiol of rats receiving 119 µg estradiol per day was significantly greater than any of the other groups.

Ovariectomy resulted in a significant (P < 0.05) decrease in uterine weight compared with that of sham ovariectomized rats, and chronic estradiol treatment of ovariectomized rats was associated with an apparent dose-dependent increase in uterine weights (Table 1). No significant differences in pancreatic weights were found between any of the groups examined.

Estradiol influence on amylase content and release from exocrine pancreas. The cellular amylase content of acini isolated from estradiol-treated ovariectomized rats appeared to increase in a dose-dependent manner (Fig. 1). Cellular amylase content was significantly (P < 0.01) increased by 119 µg estradiol per day, compared with acini isolated from any of the other three groups. There appeared to be a trend toward dose-dependent increases in basal amylase release in ovariectomized rats treated with estradiol, but no significant differences were observed (data not shown).

Chronic treatment of rats with estradiol was associated with apparent dose-dependent decreases in the magnitude of percent initial amylase release from acini in response to graded doses of CCK-8 (Fig. 2). The maximal percentage of initial cellular amylase released by acini isolated from ovariectomized rats was 34.4 ± 1.9%. Maximal release was decreased to 31.2 ± 4.1% by 0.5 µg estradiol per day, to 23.3 ± 3.4% by 24 µg estradiol per day, and to 9.4 ± 1.6% by 119 µg estradiol per day. Estradiol at a daily dose of 24 µg significantly decreased percent initial amylase release at doses of CCK-8 ranging from 10 to 300 pM, compared with ovariectomized rats treated with either vehicle or 0.5 µg estradiol per day. Percent initial amylase release was significantly decreased by 119 µg estradiol per day, at doses of CCK-8 ranging from 3 pM to 1 nM, compared with all other groups.

Effects of estradiol on CCK receptor binding. The affinity of the CCK receptor on pancreatic acinar cell membranes was not significantly affected by estradiol treatment of ovariectomized rats (Table 2). However, CCK receptor capacity was significantly greater on membranes from vehicle-treated ovariectomized rats compared with ovariectomized rats treated with 24 or 119 µg estradiol per day.

Table 1. Effects of ovariectomy and estradiol on plasma estradiol, uterine, and pancreatic weights of rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>n</th>
<th>Plasma E2 (pg/ml)</th>
<th>Uterine Wt, mg</th>
<th>Pancreatic Wt, mg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham Ovx+ veh</td>
<td>6</td>
<td>25.3 ± 1.3*</td>
<td>351 ± 55*</td>
<td>1,120 ± 55*</td>
</tr>
<tr>
<td>Ovx+ veh</td>
<td>8</td>
<td>&lt;5</td>
<td>102 ± 6</td>
<td>1,153 ± 40</td>
</tr>
<tr>
<td>Ovx+ 0.5 µg E2/day</td>
<td>6</td>
<td>&lt;5</td>
<td>211 ± 16</td>
<td>1,140 ± 29</td>
</tr>
<tr>
<td>Ovx+ 24 µg E2/day</td>
<td>5</td>
<td>188 ± 62</td>
<td>322 ± 55†</td>
<td>1,141 ± 61</td>
</tr>
<tr>
<td>Ovx+ 119 µg E2/day</td>
<td>6</td>
<td>613 ± 140‡</td>
<td>772 ± 60‡</td>
<td>1,196 ± 11</td>
</tr>
</tbody>
</table>

Values are means ± SE; n = no. of measurements. E2, estradiol; Ovx, ovariectomized; veh, vehicle. Eighteen days after surgery and initiation of estradiol treatment, plasma samples were collected, uteri and pancreata were removed, and wet weight was measured. Commercial radioimmunoassay kit was used to measure plasma estradiol. *Significantly (P < 0.05) greater than Ovx+ veh. †Significantly (P < 0.05) greater than Ovx, and Ovx+ 0.5 µg E2/day. ‡Significantly (P < 0.05) greater than all other groups.

Fig. 1. Total cellular amylase content of pancreatic acini isolated from sham ovariectomized (Sham), ovariectomized (Ovx), and estradiol (E2)-treated ovariectomized rats. Rats were implanted with 21-day time-release pellets that yielded the daily doses indicated. After 18 days rats were killed, pancreata were removed, acini were isolated, and CCK-stimulated amylase release was measured. *Significantly (P < 0.05) greater than all other groups.

Fig. 2. Amylase release expressed as a percentage of initial cellular amylase content from pancreatic acinar cells isolated from sham ovariectomized (S-Ovx), ovariectomized, and estradiol-treated ovariectomized rats. Veh, vehicle. *Significantly (P < 0.05) less than Ovx veh. **Significantly (P < 0.05) less than Ovx veh and 0.5 µg E2. ***Significantly (P < 0.05) less than all other groups.
Examination of possible short-term estradiol effects.

To determine whether the effects of estradiol could be observed over the short term, estradiol was included in the incubation medium along with 100 pM CCK, at concentrations between 1 pM and 100 nM. In this acute study no apparent effect of estradiol was observed on CCK-8-stimulated amylase release or total CCK binding (data not shown).

