Impaired ghrelin signaling is associated with gastrointestinal dysmotility in rats with gastroesophageal reflux disease

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Submitted 2 November 2011; accepted in final form 17 April 2012

Gastroesophageal reflux disease (GERD) is a condition in which regurgitation of gastric acid and other gastric contents into the esophagus causes discomfort and complications (49). Decreased the lower esophageal sphincter function and esophageal clearance are involved in impairment of the esophageal mucosa due to gastric acid (4), and proton pump inhibitors, which suppress gastric acid secretion, have been the drugs of choice for the treatment of GERD. Delayed gastric emptying occurs frequently among patients with acid reflux (3, 31), and treatment with prokinetic agents, which are agonists of the 5-hydroxytryptamine 4 (5-HT4) receptor, partially improves symptoms in patients with GERD (16, 39). Thus it was concluded that gastric dysmotility is also a factor involved in acid reflux into the esophagus (3, 10, 29, 31). However, the reasons for gastric dysmotility remain unclear.

Ghrelin is a 28-amino-acid endogenous ligand of the growth hormone secretagogue receptor (GHS-R) and is primarily secreted from gastric endocrine cells (8, 26). In addition to its secretagogue action on growth hormone (GH), ghrelin is known to have a strong orexigenic effect (26, 34) and has been reported to enhance gastrointestinal motility (12, 20, 30). In vitro, ghrelin contracts muscle strips of rat forestomach and antrum (7, 15). Ghrelin administration induces dose-dependent phase III-like contractions in the antrum and increases the motility index (MI) in rodents (11, 45, 52) and humans (42). In addition, treatment with ghrelin and GH-releasing peptide-6, which is an agonist of ghrelin receptors, improves delayed gastric emptying in mice administered with cisplatin (28) and diabetic mice with gastroparesis (37). These effects of ghrelin are believed to act on the central nervous system via the afferent vagus nerve and promote gastrointestinal motility through an efferent pathway (1, 9, 30). However, whether impaired ghrelin signaling is involved in gastrointestinal dysmotility in GERD remains unknown.

Rikkunshito is a Kampo medicine (traditional Japanese medicine) that is widely prescribed to patients with various gastrointestinal symptoms. Clinically, it has been reported that rikkunshito improves gastrointestinal complaints related to functional dyspepsia (47, 51), and its efficacy has been proven in a multicenter double-blind study (17). Recently, it has also been reported that rikkunshito leads to a fundamental and clinical reduction in esophageal acid reflux and related symptoms in patients with GERD (21–23, 32). In addition, we and others have found that rikkunshito promotes ghrelin secretion in the stomach (13, 44) and increases ghrelin receptor sensitivity (14).

We hypothesized that impaired ghrelin signaling is involved in gastrointestinal dysmotility in GERD. To examine this hypothesis, we investigated endogenous ghrelin levels and the effects of exogenously administered ghrelin in an experimental model of GERD. Furthermore, we also examined whether rikkunshito improves ghrelin signaling in this GERD model.
MATERIALS AND METHODS

All experiments were performed in accordance with the protocols approved by the Animal Experiments Review Board of Tsumura (approved protocol No. 09-22, 10-047, 11-010).

Animals. Eight-week-old male Wistar rats were obtained from CLEA Japan (Tokyo, Japan) and maintained under stable conditions of temperature and humidity and a 12-h:12-h dark:light cycle (7 AM to 7 PM). Rats had access to food and water ad libitum and were housed in groups of four or five. All animal experiments were performed between 9 AM and 6 PM. To avoid the influence of diurnal variations, feeding was stopped 24 h before the experiment, and blood and tissue samples were collected between 1 PM and 4 PM. During intravenous administration through the tail vein, we held the conscious rat lightly with a rat holder and warmed the tail in hot water. The test substance was administered slowly using 26-gauge needles to cause less stress. This procedure was concluded within 1 min.

