Serotonin stimulates endotoxin translocation via 5-HT₃ receptors in the rat ileum

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Abstract

Serotonin stimulates endotoxin translocation via 5-HT₃ receptors in the rat ileum. Am J Physiol Gastrointest Liver Physiol 284: G782–G788, 2003; 10.1152/ajpgi.00376.2002.—Because few previous studies have investigated the mechanisms of endotoxin translocation induced by intestinal obstruction, we aimed to clarify whether or not serotonin [5-hydroxytryptamine (5-HT)], which is released from enterochromaffin (EC) cells, is responsible for alterations of the mucosal permeability to endotoxin and to identify the 5-HT receptor subtypes that mediate this action. FITC-labeled LPS (FITC-LPS) was injected into the ileum of rats, and the FITC-LPS level in the superior mesenteric vein was subsequently measured by using a fluorescence spectrophotometer. To measure the 5-HT release induced by high intraluminal pressure, ex vivo preparation of vascularity and luminally perfused rat ileum was used. Results demonstrated that elevated intraluminal pressure stimulates the translocation of FITC-LPS and the release of 5-HT from the EC cells into the intestinal lumen but not into the portal circulation. This FITC-LPS translocation, which was stimulated by exogenously applied 5-HT in the lumen and the jugular vein, was inhibited by 5-HT₃ receptor antagonist administration both intamurally and intravenously. The stimulatory effect of elevated intraluminal pressure on the translocation of FITC-LPS was inhibited by the intraluminal and intravenous administration of 5-HT₃ receptor antagonist. These results suggest that 5-HT released from EC cells may be involved in the translocation of FITC-LPS induced by elevated intraluminal pressure and that this effect is mediated by 5-HT₃ receptors that may be located in the intestinal epithelium.

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the blood circulation. We used an in vivo model of the rat ileum to investigate the effect of 5-HT on the mucosal permeability induced by high intraluminal pressure and to identify the 5-HT receptor subtype that mediates this action.

MATERIALS AND METHODS

Measurement of FITC-labeled LPS translocated from ileum to superior mesenteric vein. Male Wistar rats weighing 500–600 g were fasted overnight (16–18 h) before the experiment. Animals were anesthetized with an intraperitoneal injection of pentobarbital sodium (50 mg/kg Nembutal; Abbott Laboratories). The terminal ileum, measuring ~10 cm in length, was prepared for in vivo luminal perfusion. Luminal injection was performed through a cannula inserted into the proximal end of the segment, and 0.1 M PBS (pH 7.4) containing 50 μg/ml of FITC-LPS (from Escherichia coli, serotype 0111B4; Sigma) was perfused at a speed of 0.5 ml/min and was drained from the distal end of the ileal segment. Blood samples were collected by using a heparinized catheter (0.5 mm ID, 0.8 mm OD, model SV 31; Natsume, Tokyo, Japan) inserted into the splenic vein with the tip placed in the superior mesenteric vein. Blood was collected in 2-ml samples for 45 min at 5-min intervals into ice-cold vials. Blood samples were centrifuged (3,000 rpm) for 5 min, and the plasma levels of FITC were measured by using a fluorescence spectrophotometer (model F-4500) at an excitation of 485 nm and an emission of 535 nm. After a 15-min equilibration period, the intraluminal pressure was raised to 40 mmHg during this time. Instead of the saline-injected controls, in some experiments, chemicals were injected intraluminally or intravascularly as described below, and the FITC levels in the blood were then compared with those of the nontreated controls. In some experiments, chemicals were injected intraluminally or intravascularly as described below, and the FITC levels in the superior mesenteric vein were compared with those of the saline-injected controls. Luminal injection was performed through a cannula inserted into the ileal segment, and vascular injection was performed through a catheter inserted into the jugular vein. 5-HT (Sigma), at a concentration of 10, 1.0, or 0.1 μmol·kg⁻¹·min⁻¹, was injected into the lumen for 15–35 min. 5-HT at a concentration of 0.1 or 0.01 μmol·kg⁻¹·min⁻¹ was injected into the jugular vein for 15–35 min. The luminal injection of 5-HT (10 μmol·kg⁻¹·min⁻¹) and vascular injection of 5-HT (0.1 μmol·kg⁻¹·min⁻¹) was combined with the luminal (10 μg·kg⁻¹·min⁻¹) or vascular (1 μg·kg⁻¹·min⁻¹) injection of methysergide, a 5-HT1/5-HT2 receptor dual antagonist (San- doz Pharmaceutical, Basle, Switzerland), the luminal (100 μg·kg⁻¹·min⁻¹) or vascular (10 μg·kg⁻¹·min⁻¹) injection of ketanserin, a 5-HT2 receptor antagonist (Janssen Pharmaceu- tical, Beerse, Belgium), the luminal (1 μg·kg⁻¹·min⁻¹) or vascular (0.1 μg·kg⁻¹·min⁻¹) injection of azaestrone, a 5-HT3 receptor antagonist (Yoshitomi Pharmaceutical), or the luminal (1 μg·kg⁻¹·min⁻¹) or vascular (0.1 μg·kg⁻¹·min⁻¹) injection of SB-204070, a 5-HT4 receptor antagonist (Glaxo SmithKline, London, UK). In these experiments, a 5-HT receptor antagonist was administered for 10–35 min. In some experiments, the luminal (1 μg·kg⁻¹·min⁻¹) or vascular (0.1 μg·kg⁻¹·min⁻¹) injection of azaestrone was performed for between 10 and 35 min, and the intraluminal pressure was raised to 40 mmH2O during this time. Instead of the above chemicals, saline was injected into the controls. The dose of each 5-HT receptor antagonist used in the present study was determined by the dose that affects the intestinal motility shown in the previous studies (26, 31), because no previous studies have examined the effects of 5-HT receptor antagonists on intestinal permeability.

