P2X7 receptor-mediated purinergic signaling promotes liver injury in acetaminophen hepatotoxicity in mice

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Hoque R, Sohail MA, Salhanick S, Malik AF, Ghani A, Robson SC, Mehal WZ. P2X7 receptor-mediated purinergic signaling promotes liver injury in acetaminophen hepatotoxicity in mice. Am J Physiol Gastrointest Liver Physiol 302: G1171–G1179, 2012. First published March 1, 2012; doi:10.1152/ajpgi.00352.2011.—Inflammation contributes to liver injury in acetaminophen (APAP) hepatotoxicity in mice. APAP hepatotoxicity was assessed in mice genetically deficient in P2X7, the key inflammatory receptor for nucleotides (P2X7−/−), and in wild-type mice. P2X7−/− mice had significantly decreased APAP-induced liver necrosis. In addition, APAP-poisoned mice were treated with the specific P2X7 antagonist A438079 or etheno-NAD, a competitive antagonist of NAD. Pre- or posttreatment with A438079 significantly decreased APAP-induced necrosis and hemorrhage in APAP liver injury in wild-type but not P2X7−/− mice. Pretreatment with etheno-NAD also significantly decreased APAP-induced necrosis and hemorrhage in APAP liver injury. In addition, APAP toxicity in mice lacking the plasma membrane ecto-NTPDase CD39 (CD39−/−) that metabolizes ATP was examined in parallel with the use of soluble apyrase to deplete extracellular ATP in wild-type mice. CD39−/− mice had increased APAP-induced hemorrhage and mortality, whereas apyrase also decreased APAP-induced mortality. Kupffer cells were treated with extracellular ATP to assess P2X7-dependent inflammasome activation. P2X7 was required for ATP-stimulated IL-1β release. In conclusion, P2X7 and exposure to ATP and NAD that serve as damage-associated molecular patterns (DAMPs) (12). This results in the activation of caspase-1 and cleavage of pro-IL-1β and pro-IL-18 to IL-1β and IL-18.

It is unclear what signals the assembly of the inflammasome following APAP-induced hepatotoxicity. It is known that P2X7 receptor stimulation by extracellular ATP leads to inflammation during APAP-induced hepatotoxicity (10). Proinflammatory cytokines IL-1β and IL-18 are thought to be crucial to the propagation of inflammation during APAP-induced hepatotoxicity (10). Production of these cytokines is dependent on cleavage by caspase-1, which is, in turn, dependent on the assembly of the inflammasome, a protein scaffold that assembles in response to a variety of danger signals or as damage-associated molecular patterns (DAMPs) (12). This results in the activation of caspase-1 and cleavage of pro-IL-1β and pro-IL-18 to IL-1β and IL-18.

The role of purinergic signaling mechanisms in APAP-induced hepatotoxicity is not known and has therapeutic implications as active targets for anti-inflammatory drug development that would include receptor agonists/antagonists or inducing or inhibiting the hydrolysis of ATP and other ligands. In conclusion, P2X7 and exposure to ATP and NAD that serve as damage-associated molecular patterns (DAMPs) (12). This results in the activation of caspase-1 and cleavage of pro-IL-1β and pro-IL-18 to IL-1β and IL-18.

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naling, therefore, provides attractive targets for therapeutic intervention in APAP toxicity.

MATERIALS AND METHODS

Animals. C57BL/6 male mice 5 to 8 wk of age were purchased from the National Cancer Institute or Taconic laboratories. P2X7−/−, and CD39−/− mice have been described (8, 32). All experiments and animal handling were performed under approved protocols at the Yale University and Beth Israel Deaconess Medical Center Institutional Animal Care and Use Committees.