Chemicals. Rat ghrelin was obtained from the Peptide Institute (Osaka, Japan) and was dissolved in 0.9% sterilized physiological saline (Otsuka Pharmaceutical, Tokyo, Japan). Cisapride (an agonist of the 5-HT4 receptor) was obtained from Sigma-Aldrich (St. Louis, MO) and was suspended in 0.1% carboxymethyl cellulose (Maruiihi Pharmaceutical, Osaka, Japan). Rikkunshito (Tsumura, Tokyo, Japan) was made from a hot-water extract of a mixture of eight varieties of crude drugs: sojutsu (Atractylodis lanceae rhizoma), ninjin (Ginseng radix), hange (Pinelliae tuber), bakuchu (Hoelen), taiso (Zizyphi fructus), chinpi (Aurantii nobilis percarpium), kanzo (Glycyrrhizae radix), and shokyo (Zingiberis rhizoma), which was then spray dried. Rikkunshito was suspended in distilled water (DW) at a concentration of 1.2% wt/vol and ingested ad libitum by GERD rats for 10 days (32). The approximate daily intake of rikkunshito was 1,000 mg/kg.

Preparation of the GERD model. GERD was surgically induced in rats according to the method of Omura et al. (36, 40). In brief, rats were first anesthetized with ether after 24 h of fasting, and then laparotomy was performed at the midline. The stomach and duodenum were exposed extracorporeally, and a transitional (bound-ary) section from the forestomach to the glandular stomach was ligated using 1–0 silk thread (Natsume Seisakusho, Tokyo, Japan). The duodenal side of the pylorus was covered with a 2-mm-wide 18-Fr nelaton catheter (Terumo, Tokyo, Japan) that was sutured and fixed to the surface of the duodenal serous membrane using a 5–0 nylon yarn. The stomach and duodenum were returned into the abdominal cavity, which was then closed. Sham-operated rats were first laparatomized to expose their stomach and duodenum for about 1 min, after which their abdominal cavities were closed. After surgery, rats were fasted for a further 24 h (resulting in a total fasting of 48 h).

Sham-operated and GERD rats were given food ad libitum from the day after surgery, and the pair-fed sham-operated rats were given the same amount of food as that consumed by GERD rats on the day before to examine the influence of nutritional decline.

Measurement of gastric emptying. This experiment was performed 10 days after GERD induction (10 AM to 3 PM). Gastric emptying was assessed according to a previously reported method (33). Sham-operated and GERD rats were orally administered 0.5 ml of the test meal prepared by mixing standard powdered chow (Oriental Yeast, Tokyo, Japan) and glass beads (0.2-mm diameter, BZ-02; AS ONE, Osaka, Japan) using Teflon tubes (internal diameter, 1.68 mm) connected to a 2.5-ml syringe with a 10-Fr nelaton catheter 24 h after surgery, laparotomy was performed at the midline. Thereafter, a strain gauge force transducer (F-08IS; Star Medical, Tokyo, Japan) was sutured to the surface of the antral and duodenal serous membrane to measure circular muscle contractions. In addition, a catheter filled with heparin in physiological saline was fixed to the jugular vein. After surgery, rats were housed separately and fasted for a further 24 h (resulting in a total fasting of 48 h).

Analysis of gastroduodenal motility. After GERD induction, rikkunshito (1,000 mg/kg) was administered to rats ad libitum for 10 days. On day 10 (9 AM to 3 PM), after 24 h of fasting, gastroduodenal motility was measured in conscious, freely moving rats. The control group, sham-operated and GERD rats were given DW only. The transducer was connected to a preamplifier (FS-04M; Star Medical) via a bridge box (FB-01; Star Medical). Data were recorded using an MP150 (Biopac Systems, Aero Camino Goleta, CA) and analyzed using AcqKnowledge (Biopac Systems). To observe phase III-like contractions, gastroduodenal motility was monitored for 2–3 h.

To examine the effects of exogenous ghrelin on gastroduodenal motility in sham-operated and GERD rats, rat ghrelin (3 nmol/rat) or saline was injected intravenously via the tail vein and food intake was measured 1 h later. Food intake was calculated as the difference between the preprandial and postprandial weight of the food. The experiment was conducted between 10 AM and 12 PM under fed conditions 10 days after GERD induction.

Fixation of a strain gauge force transducer. After 24 h of fasting, rats were anesthetized by intraperitoneal injection of pentobarbital sodium (Kyoritsu Seiyaku, Tokyo, Japan). During GERD-inducing surgery, laparotomy was performed at the midline. Thereafter, a strain gauge force transducer (F-08IS; Star Medical, Tokyo, Japan) was sutured to the surface of the antral and duodenal serous membrane to measure circular muscle contractions. In addition, a catheter filled with heparin in physiological saline was fixed to the jugular vein. After surgery, rats were housed separately and fasted for a further 24 h (resulting in a total fasting of 48 h).

