Part of quercetin absorbed in the small intestine is conjugated and further secreted in the intestinal lumen

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Crespy, Vanessa, Christine Morand, Claudine Manach, Catherine Besson, Christian Demigne, and Christian Remesy. Part of quercetin absorbed in the small intestine is conjugated and further secreted in the intestinal lumen. Am. J. Physiol. 277 (Gastrointest. Liver Physiol. 40): G120–G126, 1999.—Rutin and quercetin absorption and metabolism were investigated in rats after in situ perfusion of jejunum plus ileum (15 nmol/min). In contrast to rutin, a high proportion of quercetin (two-thirds) disappeared during perfusion, reflecting extensive transfer into the intestinal wall. Net quercetin absorption was not complete (2.1 nmol/min), inasmuch as 52% were reexcreted in the lumen as conjugated derivatives (7.7 nmol/min). Enterohepatic recycling contribution of flavonoids was excluded by catheterization of the biliary duct before perfusion. After a 30-min perfusion period, 0.71 µM of quercetin equivalents were detected in plasma, reflecting a significant absorption from the small intestine. The differential hydrolysis of effluent samples by glucuronidase and/or sulfatase indicates that the conjugated forms released in the lumen were 1) glucuronidated derivatives of quercetin and of its methoxylated forms (64%) and 2) sulfated form of quercetin (36%). In vitro quercetin glucuronides synthetized using jejunal and ileal microsomal fractions were similar to those recovered in the effluent of perfusion. These data suggest that glucuronidation and sulfatation take place in intestinal cells, whereas no glucuronosulfatoconjugates could be detected in the effluent. The present work shows that a rapid quercetin absorption in the small intestine is very effective together with its active conjugation in intestinal cells.

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Intestinal Absorption and Metabolism of Quercetin

Agronomique Ethics Committee, in accordance with decree no. 87-848.

Sampling Procedure

Rats were anesthetized with pentobarbital sodium (40 mg/kg body wt) 18 h after the beginning of the meal and maintained alive during the perfusion period.

After cannulation of the biliary duct, a perfusion of jejunal plus ileal segment of intestine (5 cm distal from the flexura duodenoejunalis to the valvula ileocecalis) was prepared by installing cannulas (external diam 4 mm, internal diam 2.5 mm) at each extremity. This segment was continuously perfused in situ with a buffer containing (in mM) 2.5 KH$_2$PO$_4$, 2.5 K$_2$HPO$_4$, 5 NaHCO$_3$, 50 NaCl, 50 KCl, 2 CaCl$_2$, 2 MgCl$_2$, 2.5 HEPES Na, 2.5 HEPES acid, pH 6.7, 8 glucose, and 1 taurocholic acid at a flow rate of 1 ml/min, at 37°C, and supplemented with 15 µM of quercitin or rutin. An aliquot of effluent was directly collected at the exit of the ileum in plastic tubes (1.5 ml) during the last 5 min of perfusion. The bile was collected all along the perfusion step.

At the end of the experiment, blood samples were withdrawn from the abdominal aorta into heparinized tubes. Plasma, bile, and perfusate samples were acidified with 10 mM acetic acid and then stored at −20°C.

HPLC Analysis

Samples treatment. Plasma and bile were spiked with 3.5 µmol/l of diosmetin (final concentration) used as an internal standard and acidified (to pH 4.9) with 0.1 vol of acetic acid, 0.58 M. The samples were treated for 30 min at 37°C with 5 × 10$^6$ U/l β-glucuronidase and 2.5 × 10$^6$ U/l sulfatase (for visualizing the total forms). The reactions were stopped by addition of 2.85 vol of acetone, and the resulting mixtures were centrifuged. This procedure was sufficient to totally extract quercetin bound to plasma albumin (6, 16). After this extraction step, 20 µl of supernatant were injected and analyzed by HPLC.

Effluent samples (100 µl) were acidified with 0.1 vol of acetic acid, 0.58 M, and directly analyzed or treated as follows: 1) incubated for 30 min at 37°C with 50 U β-glucuronidase and 2.5 U sulfatase from Helix pomatia, 2) incubated for 120 min at 37°C with 1,000 U of bovine β-glucuronidase, and 3) incubated for 240 min at 37°C with 50 U of aryl sulfatase from A. aerogenes.

In each case, the reactions were stopped by adding 150 µl of acetone. After centrifugation, a 20-µl aliquot of supernatant was injected in the HPLC system for analysis. The concentrations in conjugated derivatives present in a sample were estimated as the difference between the concentrations of quercetin recorded before and after the adequate enzymatic treatment.

