HIV protease inhibitors activate the unfolded protein response and disrupt lipid metabolism in primary hepatocytes

Huiping Zhou,1 Emily C. Gurley,1 Sirikalaya Jarujaron,1 Hong Ding,1 Youwen Fang,1 Zhumei Xu,2 William M. Pandak, Jr.,3 and Phillip B. Hylemon1

1Departments of Microbiology and Immunology, 2Biochemistry, 3Internal Medicine, and McGuire Veterans Affairs Medical Center, Virginia Commonwealth University, Richmond, Virginia

Submitted 1 May 2006; accepted in final form 17 July 2006

HIV protease inhibitors activate the unfolded protein response (UPR), induce apoptosis, and promote foam cell formation in macrophages. In the present study, we examined the effects of three different HIV PIs, including amprenavir, atazanavir, and ritonavir, on the UPR activation and the expression of key genes involved in lipid metabolism in primary rodent hepatocytes. Both atazanavir and ritonavir activated the UPR, induced apoptosis, and increased in lipid metabolism in primary rodent hepatocytes. Our previous studies demonstrated that HIV PIs differ in their ability to increase the levels of transcriptionally active sterol regulatory element-binding proteins (SREBPs), activate the unfolded protein response (UPR), induce apoptosis, and promote foam cell formation in macrophages. In the present study, we examined the effects of three HIV PIs, including amprenavir, atazanavir, and ritonavir, on the UPR activation and the expression of key genes involved in lipid metabolism in primary rodent hepatocytes. Both atazanavir and ritonavir activated the UPR, induced apoptosis, and increased in lipid metabolism (22).

Although the mechanisms underlying HIV PI-associated dyslipidemia are not fully understood, an increasing body of evidence suggests that multiple cellular mechanisms may be involved and individual HIV PI may have different effects on lipid metabolism (22).

The liver plays a central role in maintaining lipid homeostasis in the body. Under normal physiological conditions, lipid input is equal to lipid output from the body (20). Disruption of either the input or output pathways will result in dysregulation of lipid metabolism. Previous studies have shown that indinavir alters sterol and fatty acid metabolism in primary rat hepatocytes by increasing levels of activated sterol regulatory element-binding proteins (SREBPs) and decreasing cholesterol 7α-hydroxylase (CYP7A1) mRNA levels (32, 40). In addition, several studies have shown that perturbation of proteasome activities by HIV PIs may also contribute to dyslipidemia (27, 29, 35). Furthermore, den Boer et al. (12) recently reported that ritonavir specifically inhibited postheparin lipoprotein lipase activity, which may decrease plasma triglyceride clearance and ultimately results in hyperlipidemia. Several clinical studies suggest that amprenavir and atazanavir are less likely than other HIV PIs to induce dyslipidemia (1, 17). However, the clinical significance of these observations in terms of decreased cardiovascular risk is still unknown because of limited clinical information. In addition, similar to other HIV PIs, atazanavir also can cause hyperglycemia, insulin resistance, and lipodystrophy, which are independent risk factors for cardiovascular disease (4, 25, 26, 39).

HIV PI-induced endoplasmic reticulum (ER) stress and subsequent activation of the unfolded protein response (UPR) may represent an important cell signaling mechanism of HIV PI-induced metabolic syndromes (28, 44). We have demonstrated that HIV PI ritonavir increases the accumulation of free cholesterol, depletes the ER calcium stores, activates the unfolded protein response (UPR), induces apoptosis, and promotes foam cell formation in macrophages (44). In addition, our recent studies show that all the HIV PIs, except amprenavir, activate the UPR and induce apoptosis to different extents in macrophages (Zhou H, Sirikalaya S, Gurley EC, Pandak WM, and Hylemon PB, unpublished data). However, the effect of individual HIV PI has on activation of the UPR, induction of cell apoptosis, and disruption of the lipid metabolism in primary hepatocytes has not been fully explored.

The objective of the present study was to compare the effects of three different HIV PIs on activation of the UPR, as well as
on lipid metabolism in primary rodent hepatocytes. The results show that both atazanavir and ritonavir activated the UPR and induced apoptosis in hepatocytes, but amprenavir had no significant effect on UPR activation or lipid metabolism. These three HIV PIs also showed different effects on the expression of key genes involved in lipid metabolism, including CYP7A1, 3-hydroxy-3-methyl-glutaryl-coenzyme A reductase (HMG-CoA-R), and low-density lipoprotein receptor (LDL-R). The current results may help to provide a better understanding of the cellular mechanisms of lipid dysregulation induced by different HIV PIs and also provide useful information for the development of new therapeutic strategies to control HIV PI-associated clinical problems.

