Localization of the sulfate/anion exchanger in the rat liver

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SAT-1. SAT-1 may be responsible for the uptake of inorganic sulfate from the blood into hepatocytes to enable sulfation reactions. In oocytes was enhanced in the presence of bicarbonate, indicating sulfate/bicarbonate exchange. Estrone sulfate was not transported by sat-1. Sat-1 mRNA was found in hepatocytes and endothelial cells. Sat-1 protein was localized in sinusoidal membranes and along the borders of hepatocytes. The canalicular region and bile capillaries were not stained. Sulfate uptake was only slightly affected by sulfamyl diuretics or organo sulfates. Sulfate efflux from sat-1-expressing cells was not a pH gradient (pH 8.0 inside and pH 6.0 outside) stimulated sulfate uptake (22).

Sulfation of all naturally occurring sulfated compounds depends on the availability of intracellular inorganic sulfate. Inorganic sulfate, which accounts for >90% of the total body sulfate, is the precursor of 3'-phosphoadenosine 5'-phosphosulfate (PAPS), the activated form of sulfate and a substrate for sulfotransferases (7). Because sulfate is a divalent hydrophilic anion, it cannot pass freely through the phospholipid bilayer of plasma membranes and is therefore dependent on transport systems to allow its movement into or out of cells.

Na⁺-dependent as well as Na⁺-independent sulfate uptake was measured in the isolated perfused rat liver (3) and isolated hepatocytes (34). Sulfate uptake in hepatocytes was further characterized in rat liver sinusoidal (8) and canalicular (22) membrane vesicles. A transmembrane pH difference stimulated sulfate uptake in sinusoidal membrane vesicles, suggesting sulfate/hydroxyl ion exchange (8). Cis-inhibition and trans-stimulation of sulfate uptake were observed with sulfate, thiosulfate, oxalate, and succinate but not with chloride, bicarbonate, monocarboxylates, or glutamates and aspartate (8). In contrast, in rat canalicular membrane vesicles, a bicarbonate (50 mM bicarbonate inside and 5 mM bicarbonate outside) but not a pH gradient (pH 8.0 inside and pH 6.0 outside) stimulated sulfate uptake (22). Cis-inhibition of bicarbonate-stimulated sulfate uptake was observed with sulfate, thiosulfate, and oxalate but not with chloride, nitrate, phosphate, monocarboxylates, or glutathione (22). The sinusoidal transport system exhibited a low affinity for sulfate [Michaelis-Menten constant (Km) ~16 mM] (8), whereas the canalicular system showed a high affinity for sulfate (Km ~0.3 mM) (22).

In recent years, a Na⁺-dependent as well as a Na⁺-independent sulfate transporter have been cloned from various tissues and designated as NaSi-1 (19) and sat-1 (2), respectively. Whereas NaSi-1, the Na⁺-dependent sulfate transporter, was not detected in the liver (19), during the course of our study, a rat Na⁺-dependent sulfate transporter (rNaS2) was cloned (4). rNaS2 is expressed in the liver (4); however, BLASTing of its mRNA or amino acid sequences showed no sequence homology with sat-1. Sat-1 has been cloned from rats, humans, and mice (2, 18, 31). Functional studies in Xenopus laevis oocytes (2, 11, 20, 32, 35) and Sf9 insect cells (11) have suggested that sat-1 translocates sulfate and oxalate. Sulfate uptake was inhibited by selenate, thiosulfate, and the stilbene derivative DIDS. Sat-1 transcripts have been found in the liver and kidneys (2, 32), but localization of sat-1 has only been studied in the kidney (11).

In the liver, the localization of sat-1 is not known. The liver contributes to the biotransformation of drugs via sulfation; therefore, it is important to identify sulfate transport systems either for import of sulfate or export of sulfated compounds. NaSi-1 has not been detected in the liver (19), and rNaS2 has shown limited substrate specificity (4). In addition, the organic anion-transporting polypeptides present in the sinusoidal membrane of hepatocytes do not handle anorganic sulfate (10) but translocate estrone sulfate (ES) (17). Canaliculur excretion of organo sulfates is facilitated by multidrug resistance-associated protein 2 (MRP2) (12).