APAP-induced hepatotoxicity. APAP (Sigma-Aldrich, St. Louis, MO) solution was prepared as described (10). APAP was dosed at 500 mg/kg and administered by intraperitoneal injection after 15 h of starvation. Animals were euthanized by isoflurane or ketamine/xyla-

zine at 6 and 12 h for collection of serum, isolation of liver lymphocytes, or collection of liver tissue for histology, or they were observed every 4 h for 72 h until they became moribund.

Treatment with apyrase, etheno-NAD, and A438079. Mice were treated by intraperitoneal injection with potato apyrase (Sigma-Al-
drich) at 4 U/mouse 30 min prior and 6 h after APAP injection, etheno-NAD (Sigma-Aldrich) at 2 mg/mouse 1 h prior and 6 h after APAP injection, and A438079 (Tocris, Ellisville, MO) at 2 mg/mouse either 1 h prior or 2 h after APAP injection.

Liver histology scoring. Liver histology was scored in a blinded manner in hematoxylin and eosin-stained, paraffin-embedded sec-
tions. Necrosis was scored from 0 to 3 when present in 0, 0 – 25%, 25–50%, or > 50% of the field, respectively. Hemorrhage was scored from 0 to 3 when present in 0, 0–25%, 25–50%, or > 50% of the field, respectively. At least five fields per section were examined under ×4 magnification and data are expressed as mean scores per experimental group.

Quantitation of liver-infiltrating neutrophils. Neutrophil quantita-
tion was performed in paraffin-embedded liver sections after immu-
nolabeling with GR-1 monoclonal antibody (BD Biosciences, San Jose, CA) by scoring for positive cells in five high-power fields (×40). To confirm our results for neutrophil immunostaining, we immuno-
labeled liver sections for another neutrophil-specific epitope using Ly-6B.2 monoclonal antibody (AbD Serotec, Raleigh, NC). Imaging results represent Ly-6B.2 immunostained images.

Serum ALTs. Serum was isolated from mice and alanine amino-
transferase (ALT) levels were determined in the Yale New Haven Hospital clinical chemistry laboratory.

Quantitation of CYP 2E1 expression and APAP adducts. Western blots of liver lysates were immunostained with rabbit anti-mouse IgG for CYP 2E1 (Abcam, Cambridge, MA), rabbit anti-APAP adduct IgG (gift of Lance Pohl, National Heart, Lung, and Blood Institute, Bethesda, MD), or rabbit anti-mouse IgG for β-actin (Abcam). The secondary antibody for immunodetection was goat anti-rabbit IgG horseradish peroxidase conjugate (Santa Cruz Biotechnology, Santa Cruz, CA). Immunodetection was performed with SuperSignal West Pico Chemiluminescent Substrate (Thermo Scientific, Logan, UT). Densitometry of the predicted bands was determined using a Fotof/ Analyst Investigator digital imager (Fotodyne, Hartland, WI) and PC Imager software. The ratio of CYP2E1 and APAP adduct bands to β-actin bands was determined and normalized to the value of un-
treated wild-type animal liver run on the same Western blot analysis, which was set to one.

Caspase-1 activity assay. Snap-frozen liver tissue stored in liquid nitrogen was homogenized with a rotor/stator homogenizer in cell lysis buffer, and 300 mg of liver protein was then incubated in a 96-well microtiter dish for 1 h at 37°C with the fluorescent caspase-1 substrate YVAD-AFC as per the supplier (Biovision, Mountain View, CA). Change in fluorescence at 505 nm after excitation at 400 nm was then determined with a Biotek Synergy fluorescent plate reader (Biotek, Winoski, VT). Values were normalized to blank samples containing assay buffer, YVAD-AFC substrate, and no liver protein, and expressed as fold change from untreated wild-type liver lysate run in the same experiment.