Measurement of 5-HT release from vascularly and luminal perfused rat ileum. Male Wistar rats weighing 250–350 g were fasted overnight (16–18 h) before the experiment. Animals were anesthetized with an intraperitoneal injection of pentobarbital sodium (50 mg/kg Nembutal; Abbott Laboratories). The terminal ileum, measuring ~7 cm in length, was prepared for in situ vascular and intraluminal perfusion (9). The blood vessels that supply the stomach, duodenum, jejunum, proximal ileum, colon, pancreas, spleen, kidneys, and adrenal glands were excluded from perfusion. Arterial perfusion was performed by inserting a cannula into the aorta with its tip lying close to the superior mesenteric artery. The aorta was tied off above the celiac artery. The vascular effluent was collected through a portal vein cannula. Intraluminal perfusion was performed by inserting into the ileal segment with the tip in the midportion of the segment. This catheter was connected to a transducer (model TP-400T, Nihon Koden), and the pressure was amplified by an ampli- fier (model AP-601G; Nihon Koden). Intraluminal pressure was continuously monitored and recorded on a polygraph (model RM-6100; Nihon Koden).

The vascular perfusate consisted of Krebs solution containing 3% dextran, 0.2% bovine serum albumin, and 5 mM glucose. The perfusate was saturated with 95% O2-5% CO2 gas to maintain pH 7.4. The luminal perfusate was 0.1 M PBS, pH 7.4. The flow rate of vascular and luminal perfusion was maintained at 3 and 1 ml/min, respectively. After a 30-min equilibration period, the vascular and luminal effluents were collected for 30 min at 3-min intervals into ice-cold vials. At 18 min, the intraluminal pressure was raised by clamping the luminal effluent. The clamp was released when the pressure reached 40 cmH2O. In some experiments, TTX (Sankyo) was introduced into the vasculature to achieve a final concentration of 1 μM from 12 to 21 min, when the intraluminal pressure was high (≤40 cmH2O).

The level of 5-HT was determined by HPLC. Each 1 ml of effluent was added to 10 μl of 57 mM ascorbic acid, 10 μl of 10 mM disodium EDTA, 10 μl of 1 M perchloric acid, and 10 μl of 51 mM pargyline hydrochloride. Vascular effluent was filtered by using Ultrafree-MC (model 30000 MWL, Nihon Millipore) by centrifuging the samples for 30 min at 10,000 rpm at 4°C. Luminal effluent was filtered manually by using a 0.22-μm-pore desk filter (Milllex-GV; Nihon Millipore). Ali- quotes of 100 μl were subjected to HPLC, and the 5-HT content was measured (12).

Statistical analysis. Statistical analysis of the data shown in Figs. 1, 2, and 3A and B were performed by using a one-way ANOVA, followed by Fisher’s paired least significant difference test. The degree of changes in the FITC level (%stimulation) was calculated as: (the mean of the FITC-LPS levels marked with asterisks in the figures/mean of the FITC-LPS levels in nontreated or saline-injected controls) × 100. A paired t-test (two-tail) was used to compare the values between the saline-injected controls and those induced by chemical infusion. P values of <0.05 were considered significant.