Effects of estradiol on $G_{\alpha_{q11}}$ expression in the exocrine pancreas.

Ovariectomy had no significant effect on pancreatic $G_{\alpha_{q11}}$ expression compared with sham ovariectomized rats (data not shown). However, treatment of ovariectomized rats with increasing concentrations of estradiol was associated with significant ($P < 0.001$) increases in $G_{\alpha_{q11}}$ expression (Fig. 3).

DISCUSSION

We have previously observed that chronic ovariectomy is associated with increased sensitivity of the exocrine pancreas to CCK stimulation and increased CCK receptors on pancreatic acinar cell membranes, whereas a dose of estradiol in the upper physiological range was associated with both suppression of the magnitude of CCK-stimulated amylase release and a decrease in the number of CCK receptors (2). The present study examined the influence of chronic estrogen deficiency induced by ovariectomy and chronic exposure to a range of doses of estradiol on pancreatic acinar cell amylase content, CCK-stimulated amylase secretion, CCK receptor binding characteristics, and expression of $G_{\alpha_{q11}}$. Plasma estradiol levels were also measured, and uterine weights of ovariectomized rats treated with vehicle or estradiol were used to assess the bioactivity of the estradiol preparation employed in these studies (14, 21). Plasma estradiol of the rats administered the two highest doses attained levels found during pregnancy and in individuals receiving very high doses of estradiol (27).

The dose-dependent increases in total cellular amylase associated with chronic estradiol administration are likely the result of increases in either synthesis or storage of amylase. The decreased responsiveness of acini from estradiol-treated rats suggests that the increased total cellular amylase is probably due to increased intracellular amylase stores resulting from decreased stimulated secretion. The present study did not directly address the question of estradiol effects on amylase synthesis.

In the present study we found that chronic administration of estradiol to ovariectomized rats was associated with dose-dependent decreases in the magnitude of CCK-stimulated amylase release. Guo and Singh (13) found that total stimulated amylase release was significantly decreased when ovariectomized guinea pigs were administered a daily injection of estradiol. This finding indicates that chronic estradiol administration decreases the responsiveness of CCK stimulus-response coupling in pancreatic acini.

Recently, there has been increased interest in the nongenomic effects of steroid hormones, and membrane-bound binding sites for estrogens have been found in the brain (6). Whether such receptors exist in the exocrine pancreas is not presently known. However, pancreatic tissue has a great capacity to retain estro-

<table>
<thead>
<tr>
<th>Treatment</th>
<th>n</th>
<th>$K_d$, pM</th>
<th>$B_{max}$, pmol/mg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham Ovx + veh</td>
<td>6</td>
<td>62.6 ± 5.9</td>
<td>373.8 ± 26.2*</td>
</tr>
<tr>
<td>Ovx + veh</td>
<td>8</td>
<td>51.8 ± 3.6</td>
<td>1,051.6 ± 115.5</td>
</tr>
<tr>
<td>Ovx + 0.5 µg E/day</td>
<td>6</td>
<td>55.0 ± 10.0</td>
<td>791.8 ± 153.6</td>
</tr>
<tr>
<td>Ovx + 24 µg E/day</td>
<td>5</td>
<td>55.2 ± 4.6</td>
<td>405.5 ± 42.2*</td>
</tr>
<tr>
<td>Ovx + 119 µg E/day</td>
<td>6</td>
<td>67.9 ± 10.3</td>
<td>424.4 ± 107.3*</td>
</tr>
</tbody>
</table>

Values are means ± SE; n = no. rats. Pancreatic acinar cell membranes were incubated with 7 pM 125I-labeled BH-CCK-8 and doses of synthetic CCK-8 ranging from 10 pM to 100 nM. At 150 min, incubation mixture was filtered and washed, and bound radioactivity was determined. Equilibrium dissociation constant ($K_d$) and maximal binding capacity ($B_{max}$) were determined using LIGAND nonlinear curve fitting program. *Significantly ($P < 0.01$) less than Ovx + veh.

Fig. 3. A: representative immunoblot of enriched rat pancreatic membranes from ovariectomized rats chronically treated with increasing doses of estradiol. Blot was probed with antisera raised against $G_{\alpha_{q11}}$. B: pooled data of $G_{\alpha_{q11}}$ band densities, measured by scanning and densitometry using the Un-Scan-it program (Silk Scientific; n = 5). *Significantly ($P < 0.01$) greater than 0 or 0.5 µg estradiol/day.
gens, and the mechanism underlying this ability is an estrogen binding protein that is localized mainly, if not exclusively, to cytosol of pancreatic acini (25, 32, 33). This estrogen binding protein constitutes 2–4% of total cytosolic protein and binds estrogens with characteristics distinct from those of the uterine estrogen receptor (4, 22). The pancreatic estrogen binding protein does not translocate into isolated nuclei, and on fractionation of pancreatic tissue, most of the estrogen binding activity is localized to the microsomal fraction. Grossman et al. (12), using immunocytochemical methods, have shown that the pancreatic estrogen binding protein is distributed in both the endoplasmic reticulum and mitochondria. The presence of this estrogen binding protein in such considerable proportions and its distinct estrogen binding characteristics suggest that it plays an as yet undefined role in exocrine pancreatic physiology. Based on localization it is reasonable to speculate that if pancreatic estrogen effects are mediated through this protein, estrogens may possibly have effects on cellular calcium metabolism and/or translation of mRNA. Whether the observed effects of estrogens on the exocrine pancreas are mediated partially or wholly through the activity of this pancreatic estrogen binding protein is unclear. The findings of estradiol-induced alterations in amylase content and release in the present study may suggest that this binding protein subserves a regulatory function in the exocrine pancreas.