Chromatographic conditions. The HPLC system used consisted of an autosampler (Kontron 360), an ultraviolet detector (set at 370 nm), and a software program for data recording and processing. The system was fitted with a 5-µm C-18 Hypersil based deactivated silica analytical column (150 × 4.6 mm; Life Sciences International, Cergy, France). The mobile phase consisted of water and H$_3$PO$_4$ (99.5:0.5, solvent A) and acetonitrile (solvent B).

To separate the conjugated metabolites of flavonoids, the chromatographic conditions were as follows (flow rate 1 ml/min): 0–2 min solvent A 85%/solvent B 15%; 2–22 min solvent A 85%/solvent B 15% → solvent A 60%/solvent B 40% 22–24 min solvent A 60%/solvent B 40% 24–27 min return to initial mobile phase conditions and then equilibration for 8 min.

In Vitro Assay of Flavonoid Glucuronidation

Jejunal and ileal microsomes from rat intestine were prepared by differential ultracentrifugation at 105,000 g at 4°C for 1 h. To prepare intestinal microsomes, mucosal scraping (50 cm for jejunal and 20 cm for ileal intestine) were homogenized in ice-cold buffer containing Tris-HCl (50 mM, pH 7.2), sucrose (100 mM), EDTA (10 mM), dithiothreitol (2 mM), leupeptin (1 µM), and trypsin inhibitor (25 mg/100 ml). The final microsomal pellet was suspended in a buffer containing 100 mM HEPES, pH 7.2, sucrose (100 mM), and trypsin inhibitor (25 mg/100 ml) and kept in a frozen state at −20°C until use. The preparations were adjusted to have a final protein concentration of about 5 mg/ml, measured according to the Pierce bicinchoninic acid protein reagent kit (Interchim, Montluçon, France).

Incubations were carried out as follows. In a final volume of 750 µl, 540 µl of buffer (HEPES, 75 mM, pH 7.3, MgCl$_2$, 10 mM), 50 µl of UDP-glucuronic acid (4.5 mM final), and 100 µl of microsomal suspension (50 µg protein) were activated in situ by 60 µl of a 0.2% solution of Triton X-100. The reaction was started by the addition of 2 µl of quercetin (18.75 mM in DMSO). Incubations were performed at 37°C for 3 h, and then aliquots of the reaction mixture were taken and treated (with or without β-glucuronidase (and sulfatase)) for HPLC analysis, exactly as previously described.

Statistics

Values are means ± SE, and the differences between values were determined by one-way ANOVA coupled with the Student-Newman-Keuls multiple-comparison test. Values of P < 0.05 were considered significant.

RESULTS

Comparison of In Situ Intestinal Perfusion of Rutin and Quercetin in Rats

When the intestine was perfused at a flow rate of 1 ml/min during 30 min with a buffer containing 15 µM rutin, the concentrations of this glycoside in the effluent collected at the extremity of the ileum were not significantly different from those initially present in the perfusate (Fig. 1). This indicates that rutin is not noticeably absorbed nor metabolized by the intestinal wall.

By contrast, when quercetin was perfused in the jejunoileal segment (at 14.7 ± 0.6 µM, with a flow rate of 1 ml/min), the concentration of this flavonol (free and conjugated forms) in the residual perfusate was significantly decreased (12.6 ± 0.2 µM; Fig. 1). Thus 14.3% of perfused quercetin was taken up by the intestinal wall. Moreover, in plasma hydrolyzed by β-glucuronidase (and sulfatase), the concentration of quercetin was about 0.71 ± 0.06 µM (Table 1), giving additional evidence for the absorption of quercetin through the intestinal wall. Preliminary experiments have established that before perfusion no trace of quercetin was present in rat plasma (data not shown).

Differential Hydrolysis of Effluent Resulting From Intestinal Perfusion of Quercetin

The representative HPLC profile of the nonhydrolyzed effluent (Fig. 2A) is characterized by 1) a peak of quercetin at a concentration of 4.9 ± 0.4 µM (indicating

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that ~33% of infused quercetin was not metabolized by the intestinal cells) and 2) the presence of five unidentified peaks. Four of these peaks (1–4) totally disappeared after hydrolysis of the samples with β-glucuronidase (and sulfatase) from Helix pomatia (Fig. 2B), suggesting that they corresponded to quercetin conjugates. This HPLC profile (Fig. 2B) was also characterized by the presence of two small peaks (at 24.47 min and 24.72 min) corresponding, respectively, to trace amounts of isorhamnetin (about 0.2 µM) and tamarixetin (<0.1 µM), the methoxylated forms of quercetin. Moreover, the treatment of the effluent with the β-glucuronidase (and sulfatase) extract induced only a 50% decrease in the intensity of peak 5 (Fig. 2B), whereas this peak totally disappeared when the effluent was hydrolyzed by a sulfatase from A. aerogenes (Fig. 2C). This indicates that peak 5 should represent sulfated conjugate which is only partially hydrolyzed by the sulfatase component present in the Helix pomatia extract.