RESULTS

Effect of high intraluminal pressure on FITC-LPS translocation. We injected 50 μg/ml of FITC-LPS into the ileal lumen and determined that the basal FITC-
LPS level in the superior mesenteric vein before the elevation of intraluminal pressure was 9.87 ± 0.29 ng/ml (n = 5). When we elevated the intraluminal pressure to 40 cmH₂O by clamping the luminal effluent, the FITC-LPS level in the superior mesenteric vein significantly increased (19.09 ± 0.47 ng/ml, n = 5) to a level 198.4 ± 10.3% (n = 5) of the basal level (Fig. 1).

Effect of high intraluminal pressure on 5-HT release from the vascularly and luminally perfused rat ileum. We examined the effect of elevated intraluminal pressure on 5-HT release in ex vivo vascularly and luminally perfused rat ileum. The basal level of 5-HT release from the perfused ileum into the lumen and the portal circulation was 4.63 ± 0.37 ng/ml (n = 3) and 3.84 ± 0.44 ng/ml (n = 3), respectively (Fig. 2). The luminal release of 5-HT significantly increased (30.86 ± 3.83 ng/ml, n = 3) with increasing intraluminal pressure (40 cmH₂O) to a level 528.0 ± 39.8% (n = 3) above the basal level. This stimulatory effect of the intraluminal pressure on the luminal release of 5-HT was not altered by the vascular injection of TTX (471.0 ± 98.1%) (data not shown). The vascular release of 5-HT was not affected by the increasing intraluminal pressure (5.0 ± 0.6 ng/ml) (Fig. 2).

Effect of 5-HT on FITC-LPS translocation. To clarify the role of 5-HT in the effect of high intraluminal pressure on FITC-LPS translocation, we administered 5-HT into the lumen as well as into the jugular vein. We subsequently measured the FITC-LPS level in the superior mesenteric vein (Fig. 3) and found that the basal level of FITC-LPS in the superior mesenteric vein (10.48 ± 0.14 ng/ml, n = 6) significantly increased (15.76 ± 0.47 ng/ml, 152.0 ± 8.54% of basal level, n = 6) with the intraluminal injection of 10 μmol·kg⁻¹·min⁻¹ of 5-HT (Fig. 3, A and C). However, the intraluminal administration of 1 and 0.1 μmol·kg⁻¹·min⁻¹ of 5-HT did not affect the basal level of FITC-LPS in the superior mesenteric vein (Fig. 3C), whereas it significantly increased from 10.48 ± 0.14 to 13.08 ± 0.54 ng/ml (133.7 ± 7.25% of basal level, n = 6) after the vascular injection of 0.1 μmol·kg⁻¹·min⁻¹ of 5-HT (Fig. 3, B and D). The vascular administration of 0.01 μmol·kg⁻¹·min⁻¹ of 5-HT did not affect the basal level of FITC-LPS detected in the superior mesenteric vein (Fig. 3D).

Effect of 5-HT receptor antagonists on the 5-HT-induced FITC-LPS translocation. To determine which receptor subtypes mediate the effects of 5-HT on FITC-LPS translocation, we intraluminally and intravenously injected 5-HT₁, 5-HT₂, 5-HT₃, and 5-HT₄ receptor antagonists combined with 5-HT, and the FITC-LPS level in the superior mesenteric vein was subsequently measured. The FITC-LPS level in the superior mesenteric vein, which was elevated by intraluminal and intravenous injection of 5-HT, significantly decreased after luminal (Fig. 4, A and D, respectively) or vascular (Fig. 4, B and C, respectively) injection of azastron, but not by the injection of methysergide, ketanserin or SB-204070.

Effect of 5-HT₃ receptor antagonists on FITC-LPS translocation induced by high intraluminal pressure. Finally, the involvement of the 5-HT₃ receptor in the FITC-LPS translocation induced by high intraluminal pressure was examined. We intraluminally and intravenously injected a 5-HT₃ receptor antagonist and simultaneously increased the intraluminal pressure to 40 cmH₂O, on which the FITC-LPS level in the superior mesenteric vein was measured. The stimulatory effect of the high intraluminal pressure on FITC-LPS translocation was completely inhibited by both intraluminal and intravenous administration of azastron (Fig. 5).