Here, we report on the localization of sat-1 in sinusoids as well as in cell-cell borders of hepatocytes and show that sat-1

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can work as a sulfate/bicarbonate exchanger. Testing of compounds reported to affect sulfate transport in the intact kidney (33) and sinusoidal and canalicul membrane vesicles (8, 22) revealed a low affinity for bulky sulfated compounds.

**MATERIALS AND METHODS**

**In situ hybridization.** This was performed in formalin-fixed paraffin-embedded tissue sections of male rat livers according to our standard protocol (16). Specific localization of sat-1 RNA was obtained by incubation with the digoxigenin-labeled antisense probe, whereas negative controls were performed by incubating sections with a digoxigenin-labeled RNA sense probe substituted for the antisense probe simultaneously with the antisense incubation and by omitting the RNA probes in each experiment. Digoxigenin-labeled RNA probes were yielded by in vitro transcription using the RNA polymerases T7 (sense) and T3 (antisense) starting from a sequence specifically coding for rat sat-1 mRNA (bases 2011–2358, Accession No. L23413). This sequence was amplified from rat liver mRNA using specific primers for rat sat-1 flanked by the T7 and T3 promoter sequences for the forward (5'-TCTAGAACGCTCAGATATC-3') and reverse primers (5'-GCAA-TTAACCTCTCAGATGGAAGCACCGTTC-3'), respectively. These amplified a product of 392 bp bridging the 348 bp of the sat-1 sequence. The 392-bp product amplified by these primers was sequenced and shown to correspond to sat-1.

**Sat-1 expression in rat liver cells in vitro.** Hepatocytes, hepatic stellate cells, and endothelial cells from the rat liver were prepared and seeded as previously described. Freshly isolated cells were investigated for contamination with other liver cell populations using the appropriate markers (13–15, 27–29). Purity reached 99% for hepatic stellate cells, and 85% for endothelial cells, respectively. After RNA extraction from different cultures with the TRIzol reagent (Invitrogen; Karlsruhe, Germany), 2 µg of each RNA sample were reverse transcribed using oligo(dT) primer (Invitrogen) and mouse Moloney leukemia virus reverse transcriptase (Promega). cDNA was amplified by PCR using the forward primer rsat-1_355For (5’-TCCCAGATTAGCCAAAGAGCC-3’ (Invitrogen) and mouse Moloney leukemia virus reverse transcriptase using specific primers for rat sat-1 flanked by the T7 and T3 promoter sequences for the forward (5’-TCTAGAACGCTCAGATATC-3’) and the reverse primer rsat-1_869Rev (5’-GTGTGGCTGTGTTGTTGAG-3’) (H11032), which amplified a band of 514 bp (11). Semiquantitative PCR was performed by normalization to the amount of cDNA of GAPDH.

**Immunohistochemical detection of sat-1.** Formalin-fixed frozen sections of male rat livers were stained using a primary mouse-anti-sat-1 antibody (kindly provided by L. P. Karniski, University of Iowa), which was previously used to localize sat-1 in rat kidney proximal tubules (11). Before antibody incubation, intracellular peroxidase was blocked by methanol-H₂O₂, and the sections were predigested with pepsin. Immunoreactions were revealed using peroxidase-labeled anti-mouse antibodies (Dako: Hamburg, Germany) preadsorbed with normal rat serum (Dako) and developed with diaminobenzidine. Sections were counterstained, dehydrated, and coverslipped. Nonspecific binding of the secondary antibody was in some sections blocked using saturated milk powder, which preceded the incubation with the primary antibody instead of the rat serum treatment. In negative controls, the primary antibody was omitted.