MATERIALS AND METHODS

Materials. Antibodies against C/EBP homologous protein (CHOP), activating transcription factor-4 (ATF-4), X-box-binding protein-1 (XBP-1), lamin B, SREBP-1, and SREBP-2 and horseradish peroxidase (HRP)-conjugated donkey anti-goat IgG were from Santa Cruz Biotechnology (Santa Cruz, CA). Mouse monoclonal antibody against β-actin was from Calbiochem (San Diego, CA). Polyclonal antibody against mouse CYP7A1 was a generous gift from Dr. David W. Russell (The University of Texas, Southwestern Medical Center). Bio-Rad protein assay reagent, Criterion XT Precast Gel, HRP-conjugated goat anti-rabbit IgG and Precision Plus Protein Kaleidoscope Standards were obtained from Bio-Rad (Hercules, CA). Amprenavir, atazanavir, and ritonavir were generous gifts from GlaxoSmithKline (Barnard Castle, Durham, UK), Bristol-Meyers-Squibb (New Brunswick, NJ), and Abbott Laboratories (Abbott Park, IL), respectively. Biomax MS films were obtained from Eastman Kodak (Rochester, NY). RNAqueous total RNA isolation kit and MAXscript T7 and ribonuclease protection assay (RPA) II kits were from Ambion (Austin, TX). High-capacity cDNA archive kit and gene expression kits for rat LDL-R, CYP7A1, and HMG-CoA-R were from Applied Biosystems (Foster City, CA). CellTiter 96AQeuous One Solution Reagent was from Promega (Madison, WI). All other chemical reagents were from Sigma (St. Louis, MO).

Isolation and culture of primary hepatocytes. Primary hepatocytes were isolated from adult male Sprague-Dawley rats (250–300 g) or C57BL/6 mice (20–25 g) using the collagenase-perfusion technique of Bissell and Guzelian (2). Trypan blue exclusion was used to determine cell viability (>90%) before plating monolayers on collagen-coated plates (60 mm). Unless otherwise indicated, cells were cultured in serum-free Williams’ E medium containing dexamethasone (0.1 μM), insulin (100 nM), penicillin (100 units/ml), and thyroxine (1 μM). Cells were incubated from 12 to 24 h in 5% CO2 environment at 37°C before additions were made to culture medium. Amprenavir, atazanavir, and ritonavir were dissolved in DMSO. HIV PIs were added directly to culture medium (final concentrations 5 to 40 μM) and incubated for 0.5 to 24 h.

High-performance liquid chromatography assay of the metabolism of HIV PIs. A HPLC system (System Gold, Beckman Coulter, Montreal, QC, Canada) and a Beckman C18 reverse-phase column (4.6 mm × 25 cm) were used in these experiments. Chromatographic separation was obtained using an analytical C18 column (5 μm) at room temperature under isotropic conditions. The mobile phase was acetonitrile: 20 mM sodium dihydrogenphosphate, pH 6 (60:40 vol/vol) and 0.025% triethylamine. The mobile phase was delivered at 1 ml/min. The HIV PI peaks were detected spectrophotometrically at 210 nm.

Rat primary hepatocytes were treated with HIV PIs (30 μM) for various time periods (0 to 24 h). The culture medium at each time point was collected and centrifuged at 14,000 g for 1 min. The HIV PIs in media were extracted using solid phase C-18 cartridges (Water), which were conditioned with 1 ml of methanol followed by 1 ml of water. An aliquot of culture media (250 μl) and an internal standard, verapamil (final concentration 5 μM), were applied onto the cartridge. The cartridges were further washed with 1 ml water, followed by 1 ml methanol-water (30:70, vol/vol) after incubation at room temperature for 30 min. HIV PIs were eluted with 1 ml methanol. The eluent was evaporated under a nitrogen gas stream, and the residue was reconstituted in 250 μl of mobile phase. A 40-μl aliquot was injected onto the HPLC column (33).

A standard curve of each drug was constructed using weighted linear regression of peak area ratio values of the calibration standards. The percentage of drug recovery after the solid-phase extraction was determined by comparing the extracted internal standard.