When the effluent was treated with the sulfatase alone (Fig. 2C), no trace of isorhamnetin and tamarixetin was detected in this fraction, and the concentration of quercetin was estimated at 7.7 ± 0.4 µM. This indicates that the methoxylated forms of quercetin were not sulfated in the intestine and that the sulfate derivative of quercetin corresponded to 2.8 ± 0.3 µM of quercetin equivalent (calculated as the difference between the concentration of quercetin measured before and after the enzymatic treatment).

The hydrolysis of the effluent with bovine β-glucuronidase led to a complete disappearance of peaks 1–4 (Fig. 2D), whereas peak 5 was unaffected and the resulting concentration of quercetin was about 9.8 ± 0.6 µM. In such conditions, the concentration in quercetin glucuronides present in the effluent was estimated to be about 4.9 ± 0.2 µM. Moreover, isorhamnetin (~0.2 µM) and tamarixetin (~0.1 µM) are present in Fig. 2D, indicating that these compounds are submitted to an intestinal glucuronidation.

The total concentration of quercetin recovered in the hydrolyzed effluent was 12.6 ± 0.2 µM, 4.9 ± 0.2 µM (namely 39%) being unconjugated forms of quercetin and 7.7 ± 0.4 µM (namely 61%) being conjugated derivatives. These conjugates are composed of 36% sulfate derivatives and 64% glucuronides; a minor part of them could be methoxylated.

**Quercetin Fluxes After Intestinal Perfusion**

Whereas the total flux of quercetin in the perfusion was 14.7 ± 0.6 nmol/min (flow rate 1 ml/min), only 4.9 ± 0.4 nmol/min of quercetin was recovered intact in the effluent. The secretion of conjugated derivatives from the wall into the lumen could explain why the net absorption of quercetin was low compared with its total transfer into the intestinal wall (Fig. 3). By contrast, the net flux of quercetin absorption (2.1 ± 0.4 nmol/min corresponding to 14.3% of total perfused quercetin) was markedly lower than the secretion flux of conjugates from the wall into the lumen (7.7 ± 0.4 nmol/min, namely 52.4% of total perfused quercetin) (Fig. 3). The sum of these fluxes corresponded to the net transfer of quercetin into the intestinal wall (9.8 ± 0.5 nmol/min). Therefore, 78.5% of absorbed quercetin was directly conjugated and eliminated by the intestine. In comparison to intestinal secretion, the biliary secretion flux was quite limited (Fig. 3).

**In Vitro Glucuronidation of Quercetin by Intestinal Wall**

The in vitro capacity of rat jejunal and ileal fractions to transfer glucuronic acid from UDP-glucuronic acid to quercetin has been tested using microsomal preparations. It could be noted that the capacity of the jejunal wall to conjugate quercetin appeared significantly greater than that of the ileal wall (Fig. 4); the percentage of total quercetin metabolized in the jejunal microsomal fraction was 43 vs. 32% in ileal one. In both

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**Table 1. Quercetin and quercetin derivative concentrations in various biological samples after perfusion of rat intestine with quercetin**

| Quercetin conjugates, µM | %Hydrolyzed | % | Nonhydrolyzed
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<td></td>
<td></td>
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<tr>
<td>Excreted</td>
<td></td>
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<tr>
<td>Influent</td>
<td>7.7 ± 0.4</td>
<td>97.2</td>
<td>2.00</td>
</tr>
<tr>
<td>Effluent</td>
<td>0.71 ± 0.06</td>
<td>100</td>
<td>ND</td>
</tr>
<tr>
<td>Bile</td>
<td>54.9 ± 3.1</td>
<td>43</td>
<td>30</td>
</tr>
<tr>
<td>Plasma</td>
<td>0.71 ± 0.06</td>
<td>100</td>
<td>ND</td>
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Values are means ± SE; n = 8 rats. ND, not detectable. Conjugated forms, glucuronosulfato derivatives of quercetin and of its methoxylated forms. Amounts of conjugated derivatives were calculated by subtracting conjugate values from total ones.
cases, the same four metabolites (referred to as a-d, according to their increasing hydrophobicity) were present on the HPLC profiles (Fig. 4). These peaks corresponded to the glucuronidated forms of quercetin.