**Functional analysis of sat-1.** Sat-1 cDNA from the rat liver was used as a template for cRNA synthesis. Plasmids were linearized with Ncol, and in vitro cRNA transcription was performed using the T7 mMessage mMachine kit (Ambion; Austin, TX). The resulting cRNA was suspended in purified, RNase-free water to a final concentration of 1 µg/µl. Stage V and VI oocytes from X. laevis (Nasco; Fort Atkinson, WI) were separated by treatment with collagenase (type II, Biochrom; Berlin, Germany) and maintained at 16–18°C in oocyte Ringer (ORi) solution. The following reagents (Sigma; Taufkirchen, Germany) were used for the ORi solution (in mM): 110 NaCl, 3 KCl, 2 CaCl₂, and 5 HEPES-Tris (pH 7.5). One day after removal from the frog, oocytes were injected either with 30 nl cRNA coding for sat-1 or an equivalent amount of water and maintained at 16–18°C in ORi solution supplemented with 50 µM gentamycin and 2.5 mM pyruvate. After 3–4 days with medium changes, oocytes were used for tracer uptake studies.

Uptake of [³⁵S]sulfate (H₂SO₄, 1,200 Ci/mM, Hartmann S-RA-1; Braunschweig, Germany) in sat-1-expressing oocytes was assayed at room temperature. For kinetic experiments, oocytes were incubated for 1 h in ORi solution with the addition of 0.02 µM [³⁵S]SO₄⁻² (20 µCi) and increasing sodium sulfate concentrations up to 1 mM. Cis-inhibition of sulfate uptake was determined by the simultaneous application of 0.02 µM [³⁵S]SO₄⁻² in 0.5 mM NaN₂SO₄ and varying concentrations of diuretics, ES, and 2,3-dimercaptopropane sulfonate (DMPS) (Sigma) for 1 h. Uptake of [³²P]DHEA (46 Ci/mM, Perkin-Elmer NET-203; Rodgau, Germany) and [³²P]dihydroepiandrosterone (DHEA) sulfate (74 Ci/mM, Perkin-Elmer NET-860) was assayed by incubation of the oocytes for 1 h in ORi solution containing 10 nM [³²P]DHEA (0.46 µCi) in either 0.1 or 0.5 µM ES (Sigma) or 20 mM [³²P]DHEA sulfate (1.48 µCi) in either 500 mM or 500 µM DHEA sulfate (Sigma). After the incubation in the respective solutions, the radioactivity was aspirated, and oocytes were washed twice in ice-cold ORi solution. Oocytes were dissolved by gentle shaking for 2 h in 100 µl of 1 N NaOH and neutralized with 100 µl of 1 N HCl, and their ³²P or ³⁵S contents were determined by liquid scintillation counting (Tricarb 2900TR, Perkin-Elmer). In trans-stimulation experiments, oocytes were injected with 50 nl of 20 mM ES before [³⁵S]sulfate uptake was measured. Water-injected oocytes were treated in a similar manner to serve as controls. All uptake experiments were performed at least in duplicate with six to eight oocytes for each experimental condition.

For the sulfate efflux experiments, sat-1-expressing and water-injected oocytes were injected with 46 nl of [³⁵S]sulfate, which correlates to a final concentration of 0.92 pmol/oocyte. Immediately after the injection, single oocytes were placed in a 2-ml cup with ORi solution to remove contamination of [³⁵S]sulfate from the oocyte surface, aspirated, and replaced in a 2-ml cup to follow [³⁵S]sulfate efflux. Twenty-eight minutes later, oocytes were aspirated and washed twice in ice-cold ORi solution to stop [³⁵S]sulfate efflux. [³⁵S]sulfate efflux was assayed in ORi solution in which sodium chloride (110 mM) was removed and replaced by sodium methane sulfonate (110 mM) and in ORi solution in which part of the sodium chloride was replaced by sodium bicarbonate (30 mM).