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To examine the effects of exogenous ghrelin on gastroduodenal motility in sham-operated and GERD rats, rat ghrelin (3 nmol/rat) or saline was administrated through the jugular vein catheter. The effects of the drug were evaluated by frequency of phase III-like contractions and changes in MI before and after its administration.

Phase III-like contractions were defined as the existence of at least three potent contractions (>1/3 of maximum contraction magnitude) of short duration (<5 min). MI was defined as the mean of the area under the contractility recording curve per minute. The definition was slightly modified from a previous report (46).

Measurement of plasma ghrelin and GH levels. Ten days after GERD induction (1 PM to 4 PM), blood was taken after decapitation in a tube containing aprotinin (Wako Pure Chemical Industries, Osaka, Japan) and EDTA-2Na (Nacalai Tesque, Kumamoto, Japan) to obtain plasma samples as reported previously (44). Blood samples were immediately centrifuged at 4°C. Plasma was acidified with 1 mol/l HCl (1/10 volume) and stored at –80°C until measurement. Plasma ghrelin levels were measured using the Active- andDesacyl-Ghrelin ELISA Kit (Mitsubishi Chemical Medience, Tokyo, Japan). Plasma GH levels were measured using the Mouse GH ELISA Kit (Millipore, Billerica, MA) using nonacidified plasma samples. To examine the effects of exogenous ghrelin on GH release.
in sham-operated and GERD rats, rat ghrelin (3 nmol/rat) or saline was injected intravenously via the tail vein, and plasma GH levels were measured at 0, 10, 30, and 60 min.

**Immunohistochemical studies.** The stomachs of 24-h-fasted rats were excised 10 days after GERD induction (1 PM to 4 PM). A part of the gastric body was fixed for 2 days in a 10% formalin/phosphoric acid buffer solution. A 1.5-cm section was excised along the proximal to distal stomach, paraffin embedded, and immunohistochemically stained. Methanol containing 3% hydrogen peroxide was used to inhibit endogenous peroxidase activity. After wash with PBS, paraffin sections were incubated with polyclonal anti-ghrelin antibody (Trans Genic, Kumamoto, Japan) diluted to 1:500 for 90 min. The sections were then washed with PBS and incubated with the Immunohistochemical Staining Kit (MAX-PO; Nichirei Biosciences, Tokyo, Japan) for 20 min, washed again with PBS, and then incubated with a 3,3'-diaminobenzidine solution. The total number of ghrelin-positive cells in this section of the gastric body was counted and expressed as the number of ghrelin-positive cells per millimeter of mucosa in longitudinal sections.

**RNA extraction, reverse transcription, and qPCR analysis.** Gastric body and hypothalamus were excised 10 days after GERD induction (1 PM to 4 PM) and placed in a tube on dry ice for freezing. Once completely frozen, tubes were stored at −80°C. The tissue was homogenized, and total RNA was extracted using the RNeasy Universal Tissue Kit (Qiagen, Valencia, CA). Total RNA from each sample was diluted to 100 ng/μl, allowed to react for 5 min at 70°C, and finally cooled on ice. An aliquot of 1 μg of total RNA was reverse transcribed using the TaqMan Reverse Transcription Reagents (Applied Biosystems, Foster City, CA) according to the manufacturer’s protocol. qPCR analysis was performed with the PRISM 7900HT Sequence Detection System (Applied Biosystems) using the TaqMan Universal PCR Master Mix (Applied Biosystems). To compensate for the differences in the amount of total RNA added to each reaction, mRNA expression was normalized with β-actin as an endogenous control and expressed by the Δ threshold (ΔCT) value: ΔCT = 2^{-ΔA} - B0, where Δ is the number of cycles that reached the β-actin gene threshold and B is the number of cycles that reached the target gene threshold. The set of oligonucleotide primers and fluorescent probes used in the TaqMan quantitative PCR were provide by Applied Biosystems: cytoplasmic β-actin: Rn00667869_m1; preproghrelin: Rn00572319_m1; GHS-R: Rn00821417_m1; neuropeptide Y (NPY): Rn00561681_m1; and agouti-related protein (AgRP): Rn01431703_g1.

**Statistical analysis.** Statistical significance was examined using the Student’s t-test or Aspin-Welch t-test after the F test or by Dunnett’s analysis after one-way ANOVA. Data are expressed as the means ± SE of each group, and P < 0.05 was considered to be significant.