Kupffer cell isolation and treatment. Liver nonparenchymal cells were isolated as previously described with the following modifications (4). Mouse nonparenchymal cells were resuspended in 13% OptiPrep (Axis Shield, Norton, MA) in HBSS. This was layered over 18% OptiPrep in HBSS and then overlayed with HBSS. This was centrifuged at 1,400 g for 20 min at 4°C. The top layer and top interface were then recovered and plated on 24-well polystyrene dishes at 300,000 cells/well in DMEM supplemented with 10% FBS, 100 U/ml penicillin, 100 μg/ml streptomycin, and gentamicin 50 μg/ml. One to two hours after plating, nonadherent cells were removed and adherent cells were assessed for cell surface expression of CD45 and F4/80 cell surface markers by fluorescence activated cell sorting using mouse anti-mouse CD45.2 phycoerythrin-conjugated antibody (BD Biosciences) and rat anti-mouse F4/80 allopolyocya-
nin-conjugated antibody (eBiosciences, San Diego, CA). This data is shown in Fig. 5B. Cells were incubated overnight and then treated with LPS (Sigma-Aldrich) at 100 ng/ml for 6 h and then ATP (Sigma-Aldrich) at 5 mM for 20 min. Supernatant was collected, and cell lysate prepared in Cell Lysis Buffer (Cell Signaling, Danvers, MA) with Complete Protease Inhibitor (Roche, Mannheim, Germany).

ELISA for IL-1β release. High-binding, 96-well ELISA plates (BD Biosciences) were used per manufacturer instructions. The capture antibody was anti-mouse IL-1β (R&D Systems, Minneapolis, MN). 100 μl of Kupffer cell supernatant was added to each well and incubated overnight at 4°C. The detection antibody, polyclonal bio-
tylinated anti-mouse IL-1β antibody (R&D Systems), was then added for 2 h at room temperature.

Statistics. Kaplan-Meier plots and statistical analysis were performed using Microsoft Excel 2007 and MedCalc software version 9.2.0.1. Unpaired two-tailed Student’s t-test was used to compare groups. A P value of < 0.05 was considered significant.

RESULTS

P2X7 is required for maximum liver injury in APAP hepatotoxicity. Genetic deletion of P2X7 significantly reduces the liver necrosis score (1.3 ± 0.2 vs. 2.4 ± 0.3) and neutrophil count in the liver (9.9 ± 1.1 vs. 17.0 ± 1.0) relative to wild-type animals at 12 h post-APAP treatment (Fig. 1, A–B and D–E, respectively). Deletion of P2X7 also significantly decreases serum ALT values at 6 h post-APAP treatment (265 ± 53 vs. 3,526 ± 1,272) as shown in Fig. 1C.

Pretreatment with a P2X7 antagonist A438079 before APAP prevented liver necrosis (0.0 ± 0.0 vs. 1.6 ± 0.2), significantly reduced hemorrhage (0.7 ± 0.3 vs. 1.6 ± 0.2), significantly reduced neutrophil count in the liver (9.9 ± 1.1 vs. 17.0 ± 1.0), and significantly reduced serum ALT values (310 ± 358 vs. 3,526 ± 1,272) as shown in Figs. 2, A–B and 3, D–E, respectively. Posttreatment with the P2X7 antagonist A438079 after APAP significantly reduced liver hemorrhage (0.4 ± 0.2 vs. 1.7 ± 0.4) as shown in Fig. 2, A and B but did not significantly alter liver necrosis (Fig. 2, A and B) or serum ALT values at 12 h posttreatment (6,200 ± 923 vs. 5,117 ± 1,117). A438079-dependent reductions in APAP liver injury required P2X7 as pretreatment with A438079 did not significantly change liver injury in P2X7−/− animals (Figs. 2, C and D).

Genetic deficiency of P2X7 or pretreatment of wild-type animals with the P2X7 antagonist A438079 significantly decreased liver caspase-1 activity relative to wild-type animals given APAP (1.0 ± 0.2 and 1.0 ± 0.2 vs. 1.6 ± 0.2) as shown in Fig. 2E. Genetic deficiency of P2X7 or pretreatment of

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wild-type animals with the P2X7 antagonist A438079 did not significantly change liver CYP2E1 expression or APAP adduct formation 6 h post-APAP treatment compared with wild-type animals given APAP (Fig. 2).