Analysis of apoptosis by annexin V and propidium iodine staining. Rat primary hepatocytes were treated with HIV PIs (25 μM) for 24 h and stained with Annexin V-FITC and propidium iodine using BD ApoAlert Annexin V kit, according to the protocol recommended by the manufacturer. Annexin V/propidium iodine-stained cells were visualized under confocal fluorescence microscopy with a ×40 oil immersion objective using a dual-filter set for FITC and rhodamine (44).

Cell viability assay. Rat primary hepatocytes were plated in 96-well plates with a density of 1 × 104/well. The medium was replaced after 24 h. After treatment with HIV PIs (0 to 50 μM) for 24 h, 10 μl/well of CellTiter 96AQeuous One Solution Reagent was added. After 1 h incubation at 37°C in a humidified 5% CO2 atmosphere, the absorbance at 490 nm was recorded using an ELISA plate reader. Control refers to incubations in the presence of vehicle only (DMSO 0.5%) and was considered as 100% viable cells (44).

Western blot analysis. The nuclear extract and total cell lysates were prepared from cells, as previously described (40). The protein concentration was determined using Bio-Rad protein assay reagent. The nuclear extracts (15 μg of protein) or total cell lysate proteins (20 μg) were resolved on 10% Criterion XT precast gels and transferred to nitrocellulose membranes. Immunoblots were blocked overnight at 4°C with 5% nonfat milk in Tris-buffered saline buffer and incubated with antibodies to CHOP, XBP-1, ATF-4, SREBP-1, SREBP-2, or CYP7A1. Immunoreactive bands were detected using HRP-conjugated secondary antibody and the Western Lightning Chemiluminescence Reagent Plus. The membranes were stripped with stripping buffer (62.5 mM Tris-HCl, pH 6.8 containing 100 mM of β-mercaptoethanol and 2% SDS) and reprobed with antibody against lamin B or β-actin. The density of the immunoblot bands was analyzed using Image J computer software [National Institutes of Health (NIH), Bethesda, MD] (44).

RNA isolation and ribonuclease protection assay. Total RNA was isolated from rat primary hepatocytes using the guanidine thiocyanate cesium chloride centrifugation method. All RPA probes for rat CYP7A1, HMG-CoA-R, LDL-R, sterol 27-hydroxylase (CYP27), and cyclophilin were synthesized using a MAXscript T7 kit from Ambion using a probe-specific DNA fragment that was cloned into a pSP72 vector. The sequence information for all probes is listed in Table 1. The RPA probes were labeled with [α-32P]UTP and isolated using Qiaquick columns. Overnight hybridization was carried out with 8 × 106 counts per minute (cpm) for CYP7A1, CYP27, HMG-CoA-R, LDL-R, and 4 × 106 cpm for cyclophilin, which was used as an internal standard. Verapamil (final concentration 5 μM), were applied onto the cartridge. The cartridges were further washed with 1 ml water, followed by 1 ml methanol-water (30:70, vol/vol) after incubation at room temperature for 30 min. HIV PIs were eluted with 1 ml methanol. The eluent was evaporated under a nitrogen gas stream, and the residue was reconstituted in 250 μl of mobile phase. A 40-μl aliquot was injected onto the HPLC column (33).

Table 1. Sequence information of RPA probes for rat genes

<table>
<thead>
<tr>
<th>Gene Name</th>
<th>GenBank No.</th>
<th>Probe Location, bps</th>
<th>Probe Length, bps</th>
</tr>
</thead>
<tbody>
<tr>
<td>CYP7A1</td>
<td>7549753</td>
<td>238–439</td>
<td>202</td>
</tr>
<tr>
<td>CYP27A1</td>
<td>30578395</td>
<td>647–998</td>
<td>352</td>
</tr>
<tr>
<td>HMG-CoA-R</td>
<td>296924</td>
<td>31–424</td>
<td>394</td>
</tr>
<tr>
<td>LDL-R</td>
<td>31345479</td>
<td>1023–1424</td>
<td>202</td>
</tr>
<tr>
<td>Cyclophilin</td>
<td>205701</td>
<td>129–429</td>
<td>301</td>
</tr>
</tbody>
</table>

Table 1. Sequence information of RPA probes for rat genes
HIV PROTEASE INHIBITOR, THE UPR AND LIPID METABOLISM

Internal control. Twenty grams of total RNA were used in all RPA assays. After RNase digestion, samples were fractionated on 5% acrylamide/8 M urea gels, and bands were visualized by autoradiography using Kodak Biomax MS film. The density of bands was analyzed using Image J computer software (NIH) and normalized to rat cyclophilin.