Fig. 1. Effect of high intraluminal pressure on FITC-labeled LPS (FITC-LPS) translocation. The FITC-LPS level in the superior mesenteric vein was 4.63 ± 0.37 ng/ml (n = 5) and 3.84 ± 0.44 ng/ml (n = 3), respectively (Fig. 2). The luminal release of 5-HT significantly increased (30.86 ± 3.83 ng/ml, n = 3) with increasing intraluminal pressure (40 cmH₂O) to a level 528.0 ± 39.8% (n = 3) above the basal level. This stimulatory effect of the intraluminal pressure on the luminal release of 5-HT was not altered by the vascular injection of TTX (471.0 ± 98.1%) (data not shown). The vascular release of 5-HT was not affected by the increasing intraluminal pressure (5.0 ± 0.6 ng/ml) (Fig. 2).

Fig. 2. Effect of high intraluminal pressure on 5-hydroxytryptamine (5-HT) release from the vascularly and luminally perfused rat ileum. We examined the effect of elevated intraluminal pressure on 5-HT release in ex vivo vascularly and luminally perfused rat ileum. The basal level of 5-HT release from the perfused ileum into the lumen and the portal circulation was 4.63 ± 0.37 ng/ml (n = 3) and 3.84 ± 0.44 ng/ml (n = 3), respectively (Fig. 2). The luminal release of 5-HT significantly increased (30.86 ± 3.83 ng/ml, n = 3) with increasing intraluminal pressure (40 cmH₂O) to a level 528.0 ± 39.8% (n = 3) above the basal level. This stimulatory effect of the intraluminal pressure on the luminal release of 5-HT was not altered by the vascular injection of TTX (471.0 ± 98.1%) (data not shown). The vascular release of 5-HT was not affected by the increasing intraluminal pressure (5.0 ± 0.6 ng/ml) (Fig. 2).

Fig. 3. Effect of 5-HT on FITC-LPS translocation. To clarify the role of 5-HT in the effect of high intraluminal pressure on FITC-LPS translocation, we administered 5-HT into the lumen as well as into the jugular vein. We subsequently measured the FITC-LPS level in the superior mesenteric vein (Fig. 3) and found that the basal level of FITC-LPS in the superior mesenteric vein (10.48 ± 0.14 ng/ml, n = 6) significantly increased (15.76 ± 0.47 ng/ml, 152.0 ± 8.54% of basal level, n = 6) with the intraluminal injection of 10 μmol·kg⁻¹·min⁻¹ of 5-HT (Fig. 3, A and C). However, the intraluminal administration of 1 and 0.1 μmol·kg⁻¹·min⁻¹ of 5-HT did not affect the basal level of FITC-LPS in the superior mesenteric vein (Fig. 3C), whereas it significantly increased from 10.48 ± 0.14 to 13.08 ± 0.54 ng/ml (133.7 ± 7.25% of basal level, n = 6) after the vascular injection of 0.1 μmol·kg⁻¹·min⁻¹ of 5-HT (Fig. 3, B and D). The vascular administration of 0.01 μmol·kg⁻¹·min⁻¹ of 5-HT did not affect the basal level of FITC-LPS detected in the superior mesenteric vein (Fig. 3D).
DISCUSSION

Our results suggest that the increase in the intraluminal pressure of the rat ileum stimulates the translocation of FITC-LPS from the intestinal lumen to the superior mesenteric vein and that this stimulatory effect is mediated by 5-HT, which is released from EC cells, via 5-HT₃ receptors. We prepared in vivo models of anesthetized rats to measure the rate of translocation, because blood supply and extrinsic nervous innervation in the intestinal segments remain intact in this model (7, 15). For the measurement of 5-HT release from the intestinal segment, we used ex vivo models of

Fig. 3. Effect of intraluminal and intravenous injection of 5-HT on FITC-LPS translocation. FITC-LPS level in the superior mesenteric vein significantly (*P < 0.05) increases by both luminal (A) and vascular (B) injection of 5-HT (10 and 0.1 μmol·kg⁻¹·min⁻¹, respectively) compared with period before stimulation (a). However, intraluminal injection of 1.0 and 0.1 μmol·kg⁻¹·min⁻¹ of 5-HT and intravenous injection of 0.01 μmol·kg⁻¹·min⁻¹ of 5-HT have no effects (C and D). Values represent means ± SD of 5-min samples from 6 animals. %Stimulation was calculated as: (mean of the FITC-LPS levels marked with asterisks/mean of the FITC-LPS levels in saline-injected controls) × 100. *P < 0.05 vs. saline-injected controls.