**RESULTS**

Gastric emptying was delayed and food intake was decreased in GERD rats. The body weight of GERD rats after 24 h of fasting decreased significantly compared with that of sham-operated rats (sham-operated, 247.3 ± 2.0 g vs. GERD, 202.4 ± 4.7 g; P < 0.001). Gastric emptying in vehicle-treated GERD rats was significantly delayed compared with that in vehicle-treated sham-operated rats 2 h after test meal administration (Fig. 1A). The prokinetic agent cisapride (20 mg/kg) significantly accelerated gastric emptying in sham-operated rats. In addition, cisapride improved the delayed gastric emptying in GERD rats. Twenty-four-hour cumulative food intake of GERD rats was significantly decreased compared with sham-operated rats (Fig. 1B).

Interdigestive antral motility was decreased in GERD rats. Contractions were observed in the antrum and duodenum at regular intervals in sham-operated and GERD rats (Fig. 2, A and B). Although the frequency of phase III-like contractions in the antrum was significantly decreased in GERD rats compared with that in sham-operated rats (Fig. 2C). However, no difference in phase III-like contractions was observed in the duodenum between both groups (Fig. 2D).

Plasma ghrelin levels increased and the number of gastric ghrelin-positive cells decreased in GERD rats. Plasma acyl and desacyl ghrelin levels significantly increased in GERD rats compared with those in sham-operated rats (Fig. 3, A and B). We examined the influence of nutritional decline on plasma ghrelin levels. Although the body weight was approximately the same (P = 0.19) between GERD (202.4 ± 4.7 g) and pair-fed sham-operated rats (209.6 ± 2.1 g), plasma ghrelin levels in GERD rats (acyl ghrelin, 133.8 ± 9.0 fmol/ml; desacyl ghrelin, 709.0 ± 49.5 fmol/ml) increased significantly compared with those in pair-fed sham-operated rats (acyl ghrelin, 86.5 ± 7.7 fmol/ml; desacyl ghrelin, 423.5 ± 40.7 fmol/ml, P < 0.001). No difference in plasma GH levels was observed (sham-operated, 2.4 ± 1.2 vs. GERD, 4.7 ± 2.8 ng/ml; P = 0.54). The number of ghrelin-positive
cells in the gastric body in GERD rats significantly decreased compared with that in sham-operated rats (Fig. 3, C–E).

Expression of orexigenic genes in the hypothalamus increased in GERD rats. The expression of preproghrelin and GHS-R genes in the stomach of GERD rats did not differ from that of sham-operated rats (Fig. 4, A and B). In contrast, the expression of NPY and AgRP genes, which are known to be orexigenic in the hypothalamus, significantly increased in GERD rats compared with that in sham-operated rats (Fig. 4, C and D).

Ghrelin administration did not improve gastric emptying, food intake, and GH release in GERD rats. Ghrelin administration significantly accelerated gastric emptying in sham-operated rats 1 h after test meal administration in a dose-dependent manner (Fig. 5A). Gastric emptying in GERD rats 1 h after test meal administration showed a trend toward acceleration following administration of 3 or 10 nmol/rat ghrelin. Food intake in sham-operated rats after ghrelin administration (3 nmol/rat) was significantly higher than that after saline administration, but no significant difference was observed between GERD rats administered saline and ghrelin (3 nmol/rat) (Fig. 5B). In addition, plasma GH levels reached a peak 10 min after ghrelin administration (3 nmol/rat) in both sham-operated and GERD rats, but the peak level of GH in GERD rats was significantly lower than that in sham-operated rats (Fig. 5C).

Ghrelin administration did not affect gastroduodenal motility in GERD rats. Figure 6, A and B, shows gastroduodenal motility patterns in sham-operated and GERD rats administered ghrelin (3 nmol/rat). In sham-operated rats, the change in MI in both the antrum and duodenum significantly increased after ghrelin administration (Fig. 6, C and D). However, in GERD rats, the change in MI in the antrum and duodenum showed no difference after ghrelin and saline administration. In the antrum, the frequency of phase III-like contractions did not differ between sham-operated and GERD rats (Fig. 6E). In the duodenum, the frequency of phase III-like contractions significantly increased after ghrelin administration in sham-operated rats, but not in GERD rats (Fig. 6F).