Real-time quantitative PCR. Total RNA (5 µg) isolated from mouse primary hepatocytes was used for first-strand cDNA synthesis using a high-capacity cDNA archive kit. The mRNA levels of CYP7A1, HMG-CoA-R, and LDL-R were quantified using the specific gene expression assay kits for mouse CYP7A1, HMG-CoA-R, and LDL-R on an ABI PRISM7700 sequence detection system. The mRNA values for each gene were normalized to internal control β-actin mRNA. The ratio of normalized mean value for each treatment group to vehicle control was calculated.

Bile acid synthesis by primary hepatocytes. The conversion of [14C]cholesterol into methanol-water-soluble material was determined as previously described (21). Cells were pretreated with HIV PIs (30 µM) for 12 h, media were replaced and [14C]cholesterol (6 × 10^5 cpm/plate) and HIV PIs were added. After 48 h incubation, media were harvested and analyzed according to the Folch technique (15).

Statistical methods. Student’s t-test was used to analyze the differences between the sets of data. Statistics were performed using GraphPad Pro (GraphPad, San Diego, CA).

RESULTS

Metabolism of HIV PIs in rat primary hepatocytes. Pharmacokinetic studies have shown that most of the HIV PIs have very low systemic bioavailability and are metabolized primarily in the liver by cytochrome P-450 isoforms. Clinical observations indicate that HIV PIs are associated with the development of drug-induced liver injury (38). To optimize the experimental concentrations of HIV PIs in the present study, we first examined the metabolic rates of amprenavir, atazanavir, and ritonavir in primary rat hepatocytes by HPLC. As shown in Fig. 1, the rates of metabolism of tested HIV PIs were similar during the first 6 h of incubation. However, 89% of amprenavir was metabolized after 24 h, whereas larger amounts of atazanavir (45%) and ritonavir (64%) remained after 24 h (Fig. 1 and Fig. 2).

HIV PIs activate the UPR and induce apoptosis in rat primary hepatocytes. Clinical studies have indicated that HIV PIs appear to differ in their ability to cause hyperlipidemia (9, 22). Our previous studies also have shown that most HIV PIs, but not all, were able to activate the UPR and induce apoptosis in mouse macrophages (44). To examine whether HIV PIs have similar effects on the UPR activation in hepatocytes, cells were treated with HIV PIs (30 µM) for various time periods (1 to 24 h), and the expression of CHOP, ATF-4, and XBP-1 was detected by Western blot analysis. As shown in Fig. 3, both atazanavir and ritonavir significantly increased the expression of CHOP, ATF-4, and XBP-1. The expression levels of CHOP, ATF-4, and XBP-1 induced by atazanavir peaked at 2, 6, and 4 h, respectively. The expression levels of CHOP, ATF-4, and XBP-1 induced by ritonavir peaked at 2, 6, and 3 h, respectively. The induction of CHOP, ATF-4, and XBP-1 by atazanavir and ritonavir was concentration dependent (Fig. 4). At 15 µM, the expression of CHOP was increased 49% by atazanavir and 76% by ritonavir; ATF-4 was increased 43% by atazanavir and 63% by ritonavir. The basal level of XBP-1 expression was undetectable but dramatically increased after treatment with atazanavir and ritonavir. However, amprenavir did not significantly affect the expression of CHOP, ATF-4, and XBP-1 even at high concentrations (50 µM). HPLC analysis of amprenavir showed it to be >98% pure and eluted with the same hydrophobicity as reported in the literature (data not shown).

Activation of UPR has been implicated in processes that initiate apoptosis (3, 30). To further examine whether HIV PIs-induced UPR activation is correlated with the apoptosis in rat primary hepatocytes, we treated the cells with 25 µM of HIV PIs for 24 h; morphological changes characteristic of apoptosis were detected with confocal microscopy using Annexin V and propidium iodide staining. As shown in Fig. 5, both atazanavir and ritonavir induced apoptosis, but amprenavir had no significant effect. Our previous studies in macrophages suggest that HIV PIs have different effects on cell viability. In this study, we also compared the cytotoxic effects of amprenavir, atazanavir, and ritonavir on rat primary hepatocytes. As shown in Fig. 6, at 25 µM, the percentages of viable cells after 24 h treatment were 95% for amprenavir, 79% for atazanavir, and 74% for ritonavir, respectively.