Extracellular ATP and NAD promote liver injury in APAP hepatotoxicity. Treatment with apyrase or etheno-NAD significantly reduced liver necrosis (1.7 ± 0.1 and 1.4 ± 0.1 vs. 2.0 ± 0.0) and liver hemorrhage (1.8 ± 0.1 and 1.3 ± 0.2 vs. 2.4 ± 0.2) in APAP liver injury relative to saline vehicle-treated animals shown in Fig. 3, A and C. Treatment with apyrase or etheno-NAD significantly reduced neutrophil count in the liver (8.9 ± 1.4 and 9.3 ± 0.7 vs. 17.0 ± 1.0) as shown in Fig. 3D. Serum ALTs were significantly reduced with pretreatment with etheno-NAD or apyrase (4,083 ± 830 and 4,480 ± 851 vs. 7,320 ± 313) shown in Fig. 3E. Treatment with apyrase also significantly decreased mortality at 72 h post-APAP treatment relative to saline vehicle (1/10 vs. 9/10 expired animals) shown in Fig. 3F.

Genetic deletion of CD39 is expected to increase local concentrations of extracellular ATP and NAD at sites of injury and resulted in significantly increased liver hemorrhage (1.0 ± 1.0 and 3.0 ± 0.0) at 12 h post-APAP treatment shown in Fig. 4, A and B. Genetic deletion of CD39 also significantly increased mortality at 24 h post-APAP treatment relative to saline vehicle (6/8 vs. 1/7 expired animals) as shown in Fig. 4C.

P2X7 is required for IL-1β release in Kupffer cells. Liver Kupffer cells were isolated from wild-type and P2X7−/− mice. Treatment with LPS and ATP-induced caspase-1 proteolytic activation and IL-1β release in wild-type but not...
Fig. 2. Pre- and posttreatment with the antagonist A438079 prevents liver injury and inflammation in APAP-induced acute liver injury in a P2X7-dependent manner. WT mice were administered saline vehicle (n = 8) or A438079 at 300 mM/kg ip (n = 4) 1 h before or 2 h after (n = 5) administration of APAP at 500 mg/kg ip. P2X7−/− mice were administered saline vehicle (n = 5) or A438079 at 300 mM/kg ip (n = 5) 1 h before administration of APAP at 500 mg/kg ip. In control experiments, WT mice and P2X7−/− mice were also administered saline (n = 2 per group) or A438079 (n = 2 per group) 1 h prior to PBS vehicle. At 12 h post-APAP treatment, liver histology was assessed in WT mice (A and B) and P2X7−/− mice (C and D). Liver caspase-1 activity (E), liver CYP2E1 expression (F), and liver APAP adducts (F) were measured at 6 h post-APAP in WT mice (n = 5), P2X7−/− mice (n = 7), and WT pretreated with A438079 as above (n = 4). *P < 0.05.
Kupffer cells (Fig. 5). In parallel experiments, thioglycollate-elicited peritoneal macrophages obtained from wild-type but P2X7−/− mice could be induced to release IL-1β in response to LPS and ATP. The Kupffer cell isolation was >92% positive for the macrophage marker F4/80 as determined by flow cytometric analysis.

**DISCUSSION**

Liver injury following APAP overdose is initiated by the metabolism of excess amounts of APAP. A portion of ingested APAP undergoes reductive metabolism, primarily via CYP2E1 to produce the electrophilic reactive metabolite, n-aminopara-benzoquinone imine, which binds intracellular proteins resulting in mitochondrial failure (36). However, APAP metabolites are only found in the centrilobular areas of the liver following acute poisoning. As the time course of liver injury following APAP demonstrates, worsening liver injury and panlobular necrosis 8–16 h after the completion of APAP metabolism in mice and humans, this supports a second mechanism in propagating liver injury (16).