**Statistics.** Substrate-dependent sulfate uptakes were fitted to the following Michaelis-Menten equation using SigmaPlot software (Systat Software; Point Richmond, CA): \( V = V_{\text{max}} \times \frac{[S]}{(K_m + [S])} \), where \( V \) is the substrate-dependent sulfate uptake, \( V_{\text{max}} \) is the maximal uptake measured at saturating sulfate concentrations, \( K_m \) is the substrate concentration at half maximal uptake, and \( [S] \) is the sulfate concentration. An unpaired Student’s \( t \)-test was used to show statistically significant differences in sulfate uptake in the absence (control) and presence of diuretics. Statistical significance was set at \( P < 0.05 \).

**RESULTS**

In situ hybridization showed that after incubation of the sections with the antisense probes, a specific signal in the liver parenchyme was clearly detected in hepatocytes and in several endothelial cells (Fig. 1, A and B). Incubation with the sense probes (Fig. 1C) yielded no signals.

RT-PCR studies in hepatocytes and endothelial cells as well as in stellate cells were performed in vitro. Sat-1 was expressed in hepatocytes (Fig. 2A), where the level of expression did not change during the time in culture. In freshly isolated endothelial cells, sat-1 transcripts were also detected (Fig. 2B). In stellate cells, clear expression for sat-1 was seen on days 6 and 7 of culture but not on days 2 or 4 (Fig. 2C).
but also sulfated steroids, the uptake of \([3H]\)ES and \([3H]\)DHEA sulfate uptake significantly (Fig. 4A). Diuretics (Fig. 4B). Among the diuretics (Fig. 4C), only acetazolamide was able to inhibit sulfate uptake significantly \((P < 0.05)\).

To test whether sat-1 is not only able to translocate sulfate but also sulfated steroids, the uptake of \([3H]\)ES and \([3H]\)DHEA sulfate was investigated. In these experiments, oocytes from the same frog were divided randomly into three groups. One group was incubated for 1 h with ORI solution containing 0.5 mM \([35S]\)sulfate, whereas the two other groups were incubated for 1 h in ORI solution containing 0.1 or 0.5 \(\mu\)M \([3H]\)ES. In three batches of oocytes, sulfate uptake was 189.9 ± 43.9 pmol·h\(^{-1}\)·oocyte\(^{-1}\), indicating successfully expressed sat-1. ES uptake into sat-1-expressing oocytes was 0.8 ± 0.1 and 9.1 ± 0.4 pmol·h\(^{-1}\)·oocyte\(^{-1}\) at 0.1 and 0.5 \(\mu\)M ES, respectively (Fig. 5A). ES uptake into water-injected oocytes was 0.8 ± 0.1 and 8.7 ± 0.8 pmol·h\(^{-1}\)·oocyte\(^{-1}\), respectively, demonstrating the absence of sat-1-mediated ES uptake. Higher ES concentrations could not be tested because of nonspecific binding of ES to the oocytes. ES undergoes enterohepatic circulation and has to exit from the hepatocytes. To test for ES/sulfate exchange, sat-1-expressing oocytes were injected with ES before they were incubated in 0.5 mM sulfate for the \([35S]\)sulfate uptake to be measured. However, \([35S]\)sulfate uptake by sat-1 was not enhanced by preloading the oocytes with ES (Fig. 5B): it was 105.5 ± 5.7 pmol·h\(^{-1}\)·oocyte\(^{-1}\) without injection of ES and 116.3 ± 13.8 pmol·h\(^{-1}\)·oocyte\(^{-1}\) with injection of ES. Sulfate uptake in water-injected controls was 1.7 ± 0.3 and 8.0 ± 2.3 pmol·h\(^{-1}\)·oocyte\(^{-1}\). DHEA sulfate uptake in sat-1-expressing oocytes was 112.1 ± 15.3 and 599.1 ± 178.6 pmol·h\(^{-1}\)·oocyte\(^{-1}\).