Effect of HIV PIs on SREBPs in rat primary hepatocytes. The liver is the major organ responsible for maintaining lipid homeostasis in the body (20). SREBPs are membrane-bound transcription factors that play important roles in the regulation of lipid homeostasis. SREBPs directly activate dozens of genes dedicated to lipid metabolism. In mammalian cells, there are three SREBP isoforms, SREBP-1a, SREBP-1c, and SREBP-2. SREBP-1a is highly expressed in cell lines and activates all SREBP-responsive genes, including those that mediate the synthesis of cholesterol, fatty acids, and triglycerides (36). In contrast, SREBP-1c is the predominant isoform expressed in most tissues with especially high levels in the liver, white adipose tissue, skeletal muscle, adrenal gland, and brain. SREBP-1c primarily regulates the transcription of genes required for fatty acid synthesis but not cholesterol synthesis (36). SREBP-2 preferentially regulates cholesterol synthesis. SREBP-1c and SREBP-2 are the predominant isoforms expressed in the liver (19). Previous studies have shown that indinavir activates SREBP-1 and SREBP-2 in both rat primary hepatocytes and in the liver of intact mice (40). We also have shown that both atazanavir and ritonavir activate SREBPs in mouse macrophages (44). In this study, we compared the effects of three HIV PIs, amprenavir, atazanavir, and ritonavir, respectively.
on SREBP activation in rat primary hepatocytes. As shown in Fig. 7, after 18 h treatment with 30 μM of atazanavir and ritonavir, the mature forms of SREBP-1 were increased by 48% and 44%, respectively; in contrast, activated SREBP-2 levels were increased by only 13% and 8%, respectively. Amprenavir did not detectably increase either SREBP-1 or SREBP-2 levels. We also observed that activated SREBP levels were not altered by either atazanavir or ritonavir until after 12 h treatment and atazanavir and ritonavir induced SREBPs activation was concentration dependent (data not shown). These results were consistent with our previous studies with indinavir in rat primary hepatocytes (40).

**Effect of HIV PIs on mRNA levels of key genes involved in lipid metabolism.** Previous studies (40) have shown that indinavir decreases CYP7A1 mRNA levels and increases HMG-CoA-R mRNA levels, but it has no effect on CYP27 and LDL-R mRNA levels in rat primary hepatocytes. Indinavir also significantly decreases CYP7A1 mRNA half-life. In the current study, we examined the effects of three HIV PIs, amprenavir, atazanavir and ritonavir, on the mRNA levels of CYP7A1, HMG-CoA-R, LDL-R, and CYP27 in rat primary hepatocytes by RPA. Cells were treated with various concentrations of HIV PIs (0 to 100 μM) for 12 or 24 h, total RNA was isolated, and the specific mRNA species were quantified by RPA. As shown in Fig. 8A, amprenavir (25 μM) increased CYP7A1 mRNA by 90% and HMG-CoA-R mRNA by 23% after 24 h but had no significant effect on LDL-R and CYP27 mRNA levels. Atazanavir (25 μM) significantly decreased CYP7A1 mRNA by 38% and increased LDL-R mRNA by 20% but did not change the HMG-CoA-R mRNA level after 24 h treatments (Fig. 8B). At the high concentration (100 μM), atazanavir also decreased LDL-R and CYP27 mRNA levels. Ritonavir (25 μM) significantly decreased CYP7A1 mRNA by 56% but had no effect on HMG-CoA-R and CYP27. Ritonavir also decreased LDL-R mRNA at high concentrations (50 and 100 μM) (Fig. 8C).

We further confirmed the effects of HIV PIs on CYP7A1, HMG-CoA-R, and LDL-R mRNA expression in mouse primary hepatocytes using real-time PCR. We isolated mouse primary hepatocytes and treated the cells with individual HIV PI (25 μM) for 24 h; mRNA levels of HMG-CoA-R, CYP7A1, and LDL-R were quantified using real-time PCR. As shown in Fig. 9, amprenavir increased HMG-CoA-R and CYP7A1 mRNA levels by 23% and 53%, respectively, it had no effect on CYP27 mRNA. Atazanavir decreased CYP7A1 mRNA by 60% and increased LDL-R mRNA by 54%, it but had no effect on HMG-CoA-R. Ritonavir decreased CYP7A1 mRNA by 49% but did not affect HMG-CoA-R and LDL-R mRNA levels. These results are consistent with the RPA results.