Propagation of hepatocellular injury occurs through activation of the innate immune system and induction of inflammatory injury, although the contribution of this pathway is debated (5, 10, 11, 13, 20–22, 34). Necrotic cells or cells undergoing apoptosis release intracellular contents, which have been demonstrated to induce sterile inflammation in a variety of models of tissue injury (27). A key example is DNA from apoptotic cells, which can bind TOLL-like receptor 9 (TLR9) (7, 17). In the setting of APAP-induced injury, Imaeda et al. (10) identified TLR9 activation by assaying IL-1β in mice deficient in Tlr9, finding that APAP as well as DNA from apoptotic mammalian cells, increased pro-IL-1β transcript in the wild-type animals in a TLR9-dependent manner, and more importantly demonstrating that Tlr9 null animals were pro-
tected from hepatotoxicity. They further studied animals deficient in the components of the NRLP3 inflammasome required to activate caspase-1 and cleave pro-IL-1β to form IL-1β. They found that deficiency of nod-like receptor family, pyrin domain containing-3 (Nlrp3) and caspase-1 reduced liver injury and mortality without effecting pro-IL-1β transcript levels, demonstrating that posttranscriptional processing of procytokines, is also a key regulated determinant of innate immune injury in APAP.

Intrahepatic release of DAMPs has been demonstrated after APAP toxicity, and apoptotic and secondary necrotic cells following the initial APAP-induced injury release DNA, which activates TLR9 to transcribe pro-IL-1β and pro-IL-18 (23). These procytokines are cleaved by caspase-1 activated by the NLRP3 inflammasome. In turn, mature IL-1β and IL-18 upregulate innate immune responses in local immune cells (23, 33). Specifically, Kupffer cells (the resident macrophages of the liver) can propagate the hepatic innate inflammatory response through the production of proinflammatory TNF-α (19, 20). Subsequently there is a migration of neutrophils into the liver, and the ensuing cytotoxic actions are thought to cause further cause hepatocellular damage (21).

The link between the initial necrosis in APAP injury and upregulation of inflammasome activity has not, to date, been elucidated. Activation of the NLRP3 inflammasome can be triggered by extracellular ATP, a ligand for the purinergic receptor P2X7. As such, we investigated purinergic mechanisms, specifically ATP and NAD activation of the P2X7 receptor, in the setting of necrotic cell death induced by APAP. There is increasing evidence that P2X7 mediates sterile inflammation in many organs, including the lung (28) and the pancreas (9). Despite this accepted role of the P2X7 receptor in the sterile inflammatory response, there are important organ-specific differences. In the lung, inhibition of P2X7 receptor function results in a very significant reduction in inflammation and injury, but this occurs to a lesser degree in the pancreas. Of note, the contribution of P2X7 activation to sterile inflammation in the liver was previously unknown.

Using genetically deficient animals, receptor antagonists, and enzymatic depletion, we have demonstrated an important role for purinergic signaling methods in APAP liver injury and inflammation. P2X7 was required for full injury in APAP hepatotoxicity (Figs. 1B and 2B). This P2X7-dependent effect did not appear to be through regulation of CYP2E1 expression or alteration of APAP metabolism and APAP adduct formation (Fig. 2F). P2X7 was required for innate immune response to APAP hepatotoxicity, specifically hepatic caspase-1 activation (Fig. 2E) and neutrophil migration into the liver (Figs. 1E and 3D). In addition to this central finding, our work supports the use of P2X7 specific antagonists in liver injury after APAP, and may reduce the incidence of APAP-induced acute liver failure (Figs. 2B and 3E).