The HPLC profile of nonhydrolyzed effluent resulting from quercetin perfusion (Fig. 2A) has been compared with that obtained after in vitro glucuronidation of quercetin by microsomal jejunal and ileal fractions (Fig. 4). By taking into account the respective relative retention times compared with diosmetin, peaks 1, 2, and 4 (Fig. 2A) exhibited a perfect coincidence with those noted b, c, and d, obtained by the in vitro glucuronidation procedure (Fig. 4). Peak 3 could correspond to glucuronidated-methoxylated forms of quercetin (but this point was not tested). These data support the view of the existence of an UDP-glucuronyltransferase activity using quercetin as a substrate at the intestinal level.

Concentrations of Quercetin and of Its Methoxylated Forms in Various Biological Samples

At the end of the perfusion period, the concentration of quercetin in hydrolyzed plasma was about 0.71 ± 0.06 µM (Table 1). By contrast, in nonhydrolyzed plasma no trace of free quercetin could be found, indicating that the circulating forms of quercetin are glucurono-sulfoconjugates. However, these quercetin metabolites were not detected in nonhydrolyzed samples, probably because their responsiveness was too low in the present chromatographic conditions (Table 1).

In all experiments a cannulation of the biliary duct was performed to test the enterohepatic cycling of quercetin. The biliary secretion was quite noticeable at the end of the perfusion; about 55 µM of quercetin and of its methoxylated forms were found in the hydrolyzed bile samples. Quercetin constituted the main form of biliary excretion (43%), the other metabolites beingisorhamnetin and tamarixetin (30 and 27%, respectively) (Table 1). This high level of biliary excretion accounts for a very active hepatic metabolism of quercetin.

DISCUSSION

In the present experimental pattern, rutin was not absorbed and metabolized in the small intestine. These data are in accordance with those of Manach et al. (16), which showed that the absorption of rutin was delayed.
compared with that of quercetin. Indeed, the hydrolysis of rutin takes place in the cecum because the microflora possesses glycosidases liable to hydrolyze glycosides with a high degree of specificity (3). Then the free quercetin produced can be absorbed, eliminated, or degraded in phenolic acids (1, 4, 30).

In contrast to rutin, the absorption of quercetin from the small intestine appears very effective, inasmuch as 66.7% of perfused quercetin is absorbed by the small intestine, although the perfusion rate is relatively high (0.9 µmol/h of quercetin, which after extrapolation represents 6.5 mg/day). A recent study dealing with the transcellular transport of quercetin in the intestine using a Caco-2 cell monolayer model (32) corroborates the present finding. The study by Walgren et al. (32) reported that the average recovery in the basolateral side was to 67% of medium quercetin with an apical loading in this molecule.

A significant part of the quercetin recovered in the effluent was conjugated. These conjugated forms of quercetin have an intestinal origin because the biliary duct was cannulated before the beginning of the perfusion (leading to a disruption of the enterohepatic cycle). The intestinal metabolism of flavonoids is still poorly known; nevertheless previous studies performed on rats have already shown that glucuronide and sulfate forms of 1-naphthol and estrogens were excreted in the lumen during in situ perfusion of the intestine (2, 26). Thus the intestinal metabolism of quercetin and that of quercetin fluxes in different biological samples after intestinal infusion. Data are means ± SE; n = 8. Ileal plus jejunal segments were perfused with 15 µM quercetin for 30 min at flow rate of 1 ml/min as described in MATERIALS AND METHODS. The present quercetin fluxes have been calculated using data from Fig. 1. Biliary flow rate was 13.6 ± 0.7 µl/min. A, quercetin perfusion flux; B, free quercetin recovered in effluent; C, net transfer of quercetin into intestinal wall; D, net secretion of conjugated quercetin from intestinal wall into lumen; E, effective absorption of quercetin; F, biliary flux of conjugated forms.

![Fig. 3](image)

**Fig. 3.** Quercetin fluxes in different biological samples after intestinal infusion. Data are means ± SE; n = 8. Ileal plus jejunal segments were perfused with 15 µM quercetin for 30 min at flow rate of 1 ml/min as described in MATERIALS AND METHODS. The present quercetin fluxes have been calculated using data from Fig. 1. Biliary flow rate was 13.6 ± 0.7 µl/min. A, quercetin perfusion flux; B, free quercetin recovered in effluent; C, net transfer of quercetin into intestinal wall; D, net secretion of conjugated quercetin from intestinal wall into lumen; E, effective absorption of quercetin; F, biliary flux of conjugated forms.