In all liver sections treated with the primary anti-sat-1 antibody, staining was regularly seen along the sinusoidal lining (Fig. 3A). Cell-cell borders between hepatocytes (Fig. 3, A and C) were also clearly stained with the exception of the central region, which corresponds to the bile canaliculi (Fig. 3, A and C). In several corners between hepatocytes, at very high magnification (Fig. 3D), it was possible to distinguish a separation of the staining of the hepatocytic membrane (Fig. 3D) from the staining of the sinusoidal lining (Fig. 3D). In negative controls (Fig. 3B), no such staining was seen.

Incubation of 149 oocytes expressing sat-1 and 115 water-injected control oocytes from 18 different frogs in 0.5 mM \([35S]\)sulfate for 1 h led to an uptake of sulfate of 121.7 ± 32.6 and 4.4 ± 1.1 pmol·h\(^{-1}\)·oocyte\(^{-1}\), respectively (Fig. 4A). To evaluate the substrate specificity of sat-1, certain drugs were tested in cis-inhibition experiments for their ability to inhibit \([35S]\)sulfate uptake. For a better comparison of the data, sulfate uptake in the presence of these drugs was calculated as a percentage of sulfate uptake in the absence of these drugs. DMPS is used to reduce the content of mercury in the liver (36). DMPS at concentrations of 0.25 and 1 mM enhanced rather than inhibited \([35S]\)sulfate uptake (Fig. 4B). Among the diuretics (Fig. 4C), only acetazolamide was able to inhibit sulfate uptake significantly \((P < 0.05)\).

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fmol·h⁻¹·oocyte⁻¹ at 500 nM and 500 μM DHEA sulfate, respectively. In water-injected oocytes, it was 101.8 ± 17.1 and 553.1 ± 26.1 fmol·h⁻¹·oocyte⁻¹, demonstrating the absence of sat-1-mediated DHEA sulfate uptake (Fig. 5C). In addition, DHEA sulfate was not able to inhibit [³⁵S]sulfate uptake up to concentrations of 500 μM (data not shown).

For sat-1-mediated sulfate effects, oocytes were microinjected with [³⁵S]sulfate. The initial [³⁵S]sulfate concentration averaged 0.92 pmol/oocyte in sat-1-expressing as well as water-injected oocytes. In these oocytes, the amount of [³⁵S]sulfate did not leak out of the oocytes (15 oocytes from 3 donors). In sat-1-expressing oocytes, [³⁵S]sulfate decreased from 0.92 to 0.52 ± 0.08 pmol/oocyte within 30 min (15 oocytes from 3 donors; Fig. 6A). [³⁵S]sulfate efflux was slightly enhanced by chloride (Fig. 6A) but was strongly accelerated in the presence of 30 mM bicarbonate (Fig. 6A). Under this condition, nearly all of the injected [³⁵S]sulfate left the sat-1-expressing oocytes by sat-1 within 30 min, whereas [³⁵S]sulfate stayed constant in water-injected controls. In cis-inhibition experiments, [³⁵S]sulfate uptake was only marginally affected by bicarbonate (Fig. 6B): 164.4 ± 19.8 pmol·h⁻¹·oocyte⁻¹ in the absence of

Fig. 3. Localization of sat-1 in the liver of male rats. The presence of sat-1 was demonstrated by immunohistochemistry using anti-sat-1 antibodies. A: positive staining was seen along the sinusoids (closed arrows) and cell borders between hepatocytes (open arrows). Arrowheads indicate regions between hepatocytes that were not stained. Sat-1-positive cells did not form a particular pattern and were scattered throughout the liver. B: specific staining was abolished by substitution of the anti-sat-1 antibody with 1% bovine serum albumin in Tris-buffered saline. C: higher magnification of cell-cell borders between hepatocytes that were stained by the anti-sat-1 antibody. Open arrows indicate staining of the cell borders; the central region (arrowhead) was not stained. D: higher magnification of a site in which staining of the hepatic cell membrane (open arrow) can be distinguished from the staining of the sinusoidal wall (closed arrow). Scale bars = 5 μm.