**Fig. 2. Metabolism of HIV PIs in rat primary hepatocytes.** Representative chromatograms of amprenavir (A), atazanavir (B), and ritonavir (C). Rat primary hepatocytes were treated with 30 μM of HIV PIs for 0 (0) or 24 (b) h. The drug concentrations in culture media were determined by HPLC, as described under MATERIALS AND METHODS. Open arrow indicates drug peak, and solid arrow indicates drug metabolites.
Effect of HIV PIs on protein levels of CYP7A1 in rat primary hepatocytes. To further examine whether HIV PI-induced increase or decrease of mRNA level of CYP7A1 is correlated with the changes of protein expression, we performed the Western blot analysis using a polyclonal antibody against mouse CYP7A1. As shown in Fig. 10, both atazanavir and ritonavir significantly decreased CYP7A1 levels by 28%, but amprenavir had no effect.

Effect of HIV PIs on bile acid synthesis in rat primary hepatocytes. Bile acid can be synthesized by either the “neutral” or “acidic” pathway in hepatocytes (11). To further determine whether HIV PI-induced downregulation of CYP7A1 expression is correlated with a decrease in bile acid synthesis in rat hepatocytes, the bile acid biosynthesis was estimated by measuring conversion of [14C]cholesterol to water-soluble radioactivity (21) after treatment with HIV PIs for 48 h. As shown in Fig. 11, atazanavir and ritonavir significantly inhibited bile acid synthesis by 47% and 67%, respectively, but amprenavir had no significant effect.

DISCUSSION

Since the introduction of HIV PIs into HAART, the mortality rate of HIV-infected patients has dramatically decreased. However, HAART has changed the clinical profile of HIV infection from a subacute lethal disease to a chronic ambulatory disease (41). Dyslipidemia specifically associated with HIV PIs, which are one of the cornerstones in HAART, has emerged as an important issue in HIV-infected patients (9, 10, 37). More than 50% of the patients receiving HAART develop lipid abnormalities, including elevated levels of total serum cholesterol, LDL-cholesterol, and triglycerides, which are well-known risk factors for cardiovascular diseases (34). We and others have demonstrated that HIV PIs induced UPR, which may represent an important cell signaling mechanism of HIV PI-induced metabolic syndromes (28, 44).

In the present study, we compared the direct effects of three HIV PIs on activation of the UPR and hepatic lipid metabolism using rodent primary hepatocytes as a model system. Our results demonstrated that these HIV PIs, amprenavir, atazanavir, and ritonavir, had different effects on the UPR activation, cell apoptosis, and lipid metabolisms in primary hepatocytes. Pharmacokinetetic studies have shown that all of the HIV PIs are metabolized in the liver by various isoforms of the cyto-
Drug-induced hepatotoxicity has been observed in patients undergoing HAART. Although the mechanisms of drug-induced liver injury are poorly identified, it is clear that HIV PIs are a contributing factor, and different HIV PIs may have different effects. The data from the current studies clearly showed that both atazanavir and ritonavir significantly induced apoptosis in rat primary hepatocytes, but amprenavir had no significant effect. Apoptosis is a form of cell death that involves multiple pathways. In addition to a death receptor and mitochondrial pathways, ER stress-induced cellular death also plays an important role in regulating normal cell function. Our previous studies have demonstrated that HIV PIs activated the UPR and induced apoptosis, and different HIV PIs varied greatly in their ability to activate the UPR in macrophages. Recent studies done by Parker et al. also demonstrated that HIV PIs induced an ER stress response in human HepG2 and TC5 hepatocytes cell lines and mouse 3T3-L1 adipocytes, suggesting that ER stress may contribute to HIV PI-induced lipodystrophy. It also has been shown that ritonavir increases endothelial permeability, decreases levels of tight junction proteins, and increases superoxide anion production. HIV PI-induced endothelial dysfunction represents one of the important mechanisms of vascular lesion formation and also contributes to HIV PI-associated cardiovascular diseases.

Fig. 5. HIV PIs induce apoptosis in rat primary hepatocytes. Rat primary hepatocytes were treated with vehicle control or individual HIV PIs (25 μM) for 24 h, then stained with annexin V-FITC (a) and propidium iodine (b). Thapsigargin (100 nM) was used as a positive control. Images of Annexin V-FITC and propidium iodine-stained cells were visualized under confocal fluorescence microscopy with a dual filter set for FITC and rhodamine. A: vehicle control, DMSO. B: AMPV. C: ATZV. D: RITV. E: thapsigargin, TG.