Fig. 2. Fasted gastroduodenal motility in sham-operated and GERD rats. A and B: antral and duodenal motor patterns detected by a strain gauge force transducer in sham-operated and GERD rats, respectively. C and D: frequency of phase III-like contractions in the antrum and duodenum. Results are expressed as means ± SE (n = 6–8).
Rikkunshito administration restored impaired ghrelin response on gastroduodenal motility in GERD rats. Figure 7, A and B show gastroduodenal motility patterns in GERD rats administered DW or rikkunshito for 10 days after GERD induction and intravenous injection of ghrelin via the jugular vein catheter (3 nmol/rat). Before ghrelin administration, the frequency of phase III-like contractions in the antrum of GERD rats administered rikkunshito significantly increased compared with that in the antrum of GERD rats administered DW (DW administration, 1.6 ± 0.1 vs. rikkunshito administration, 2.1 ± 0.2 count/h; \( P = 0.035 \)). There was no difference in the frequency in the duodenum between both groups (DW administration, 5.4 ± 0.5 vs. rikkunshito administration, 5.7 ± 1.0 count/h; \( P = 0.78 \)). After ghrelin administration, there was no change in MI in the antrum and duodenum of GERD rats administered DW. However, a significant increase in the change in MI in the antrum (Fig. 7C) and an increasing trend in the same in the duodenum of GERD rats administered

Fig. 3. Plasma ghrelin levels and number of gastric ghrelin-positive cells in sham-operated and GERD rats. A and B: plasma acyl and desacyl ghrelin levels. C and D: ghrelin-positive cells in the gastric body of sham-operated and GERD rats (scale bars = 100 \( \mu \)m). E: number of ghrelin-positive cells per millimeter of mucosa in longitudinal sections. Results are expressed as means ± SE (n = 5–8).
Similarly, there was a significant increase in the frequency of phase III-like contractions in the antrum (Fig. 7E) and an increasing trend in the same in the duodenum of GERD rats administered rikkunshito ($P = 0.074$ vs. saline administration; Fig. 7F).

The food intake of GERD rats administered rikkunshito did not differ from that of GERD rats administered DW (9 days after GERD induction: DW administration, 19.9 ± 1.2 vs. rikkunshito administration, 17.9 ± 0.9 g/day; $P = 0.24$). Ten days after GERD induction, the effect of ghrelin administration on gastric emptying in GERD rats administered rikkunshito was examined. In GERD rats administered DW, ghrelin administration had no effect on gastric emptying (Fig. 7G). However, ghrelin administration significantly increased gastric emptying compared with saline administration in GERD rats administered rikkunshito.

**DISCUSSION**

In this study, we focused on the association between peripheral ghrelin and GERD. We demonstrated that 1) GERD rats showed decreased gastric emptying, food intake, and antral motility and increased peripheral ghrelin levels and 2) ghrelin administration to GERD rats did not improve decreased gastric emptying, food intake, and antral motility, whereas rikkunshito administration improved decreased antral motility.

Although decreased gastric emptying has already been reported in patients with GERD (3, 31), its mechanism remains unclear. However, treatment with prokinetic agents has been shown to improve several symptoms in patients with GERD (16, 39), and we have recently demonstrated that cisapride improves the motility proximal to the lower esophageal sphincter in GERD rats (40). These findings suggest that gastrointestinal dysmotility is involved in GERD progression. In the present study, cisapride administration had a significant inhibitory effect on delayed gastric emptying in GERD rats, suggesting that motility in the upper gastrointestinal tract in such rats is regulated by 5-HT$_4$ signaling. At present, no study on changes in antral motility of GERD rats has been reported. We found that antral motility significantly decreased in GERD rats; however, no difference was observed in duodenal motility. These results suggest that decreased antral motility is one of the causes of delayed gastric emptying in GERD rats.
Interdigestive contractions in the antrum are regulated in part by peripheral acyl ghrelin (13). Increased circulating acyl ghrelin levels have been shown to induce phase III-like contractions in the antrum. In the present study, plasma acyl and desacyl ghrelin levels significantly increased in GERD rats; however, the number of ghrelin-positive cells in the stomach significantly decreased. Moreover, the expression of the preproghrelin gene in the stomach of GERD rats was not affected. These results suggest that higher plasma ghrelin levels observed in GERD rats is not due to an increase in the synthesis of ghrelin in the stomach but due to enhanced secretion of ghrelin into the circulation from the stomach, resulting in a decrease in stored ghrelin in X/A-like cells, which is consistent with our observation that the number of ghrelin-positive cells decreased in GERD rats.