Fig. 4. Influence of 2,3-dimercapto-1-propane sulfonate (DMPS) and diuretics on sulfate uptake. A: 1-h [³⁵S]sulfate uptake was measured in sat-1-expressing oocytes (sat-1 oocytes; 149 oocytes) and water-injected oocytes (H₂O oocytes; 115 oocytes from 18 frogs). Sat-1-expressing oocytes were incubated in oocyte Ringer (ORi) solution containing 0.5 mM [³⁵S]sulfate (A–C) and 1 mM DMPS (B) and diuretics in the concentrations indicated (C). All data were corrected for sulfate uptake in water-injected controls under the same experimental conditions. Data are expressed as percentages of sulfate uptake in the absence of the drugs (control) and are means ± SE from at least 2 frogs and >7 oocytes for each experimental condition. *Statistically significant at P < 0.05.
Fig. 5. Comparison of sulfate uptake in the presence and absence of estrone sulfate (ES). A: 1-h uptake studies of 0.1 and 0.5 μM [3H]ES were compared. Data are means ± SE from 3 independent determinations with >6 oocytes for each experimental condition. B: [35S]sulfate uptake was determined after oocytes were injected with either 50 nl of 20 mM ES (shaded bars) or an equivalent amount of water (solid bars). Data are means ± SE of at least 8 oocytes from 3 donors for every condition. C: 1-h uptake studies of 500 nM and 500 μM [3H]dehydroepiandrosterone (DHEA) sulfate. Data are means ± SE from 3 independent determinations with >9 oocytes for each experimental condition.

Fig. 6. Dependence of sat-1 transport function on bicarbonate. A: sat-1-expressing and water-injected control oocytes were injected with 46 nl [35S]sulfate (0.92 pmol/oocyte), and the efflux of sulfate was followed for 30 min in the presence (solid bars) and absence of chloride (shaded bars) and in a solution containing 30 mM bicarbonate in chloride-containing ORi solution (open bars). Chloride was substituted by methane sulfonate and in the bicarbonate experiments, 30 mM bicarbonate was added at the expense of chloride. Experiments were performed with 5 oocytes from 3 donors. B: sat-1-expressing and water-injected control oocytes were incubated for 1 h in ORi solution containing 0.5 mM total sulfate in the absence and presence of 30 mM bicarbonate. Data are means ± SE of at least 8 oocytes for each condition from 4 donors. C: sat-1-expressing oocytes and water-injected controls were preincubated for 4 h in either the absence or presence of 30 mM bicarbonate. Bicarbonate was added at the expense of chloride. Data are means ± SE of at least 8 oocytes from 2 donors.
bicarbonate and 140.2 ± 36.5 pmol·h⁻¹·oocyte⁻¹ in the presence of bicarbonate. The values for water-injected oocytes were 3.9 ± 0.7 and 2.8 ± 0.6 pmol·h⁻¹·oocyte⁻¹, respectively. To demonstrate the direction of transport, oocytes were incubated for 4 h in ORI solution containing 30 mM bicarbonate at the expense of chloride. In two experiments, [³⁵S]sulfate uptake was enhanced by a factor of 1.59 ± 0.12 in oocytes preincubated in bicarbonate-containing solutions (Fig. 6C).

DISCUSSION

Despite the cloning of sat-1 as a hepatic sulfate transporter, neither its physiological role nor its localization within the liver has been clarified to date. Sat-1 was identified as a Na⁺-independent sulfate transporter by expression cloning from the rat liver (2), and its characteristics matched in part those identified for the sulfate transport systems from either the canalicular (22) or the sinusoidal (8) membrane of hepatocytes. In sat-1-expressing oocytes, the $K_m$ for sulfate was found to be 0.14 mM (2), which is close to that reported for rat canalicular membrane vesicles (0.3 mM) (22). Because canalicular and sinusoidal membrane vesicles may each contain several sulfate transporters, no safe conclusion from the $K_m$ to the localization of sat-1 can be drawn.