The observed estrogen-induced alteration of CCK receptor binding characteristics is one mechanism that may at least partially account for the altered responsiveness of acini to CCK. CCK receptor affinity was not significantly altered by either ovariectomy or ovarietomy plus estradiol. We have previously shown that pancreatic membranes prepared from ovariectomized rats exhibit a significantly greater number of CCK receptors than those of sham ovariectomized rats (2). The findings of the present study show that while ovarian insufficiency is associated with increased acinar cell CCK receptor capacity, chronic estrogen administration is associated with dose-dependent decreases in CCK receptor capacity. Increased CCK binding capacity on membranes prepared from acini isolated from ovariectomized rats parallels increased responsiveness of pancreatic acini to CCK stimulation, whereas decreased CCK binding capacity on membranes prepared from estradiol-treated rats dose dependently parallels decreases in responsiveness of pancreatic acini to CCK-8. It is likely that the altered responsiveness to CCK-8 of pancreatic acini isolated from ovariectomized rats is at least partially the consequence of alterations in CCK receptor numbers. The mechanisms through which estrogens induce these effects on pancreatic CCK receptors are not clear.

Because the CCK receptor on acinar cells exists in multiple interconvertible binding affinity states (3), it is possible that estrogens may influence the affinity state of the receptor. It is also possible that chronic estradiol administration may be associated with altered receptor internalization or insulation (24). Although the most dramatic decrease in CCK-8-stimulated amylase release was observed in rats treated with 119 µg estradiol per day, there was no further decrease in the number of CCK receptors between that group and the 24 µg estradiol per day group, suggesting that alterations in stimulus-secretion coupling downstream of the CCK receptor may also play a role in the effects observed at this highest dose of estradiol.

The mechanism through which binding of CCK to its receptor leads to activation of phospholipase C is believed to be via the interaction of the ligand bound receptor with a guanine nucleotide binding protein (30). Expression of the Gα subunit of Gq/11, the heterotrimeric guanine nucleotide binding protein believed to couple to the CCK receptor, was examined by Western blotting to further examine estrogen effects on the CCK signal transduction pathway as a possible mechanism mediating the alterations in secretion. Expression of Gq/11 was dose dependently increased in acinar cell membranes from ovariectomized rats treated with estradiol. Recently it has been found that Gq/11 is localized on pancreatic zymogen granules and involved in calcium-regulated amylase secretion (20). Onishi et al. (20), utilizing a known antagonist of Gq/11, demonstrated that antagonism of Gq/11 was associated with a concentration-dependent potentiation of calcium-stimulated amylase release. The authors concluded that Gq/11 on zymogen granules plays a tonic inhibitory role in calcium-regulated amylase secretion from pancreatic acini. In view of these findings, the dose-dependent increases in the expression of Gq/11 in estradiol-treated rats may be involved in the inhibition of CCK-stimulated amylase release from estradiol-treated acini. It is possible that estrogens also influence stimulus-secretion coupling at other points in the CCK signal transduction pathway or may be associated with altered receptor G protein coupling. Our present data do not rule this out but do demonstrate that the observed effects of estradiol on amylase secretion are likely mediated by alterations in CCK receptor numbers and Gq/11 expression.

The most obvious manner through which estradiol might influence the expression of CCK receptors and Gq/11 is via the classical estrogen receptor, acting to induce changes in transcriptional mechanisms (16). It will be interesting to determine whether the change in receptor density on acinar cell membranes and Gq/11 expression is the result of modified transcriptional or translational mechanisms resulting in altered production of these proteins. Moreover, estradiol may possibly be acting through membrane-bound estradiol binding sites. The present study examined the influence of ovariectomy and estradiol administration on CCK-mediated signal transduction mechanisms. Whether the effects of estradiol are specific for the CCK-9 signaling pathway or may influence signaling mediated by other acinar cell stimulants such as bombesin, vasoactive intestinal peptide, and acetylcholine is not known. These questions are presently under investigation.

In conclusion, we have characterized the effects of chronic estradiol administration on pancreatic acinar
cell amylase secretion stimulated by CCK. We have also demonstrated that estrogens decrease CCK receptor binding while increasing Gαq expression. These observations provide new insight for understanding the mechanisms through which estrogens influence exocrine pancreatic function.

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