Fig. 4. Effect of 5-HT receptor antagonists on the 5-HT-induced FITC-LPS translocation. The FITC-LPS level, which was elevated by intraluminal (A and B) and intravenous (C and D) injection of 5-HT, significantly decreased after luminal (A and D) or vascular (B and C) injection of azasetron but not by injection of methysergide, ketanserin, nor SB-204070. Values represent means ± SD from 6 animals. #P < 0.05 vs. saline-injected controls.
translocation. This method seems reliable, because re-
the plasma levels of FITC as an indicator of LPS
injected FITC-LPS into the ileal lumen, and measured
ination have not been measured. In the present study, we
circulation after the stimulation of intestinal obstruc-
(15). However, endotoxin (LPS) levels in the portal
mesenteric lymph nodes (5), liver, spleen, or kidney

We injected 50
100
200
Control
Clamp
Clamp +
Luminal inj of
Azasetron
1 μg·kg⁻¹·min⁻¹
Vascular inj of
Azasetron
0.1 μg·kg⁻¹·min⁻¹

Fig. 5. Effect of 5-HT₃ receptor antagonists on FITC-LPS transloca-
tion induced by high intraluminal pressure. The stimulatory effect of
the high intraluminal pressure on FITC-LPS translocation is com-
pletely inhibited by both intraluminal or intravenous administration
of azasetron. Values represent means ± SD from 6 animals. #P <
0.05 vs. nontreated controls.

vascularly and luminally perfused ileum, because this
preparation is useful to compare the bidirectional re-
lease of 5-HT into the lumen and blood vessels (12).

Previous studies (5, 15) have demonstrated that in-
testinal obstruction causes bacterial translocation in
both humans and experimental animals. In these stud-
ies, bacterial translocation was evaluated by counting
the number of bacteria or radiolabeled E. coli in the
mesenteric lymph nodes (5), liver, spleen, or kidney
(15). However, endotoxin (LPS) levels in the portal
circulation after the stimulation of intestinal obstruct-
ion have not been measured. In the present study, we
injected FITC-LPS into the ileal lumen, and measured
the plasma levels of FITC as an indicator of LPS
translocation. This method seems reliable, because re-
cent fluorescence microscopic study has shown that
FITC-LPS is stable in the ileal lumen and is translo-
cated from the lumen to the lamina propria, whereas
FITC-vehicle does not cross the epithelial barrier (16).
We injected 50 μg/ml of FITC-LPS into the ileal lumen.
Because the mean LPS levels in the luminal content of
the terminal ileum of rats under normal conditions is
~1.8 μg/ml, as described previously (30), 50 μg/ml of
exogenously applied LPS seems to be a pathological
dose. However, this dose is comparable to other in vivo
studies, in which 50−400 μg/ml LPS was injected into
the intestinal lumen to examine the permeability of
endotoxin during noninflammatory stress, such as im-
mobilization, foot shock, or mesenteric ischemia (7, 27,
30). We detected a basal level of 10 ng/ml of FITC-LPS
in the superior mesenteric vein, which increased to
200% of the basal level in response to high intraluminal
pressure. These results indicate that 0.02% of the
total LPS applied into the lumen was tranlocated into
the portal circulation, even in the basal condition.

These findings were consistent with previous data that
demonstrated that ~0.02% of LPS, applied exog-
enoously into the ileal lumen of rats, was detected in the
mesenteric lymph nodes, mesentery, liver, and spleen
even in the basal state (30). However, the LPS level
detected in the superior mesenteric vein in our study
was markedly high compared with the plasma levels of
LPS (2−3 pg/ml) in normal animals. This discrepancy
may be the result of the induction of mucosal hyper-
permeability to endotoxins in response to exogenously
applied LPS (19, 24).

The effect of elevated intraluminal pressure on the
release of 5-HT was examined in an ex vivo perfusion of
the rat ileum, following previous studies that demon-
strated that release of 5-HT from EC cells is stimu-
ated by luminal pressure in the guinea pig small intestine
(3) and rat duodenum (10). When we increased the
pressure in the ileal lumen to 40 cmH₂O by clamping
the luminal effluent of the perfused ileum, the release
of 5-HT into the lumen markedly increased, whereas
the amount released into the portal circulation was not
altered. Because this stimulatory effect was not altered
by TTX administration, 5-HT release induced by high
intraluminal pressure must originate from EC cells,
not from 5-HT-containing enteric neurons (11).