In the present work, we clearly showed that hepatocytes and endothelial cells express transcripts of sat-1. In contrast, stellate cells showed clear sat-1 expression only under conditions that were far from the physiological state. We also showed that sat-1 is localized along the sinusoidal borders and in the borders between hepatocytes. Unexpectedly, the canalicular region was free of staining. Therefore, we conclude that sat-1 is not located in the bile canaliculi. In contrast, our results identify sat-1 as a sulfate transporter present in the sinusoidal membrane of hepatocytes in the interhepatocytic borders and in endothelial cells.

Sulfate serves to form PAPS, a sulfate donor important in detoxification processes (7). However, in the liver, sinusoids are provided with a perisinusoidal matrix containing sulfated proteoglycans (6, 25, 26). Proteoglycans participate in many cellular processes such as migration, adhesion, proliferation, and regulation of growth factors (9). The presence of sat-1 in the sinusoidal domain of hepatocytes and in other cell types located in the proximity of the sinusoids such as endothelial cells, as shown in the present work, strongly suggests that sat-1-dependent sulfate transport may be involved in the syntheses of sulfated proteoglycans of the perisinusoidal matrix. In the present work, we also showed that, in stellate cells, sat-1 expression increases with the length of time of culture. This might be suggestive of a general tendency of stellate cells to upregulate sat-1 expression under conditions that are far from the physiological state. In this context, it is interesting to note that nonphysiological conditions in which stellate cells play a key role occur, for example, in chronic liver injury. In the course of this process, stellate cells can be activated by stimuli derived from damaged hepatocytes, endothelial cells, and Kupffer cells. Stellate cell activation results in liver fibrosis with changes in extracellular matrix composition accompanied by production of great amounts of matrix also containing sulfated proteoglycans (1, 6, 26, 30). Whether an upregulation of sat-1 occurs during activation of stellate cells in liver injury in vivo and, therefore, may account for an increased sulfate uptake in this cell type is not yet known.

Liver sulfation capability is not only aimed at synthesising sulfated proteoglycans. Sulfation enhances the water solubility of steroids and drugs and their distribution within the plasma. Sulfation also facilitates the excretion of lipophilic compounds by the kidneys, thereby contributing to detoxification. In this context, the question arises as to whether sat-1 contributes to excretion of sulfated compounds and, therefore, may participate in their excretion. The results of our experiments show that sat-1 does not translocate ES and DHEA sulfate. Therefore, it is conceivable that sulfate steroids are not excreted from the liver via a sat-1-mediated transport. The excretion of these sulfated steroids is rather mediated by other systems localized in the sinusoidal membrane (5, 33a) or, in the bile, by MRP2 (12). With the exception of acetazolamide, millimolar concentrations of diuretics were not able to reduce sulfate uptake mediated by sat-1. The preference of sat-1 for inorganic sulfate and the sinusoidal localization of the transporter suggest a role for sat-1 in the uptake of inorganic sulfate from the circulation rather than a function in the excretion of sulfated compounds from the liver. This feature enables sat-1 to continuously support hepatocytes and endothelial cells providing sufficient sulfate for sulfation.

We were also able to demonstrate sulfate efflux from oocytes that was not significantly changed when chloride was replaced by methane sulfonate but was strongly accelerated in the presence of bicarbonate. This indicates sulfate/bicarbonate rather than sulfate/chloride exchange by sat-1. In cis-inhibition experiments, [³⁵S]sulfate uptake was only marginally reduced in the presence of bicarbonate, suggesting that the affinities for bicarbonate at the extracellular site may be less strong than at the intracellular site of sat-1.

In conclusion, sat-1 is a sulfate/bicarbonate exchanger located in the sinusoidal membrane of hepatocytes as well as at their interhepatocytic borders and in endothelial cells.

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FUNCTIONAL CHARACTERIZATION AND LOCALIZATION OF SAT-1 IN THE LIVER