It is of interest that the P2X7 receptor antagonist A438079 reduced APAP liver injury to a greater degree than seen in the
P2X7-deficient mouse (Figs. 1 and 2). To confirm that this finding was not related to off-target effects of the antagonist, we demonstrated that A438079 did not significantly alter APAP liver injury in P2X7-deficient mice (Fig. 2, C and D). The reasons for these differences are therefore likely attributable to developmental adaptations of the immune system in the P2X7 receptor deficient mice, with upregulation of non-P2X7 receptor-dependent pathways.

Regarding the endogenous ligands for P2X7, NAD and ATP were first identified as DAMPs released by injured erythrocytes (30). ATP was later identified as a DAMP-type ligand for P2X7 released from extracellular mitochondria of necrotic cell origin (12). Extracellular NAD is a much more potent ligand for P2X7 receptor activation, presumably due to membrane localization through NAD-ribosylation (31). We provide evidence that endogenous ATP and NAD likely released from injured hepatocytes in APAP toxicity are required for hepatic necrosis and sinusoidal injury with intrahepatic hemorrhage. Specifically, depletion of extracellular ATP with apyrase and competitive antagonism of NAD-ribosylation sites with etheno-NAD both resulted in decreased liver necrosis and hemorrhage (Fig. 3, C and E). Additionally, apyrase and etheno-NAD also reduced neutrophil migration into the liver to a significant but lesser degree than use of a P2X7 antagonist (Fig. 3D). Finally, apyrase decreased mortality from APAP-induced acute liver failure (Fig. 3F). To further confirm that endogenous ATP and NAD are key determinants of sterile liver injury and mortality in APAP hepatotoxicity, we demonstrated that genetic deletion of CD39, the predominant ecto-NTPase in the metabolism of extracellular ATP and NAD, enhanced liver hemorrhage and increased mortality in this model (Fig. 4, B and C). Several cell types in the liver are known to express P2X7 and CD39. Liver sinusoidal endothelial cells and Kupffer cell subpopulations express P2X7, but function was not confirmed in these studies (1, 2, 35). Liver sinusoidal endothelial cells and Kupffer cell also express CD39 (1, 2). The role of Kupffer cells in promoting liver injury in APAP hepatotoxicity was demonstrated in macrophage depletion experiments employing gadolinium chloride (26). Additionally, selective deletion of macrophages employing diphtheria toxin and the CD11b-DTR transgene implicated macrophages in the progression of toxin-mediated liver injury (6). We now show that Kupffer cells respond to extracellular ATP in a P2X7-dependent manner and through activation of the NLRP3 inflammasome with IL-1β release (Fig. 5). It is possible that functional P2X7 on hepatocytes or sinusoidal endothelial cells may also mediate APAP hepatotoxicity through recognition of ATP and NAD and P2X7-mediated cell permeabilization and cell death.

**Injured Hepatocyte**  
Mitochondria  
NAD and ATP  

**DAMP Sensing Cell**  
DAMPs  
TLRs  
Pro-IL-1β  
P2X7  
ASC  
NLRP3  
Caspase-1  
IL-1β  
Liver Injury
Inflammation promotes liver injury in APAP hepatotoxicity, mediated at least in part through DAMP-mediated innate immune signaling. DAMP receptors previously implicated in APAP hepatotoxicity include TLR4 and TLR9, which recognize the DAMPs high mobility group box-1 and self DNA, respectively (1, 12, 39). Through activation of the NLRP3 inflammasome, DAMP-mediated activation of P2X7 provides a link between TOLL-like receptor mediated transcriptional induction of proinflammatory cytokines, specifically pro-IL-1β and pro-IL-18, and cytokine maturation and secretion (Fig. 6).

In summary, we have identified P2X7 and extracellular ATP and NAD as significant contributors to liver injury, inflammation, and mortality, in APAP hepatotoxicity. We have also shown that Kupffer cells have functional P2X7 and may thereby mediate these effects through activation of the NLRP3 inflammasome consistent with the paradigm of DAMP mediated sterile inflammation.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

AUTHOR CONTRIBUTIONS


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