To clarify the site of action of 5-HT released from EC
cells in response to high intraluminal pressure on the
epithelial permeability to FITC-LPS, we injected 5-HT
into the lumen and jugular vein to examine FITC-LPS
translocation. Because 5-HT released into the lumen
from EC cells may activate epithelial cells either from
the apical site or basolateral site of the epithelium, in
the latter case 5-HT may enter the lamina propria by
crossing the epithelium. In fact, a previous study (13)
has shown that intraluminally released 5-HT activates
nerve endings in the lamina propria. For intraluminal
administration, 10, 1.0, and 0.1 μmol·kg⁻¹·min⁻¹ of
5-HT (corresponding to 10⁻⁵, 10⁻⁶, and 10⁻⁷ M of
solution) were applied, because these concentrations of
5-HT influence intestinal motility when administered
intraluminally (13). For intravenous injection, 0.1 and
0.01 μmol·kg⁻¹·min⁻¹ of 5-HT were administered,
because these concentrations of 5-HT influence the intes-
tinal motility when administered intravenously (22).
The results showed that FITC-LPS levels in the su-
perior mesenteric vein were stimulated by both intralu-
minal and intravenous administration of 5-HT in
which vascularly administered 5-HT was more potent
than luminally administered 5-HT. In the ex vivo per-
fusion study, on the other hand, intraluminal release
but not vascular release of 5-HT was stimulated by
high intraluminal pressure. This discrepancy can be
explained by the fact that 5-HT released from EC cells
into the lumen may partially enter the lamina propria
and exert functions from the basolateral site of the
epithelium, which seems more potent than acting from
the luminal site.

To further identify the epithelial target site of lumi-
nally and intravenously administered 5-HT, 5-HT re-
ceptor antagonists combined with 5-HT were injected
via the same or different routes. The results showed
that the effect of luminal injection of 5-HT was inhibited by both luminal and vascular injection of 5-HT\textsubscript{3} receptor antagonist and that the effect of the vascular injection of 5-HT was inhibited by both luminal and vascular injection of 5-HT\textsubscript{3} receptor antagonist. The administration of 5-HT\textsubscript{1}, 5-HT\textsubscript{2}, and 5-HT\textsubscript{4} receptor antagonists did not affect these results. These results suggest that 5-HT, both intraluminally and intravenously administered, may affect the epithelial cells and increase the permeability via 5-HT\textsubscript{3} receptor, which is possibly localized in the epithelium.

The pathway of endotoxin translocation across the epithelium has been widely investigated in cultured epithelial cells (2, 17), Ussing chamber preparations (25), and Thiry-Vella loops of intestine (29). Proposed pathways include a transcellular route (1, 9) and a paracellular route (17, 25). In the transcellular route, endotoxin is taken by the endocytosis of the epithelial cell and transported to the basolateral side of the epithelium (2). In the paracellular route, the tight junction proteins, such as occludin, ZO-1, and ZO-2, may be involved in the regulation of epithelial permeability (1). 5-HT, both exogenously applied and endogenously released from EC cells, may influence the epithelial cells from an apical and/or basolateral site and stimulate the transcellular route, or may influence the intercellular tight junctions and stimulate the paracellular route, via 5-HT\textsubscript{3} receptors.

Our results demonstrated that FITC-LPS translocation induced by high intraluminal pressure is inhibited by 5-HT\textsubscript{3} receptor antagonist administered both intraluminally and intravenously. However, the localization of the 5-HT\textsubscript{3} receptor on either the apical or basolateral membrane of the epithelial cells has not been demonstrated. A few previous studies demonstrated that 5-HT\textsubscript{3} receptor is localized to the luminal site of the epithelium in the developing intestine (6) and on a small number of endocrine cells in the epithelium (14). Although identification of 5-HT receptors on apical and/or basolateral membrane of the epithelial cells awaits further investigation, 5-HT released from EC cells may function as a critical factor to stimulate the permeability of endotoxin through the intestinal epithelium.

In conclusion, our study demonstrated that the translocation of FITC-LPS injected into the rat ileum and the superior mesenteric vein was stimulated by the increase of intraluminal pressure, and this stimulatory effect might be mediated by 5-HT released from EC cells via the 5-HT\textsubscript{3} receptor possibly located on the epithelium.

REFERENCES