Fig. 6. Effects of HIV PIs on cell viability. Rat primary hepatocytes were treated with individual HIV PIs (0 to 50 μM) for 24 h. The cell viability was measured using CellTiter 96AQOne Solution Reagent, as described under MATERIALS AND METHODS. Control refers to incubations in the presence of vehicle only (DMSO 0.5%) and was considered as 100% of viable cells. Each bar represents the means ± SE of 3 independent experiments. Statistical significance relative to vehicle control, *P < 0.05.

Fig. 7. Effects of HIV PIs on the activation of SREBP-1 and SREBP-2 in rat primary hepatocytes. Representative immunoblots against mature forms of SREBP-1, SREBP-2, and lamin B from the nuclear extracts of rat primary hepatocytes treated with 30 μM of HIV PIs for 18 h. Lamin B was used as a loading control.
literature (data not shown). Drug metabolism analysis also indicated that the metabolic rate of these three HIV PIs were similar during the first 6 h of incubation (Fig. 1). Therefore, the different response appears to reflect the intrinsic properties of the individual HIV PI.

Cholesterol and lipid metabolisms are controlled by SREBPs, which are considered to be master regulators of lipid homeostasis (13). Our previous studies showed that indinavir increased the levels of transcriptionally active of SREBP-1 and SREBP-2 in both primary rat hepatocytes and in the liver of intact mice (40). In macrophages, we also found that HIV PIs markedly increased mature forms of nuclear SREBPs, which might contribute to an increase in intracellular lipids and foam cell formation (44). In the current study, we observed that both atazanavir and ritonavir increased the levels of mature SREBPs in hepatocytes (Fig. 7), but amprenavir had little effect. Because the activation of SREBPs by HIV PIs was detected after 12 h treatment and the metabolic rate of amprenavir after 6 h was much faster than that of atazanavir or ritonavir, a lower active drug concentration might contribute to the lower effect of amprenavir on SREBPs in hepatocytes. However,

A

B

C

Fig. 8. Effects of HIV PIs on steady-state mRNA levels of genes involved in cholesterol and bile acid metabolism in rat primary hepatocytes. Rat primary hepatocytes were treated with various concentrations of HIV PIs (0 to 100 μM) for 12 or 24 h. Total RNA was isolated. The mRNA levels of cholesterol 7α-hydroxylase (CYP7A1), 3-hydroxy-3-methylglutaryl-coenzyme A reductase (HMG-CoA-R), low-density lipoprotein receptor (LDL-R), and sterol 27-hydroxylase (CYP27) were determined by ribonuclease protection assay (RPA) analysis. Relative amounts of mRNA were determined by quantifying the densities of the radioactive bands using Image J software and normalized to rat cyclophilin mRNA as loading control. A: amprenavir. B: atazanavir. C: ritonavir.

Fig. 9. Real-time quantitative PCR of key genes involved in cholesterol and bile acid metabolism in mouse primary hepatocytes. Primary mouse hepatocytes were isolated as described under MATERIALS AND METHODS and treated with HIV PIs (25 μM) or vehicle control for 24 h. Total cellular RNA was isolated using Ambion RNAqueous kit and the first-strand cDNA was synthesized using high-capacity cDNA archive kit. The mRNA levels of HMG-CoA-R, CYP7A1, and LDL-R were quantified using the specific gene expression assay kits for mouse HMG-CoA-R, CYP7A1, and LDL-R on an ABI PRISM7700 Sequence Detection System. The mRNA values for each gene were normalized to internal control β-actin mRNA. The ratio of normalized mean value for each treatment group to vehicle control was calculated. The values are expressed as means ± SE of 3 independent experiments. Statistical significance relative to vehicle control, *P < 0.05. A: relative mRNA levels of HMG-CoA-R. B: relative mRNA levels of CYP7A1. C: relative mRNA levels of LDL-R.
HIV PROTEASE INHIBITOR, THE UPR AND LIPID METABOLISM

Fig. 10. Effects of HIV PIs on CYP7A1 protein expression in rat primary hepatocytes. A: representative immunoblots against CYP7A1 and β-actin from total cell lysates of rat primary hepatocytes treated with 25 μM of HIV PIs for 24 h. β-Actin was used as a loading control. B: relative protein levels of CYP7A1 were determined by quantifying the densities of the immunoreactive bands using Image J software and normalized to loading control (β-actin). *p < 0.05.