In rats, serum ghrelin levels increased with fasting and decreased with refeeding and administration of glucose (48). Furthermore, serum or plasma ghrelin levels are inversely proportional to body mass index in humans (19, 35). In this study, the body weight of GERD rats significantly decreased compared with that of sham-operated rats. Therefore, it was suggested that nutritional status may have influenced plasma ghrelin levels. However, the pair-fed sham-operated rats that were controlled in order that they would consume the same amount of food as that consumed by GERD rats showed plasma ghrelin levels similar to those in sham-operated rats. It has been reported that plasma ghrelin levels in rats with food intake restricted by 25% and 50% that were measured 15 days after the initiation of restriction did not differ from rats with ad libitum food intake (27). Therefore, it has been suggested that the increased plasma ghrelin levels in GERD rats may have not resulted solely from reduced food intake.

In the present study, although plasma ghrelin levels increased in GERD rats, decreased phase III-like contractions were observed in the antrum due of delayed gastric emptying and decreased food intake. Ghrelin administration increased gastric emptying, food intake, and the frequency of phase III-like contractions in the duodenum in sham-operated rats but did not improve delayed gastric emptying and decreased food intake in GERD rats. In this study, sham-operated and GERD rats were exogenously administered 3 nmol/rat ghrelin. Gastric emptying in the vehicle group of GERD rats varied significantly. The number of

![Graph](image-url)
Fig. 6. Effects of exogenous ghrelin on fasted gastroduodenal motility in sham-operated and GERD rats. A and B: antral and duodenal motor patterns detected by a strain gauge force transducer in sham-operated and GERD rats. C and D: effects of exogenous ghrelin administration (3 nmol/rat, intravenously) on the change in motility index (MI) in the antrum and duodenum. E and F: effects of exogenous ghrelin administration (3 nmol/rat, intravenously) on the frequency of phase III-like contractions in the antrum and duodenum. Results are expressed as means ± SE (n = 5–10).
animals in each group was seven. Variations in the pathological conditions may have had an effect on gastric emptying. Although GERD rats were administered a higher dosage of ghrelin than sham-operated rats when converted into kilogram of body weight (sham-operated, 12.1 nmol/kg; GERD, 14.8 nmol/kg), no significant improvements were observed in gastric emptying. In addition, although the frequency of phase III-like contractions in the duodenum of GERD rats was similar to that in the duodenum of sham-operated rats, ghrelin administration did not increase their...
It is known that ghrelin transmits signals to the afferent vagus nerve and passes through the central nervous system to induce gastrointestinal contractions through the efferent vagus nerve. In GERD rats, we speculate that the vagus nerves are impaired and may lead to decreased ghrelin response. Therefore, to determine whether the decreased response to ghrelin in GERD rats is due to decreased functionality of the ghrelin receptors, we investigated the effects of concomitant administration of rikkunshito and ghrelin on the antral and duodenal motility and gastric emptying. Ghrelin receptors are G protein-coupled receptors and mediate Ca\(^{2+}\) influx through the phospholipase C/protein kinase C/inositol triphosphate pathway (18) and adenylate cyclase/cyclic AMP/protein kinase A pathway (25). In AgRP/NPY neurons in the arcuate nucleus, ghrelin signaling is blocked by the action of phosphodiesterase type III and phosphatidylinositol 3-kinase, which decrease cyclic AMP (24). Rikkunshito contains heptamethoxyflavone and nobiletin, which have been shown to inhibit phosphodiesterase type III activity in vitro (43). From those findings, it was speculated that the increase in the frequency of spontaneous phase III-like contraction in the antrum and ghrelin response in GERD rats administered rikkunshito was due to the stimulation of ghrelin signaling by protein kinase A because of the prevention of cyclic AMP decomposition by rikkunshito, which inhibits phosphodiesterase type III activity. Therefore, it is possible that factors that decrease cyclic AMP may increase in the vagal nerve endings with ghrelin receptors in our GERD model.

In conclusion, impaired ghrelin signaling is involved in gastrointestinal dysmotility in GERD rats. Moreover, rikkunshito improves gastrointestinal motility by enhancing the decreased response to ghrelin. Improvement of ghrelin signaling may provide a new therapeutic approach for GERD.

**REFERENCES**