![Fig. 4](image)

**Fig. 4.** Representative HPLC chromatograms obtained after glucuronidation of 50 µM quercetin with microsomal preparations from jejunum (A) and ileum (B) walls. In vitro glucuronidation procedure was detailed in MATERIALS AND METHODS. After a 3-h incubation period at 37°C, aliquots of the reaction mixture were directly extracted with acetone and glucuronidated forms of quercetin were visualized by HPLC analysis. Q, quercetin; D, diosmetin (internal standard). a, b, c, d, Glucuronide derivatives.
these compounds seems quite similar. In view of this, Sfakianos et al. (27) have described an in vivo glucuronidation of an isoflavone (genistein) by the intestinal wall.

The existence of an intestinal UDP-glucuronyltransferase activity was confirmed by the present in vitro experiments of quercetin glucuronidation from jejunal and ileal microsomal preparations. The glucuronidation yield measured on these fractions appears lower than that observed with liver microsomes (18); however, it was still noticeable. Glucuronidation was not the sole conjugative activity present in the intestine since methoxylated forms of quercetin (3- and 4-O-methylquercetin) are released after hydrolysis of the O-glucuronides. Indeed, the activity of catechol-O-methyltransferase has been reported to be maximum in the liver and kidneys (29), it is also present in the enterocytes (21). These cells can combine processes of glucuronidation and methylation. The glucuronidation has been described as a rapid process (2, 22) that takes place for the high concentrations of metabolites (12). This could explain why the percentage of glucuronidation in the intestine (64%) appeared greater than that of methylation (4%) (12). This result is in accordance with those of Piskula and Terao (21), which showed that the glucuronidation of epicatechin was more important than the methylation in the intestine. Moreover, these data have been obtained in vitro, and additional experiments could be performed to test in vivo their validity. In keeping with a previous report (25), the present work shows the existence of a sulfotransferase activity in intestinal cells. It is well established that whatever its origin, the sulfotransferase activity is inversely related to the concentration of quercetin (25, 28, 33). However, in the present conditions of perfusion, even if the glucuronidation prevailed (64%), sulfatation was substantial (36% of the total conjugation). At the intestinal level, quercetin was sulfated but no trace of mixed glucurono-sulfoconjugates was found. By contrast, it has been shown (18) that in rats adapted to a quercetin diet the major circulating forms of quercetin are constituted by glucurono-sulfoconjugates. The absence of these compounds at the intestinal level strongly suggests that they are preferentially synthetized in the liver.

According to the present data and to those found in the literature (11, 15, 16, 18), it could be possible to depict the major features of quercetin absorption and the metabolism at the intestinal level (Fig. 5). Quercetin enters the cells by a still unidentified mechanism. In these cells, quercetin is readily glucuronidated, methoxylated, and sulfated. The resulting metabolites can subsequently leave the intestinal cell possibly via a facilitated transport system, either across the apical pole (secretion of the conjugated forms in the lumen) or across the basolateral side for their further transfer into the portal vein. However, disposal data are not sufficient to exclude that a part of the absorbed quercetin can circulate in the blood as aglycone (Fig. 5).

The liver further metabolizes the conjugated compounds arising from intestine, leading to an increase of their degree of glucuronidation, sulfatation, and methylation. Thus no trace of free quercetin could be detected in the plasma of rats infused with quercetin in the small intestine. This is in accordance with previous studies, in which it was reported that the circulating metabolites of quercetin recovered in the plasma are glucurono-sulfoconjugates with a high degree of methylation (16, 18). In vitro the rate of intestinal conjugation and of secretion of quercetin metabolites toward the lumen is quite substantial, but in vivo the intestinal secretion of conjugated derivatives could be lower. In vivo the presence of biliary quercetin conjugates in the lumen could decrease the concentration gradient between the enterocytes and the digestive content and therefore the flow of conjugates could be preferentially directed toward the basolateral side.

In the present work, both production and elimination of conjugates seen essentially occur via the intestinal pathway, rather than by the biliary route. Whatever their origin (biliary or intestinal), quercetin conjugates could be recycled after their metabolism by the microflora, in the large intestine (9). This phenomenon can contribute to extend the bioavailability of quercetin all along the light-dark cycle (9). However, bacterial degra-
Conjugation of quercetin into phenolic acids (4) may depress the recovery of the intestinal conjugates.

In conclusion, the intestinal secretion of quercetin conjugates constitutes an elimination pathway that should be taken into account, in addition with the urinary and biliary pathways. Nevertheless, it seems essential to characterize the transport systems for the aglycone and conjugated forms across the intestinal membrane.

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REFERENCES