amprenavir also did not alter SREBP expression, even though it was very slowly metabolized in macrophages (43). It is currently unclear how HIV PIs affect SREBPs’ maturation and why the UPR activation is associated with an increase in the mature forms of SREBPs. Several studies suggest that one potential mechanism may be that HIV PIs inhibit the degradation of the SREBPs by modulating proteasome activity (18, 27, 35). However, recent studies by Lee and Ye (23) showed that the proteolytic activation of SREBPs may occur in UPR-activated cells as a result of cellular depletion of insig-1. Insig-1 (1 and 2) reside in the ER membrane and play an essential role in the processing of SREBPs. SREBPs form a complex with SREBP cleavage-activating protein (SCAP) in the ER. If the cell is depleted of sterols, SCAP is involved in the transport of SREBPs to the Golgi for proteolytic processing (13). When cellular cholesterol levels are high, SCAP binds to Insig-1 and inhibits the movement of SREBP-SCAP complex to the Golgi. Activation of the UPR causes a marked decrease in the rate of protein synthesis (31). Because the half-life of Insig-1 is estimated to be less than 2 h, cells become rapidly depleted of Insig-1 after UPR activation. This may allow for the proteolytic processing of SREBPs even at high cellular levels of cholesterol. Whether HIV PIs directly inhibit insig-1 expression in hepatocytes awaits further study.

The liver is the major organ responsible for maintaining lipid homeostasis in the body. Under normal physiological conditions, cholesterol input equals cholesterol output (20). Cholesterol input pathways in the liver include receptor-mediated endocytosis of cholesterol-carrying lipoproteins and de novo cholesterol biosynthesis. There are two major cholesterol output pathways: direct secretion of excess cholesterol into the canaliculus and bile acid synthesis. CPY7A1 and CYP27 are the key enzymes in the two major pathways of bile acid biosynthesis: “neutral pathway” and “acidic pathway,” respectively (20). Previous studies have shown that indinavir induces free intracellular cholesterol accumulation in rat primary hepatocytes. This is followed by upregulation of HMG-CoA-R and downregulation of CPY7A1 but no effect on CYP27 and LDL-R expression (40). In the present study, we found that HIV PIs had a different effect on CPY7A1 and HMG-CoA-R expression. Atazanavir and ritonavir significantly inhibited CPY7A1 mRNA and protein expression but had no effect on HMG-CoA-R, both in rat and mouse hepatocytes. Our previous studies have shown that indinavir decreased the half-life of CPY7A1 mRNA (40). We also found that both atazanavir and ritonavir decreased CPY7A1 mRNA half-life in rat primary hepatocytes (data not shown). Consistently, bile acid biosynthesis in atazanavir- and ritonavir-treated cells was significantly decreased. Although amprenavir significantly increased CPY7A1 mRNA level, it had no significant effect on CPY7A1 protein levels and rates of bile acid synthesis (Figs. 10 and 11). Clinical observation indicates that atazanavir and amprenavir seem to have less impact on the lipid profile in patients (4, 11, 16, 26). The results from the current study suggest that the increase of LDL-R expression by atazanavir and CYP7A1 expression by amprenavir may contribute, at least partially, to their lower impact on dyslipidemia. However, HIV PIs tested in vitro may have different biological activities in vivo because of the plasma protein-binding properties of these drugs. All of the HIV PIs currently used in clinical trials, except indinavir, are tightly bound to serum proteins. The concentration tested in vitro is much higher than the calculated free plasma concentration. The kinetics between plasma-bound and free HIV PIs in vivo is still not clear. Further in vivo investigations are needed to elucidate the cellular mechanisms of HIV PI-induced dyslipidemia.

In summary, we have demonstrated that individual HIV PIs have different effects on the UPR activation and expression of key genes involved in lipid metabolism in hepatocytes. HIV PI-induced UPR appears to contribute to lipid dysregulation. These in vitro models may be useful in predicting possible clinical adverse effects by HIV PIs on lipid metabolism in vivo.
ACKNOWLEDGMENTS

We would like to thank the following companies for providing us with the following compounds used in this research: GlaxoSmithKline (amprenavir); Abbott Laboratories (ritonavir); and Bristol-Myers-Squibb (atazanavir). We also want to thank Dr. David W. Russell for providing us with the CYP7A1 antibody.

Present address for Hong Ding: Department of Pharmacology, School of Pharmacy, Wuhan Univ., Wuhan, P.R. China, 430072.

GRANTS

This work is supported by grants from the National Institutes of Health (RO1 AR05189 and F10 NK 038030), GlaxoSmithKline research fund, and the A.D. Williams fund.

REFERENCES


