Nesfatin-1 inhibits gastric acid secretion via a central vagal mechanism in rats

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Nesfatin-1 inhibits gastric acid secretion via a central vagal mechanism in rats. Am J Physiol Gastrointest Liver Physiol 303: G570–G577, 2012. First published June 21, 2012; doi:10.1152/ajpgi.00178.2012.—Nesfatin-1, a novel hypothalamic peptide, inhibits nocturnal feeding behavior and gastrointestinal motility in rodents. The effects of nesfatin-1 on gastrointestinal secretory function, including gastric acid production, have not been evaluated. Nesfatin-1 was injected into the fourth intracerebral ventricle (4V) of chronically cannulated rats to identify a nesfatin dose sufficient to inhibit food intake. Nesfatin-1 (2 μg) inhibited dark-phase food intake, in a dose-dependent fashion, for >3 h. Gastric acid production was evaluated in urethane-anesthetized rats. Nesfatin-1 (2 μg) was introduced via the 4V following endocrine stimulation of gastric acid secretion by pentagastrin (2 μg·kg−1·h−1 iv), vagal stimulation with 2-deoxy-D-glucose (200 mg/kg sc), or no stimulus. Gastric secretions were collected via gastric cannula and neutralized by titration to determine acid content. Nesfatin-1 did not affect basal and pentagastri-nin-stimulated gastric acid secretion, whereas 2-deoxy-D-glucose-stimulated gastric acid production was inhibited by nesfatin-1 in a dose-dependent manner. c-Fos immunofluorescence in brain sections was used to evaluate in vivo neuronal activation by nesfatin-1 administered via the 4V. Nesfatin-1 caused activation of efferent vagal neurons, as evidenced by a 16-fold increase in the mean number of c-Fos-positive neurons in the dorsal motor nucleus of the vagus (DMNV) in nesfatin-1-treated animals vs. controls (P < 0.01). Finally, nesfatin-1-induced Ca2+ signaling was evaluated in primary cultured DMNV neurons from neonatal rats. Nesfatin-1 caused dose-dependent Ca2+ increments in 95% of cultured DMNV neurons. These studies demonstrate that central administration of nesfatin-1, at doses sufficient to inhibit food intake, results in inhibition of vagally stimulated secretion of gastric acid. Nesfatin-1 activates DMNV efferent vagal neurons in vivo and triggers Ca2+ signaling in cultured DMNV neurons.

Nesfatin-1, an 82-amino acid peptide derived from the parent peptide nucleobindin-2, was first identified as a satiety molecule in the hypothalamus by Oh et al. (33). Inhibition of rodent nocturnal feeding behavior by nesfatin-1 administered into the intracranial ventricles has been consistently observed (18, 25, 34, 40, 42). Most centrally acting peptides that influence food intake also regulate the processes of digestion, including gastrointestinal motility and secretory function (45). In rodents, nesfatin-1 inhibits gastroduodenal motility and gastric emptying (3, 41). We postulate that nesfatin-1 may also affect secretion of gastric acid.

Gastric acid secretion is regulated by neural, hormonal, and paracrine pathways (38, 39). Gastrin, a hormone released from antral G cells, binds to gastrin receptors on parietal cells and also stimulates cholecystokinin-2 receptors on gastric enterochromaffin cells, resulting in histamine release. In paracrine fashion, histamine binds its receptor on the gastric parietal cell, stimulating the production of hydrochloric acid (23). The actions of gastrin constitute a major peripheral mechanism for gastric acid production. Central stimulation of gastric acid secretion is mediated by the vagus nerve. Gastric acid secretion can be stimulated experimentally by 2-deoxy-D-glucose (2-DG), a stable glucose analog that cannot undergo further glycolysis. Systemic administration of 2-DG initiates a state of simulated cytoglycopenia, leading to altered hypothalamic input to the dorsal-vagal complex (DVC) in the brain stem and resultant activation of bilateral vagus nerves (10, 19, 30). 2-DG has been widely used to study gastric acid secretion mediated by the vagus nerve (19, 32, 36). Using gastrin and 2-DG to evaluate peripheral and central mechanisms of stimulated acid production, we have explored the effects of centrally administered nesfatin-1 on gastric acid secretion.

MATERIALS AND METHODS

Animals. Adult male Sprague-Dawley rats (240–280 g body wt) were housed in a 12:12-h light-dark cycle and had access to food and water ad libitum.

Chemicals and solutions. Nesfatin-1 was purchased from Phoenix Pharmaceuticals (Burlingame, CA); Neurobasal A medium, B27 supplement, penicillin-streptomycin, L-glutamine, trypsin type I, β-FGF, fura 2-AM, goat serum, and ProLong antifade with 4,6-diamidino-2-phenylindole from Life Technologies, formerly Invitrogen (Carlsbad, CA); Triton X-100 from Thermo Fisher (Waltham, MA); 5-thio-D-glucose from MP Biomedicals (Solon, OH); ketamine, poly-L-lysine, pentagastrin, 2-DG, NiCl2, verapamil, α-conotoxin GVIA, and mibe-fradil dihydrochloride from Sigma-Aldrich (St. Louis, MO); xylazine from Lloyd (Shenandoah, IA); optimal cutting temperature (OCT) compound from Sakura Finetek USA (Torrance, CA); and rabbit anti-c-Fos antibody from Calbiochem (San Diego, CA), formerly Oncogene (Cambridge, MA). FITC-conjugated goat anti-rabbit IgG (Jackson ImmunoResearch, West Grove, PA) was used as secondary antibody. Intracerebroventricular cannulas were purchased from and cut to 7.5 mm by Plastics One (Roanoke, VA).

Fourth intracerebral ventricle cannula placement. Male Sprague-Dawley rats (240–280 g body wt) were anesthetized with ketamine (90 mg/kg ip) and xylazine (10 mg/kg ip) and placed on a stereotaxic instrument (Stoelting, Wood Dale, IL). A 22-gauge single-lumen cannula (Plastics One) was inserted into the 4V with the tips of the guide cannulae aimed at the following coordinates: 1.0 mm posterior, 1.0 mm lateral to the midline, and 3.5 mm ventral to the dura mater.
cannula was implanted into the fourth intracerebral ventricle (4V) according to coordinates published by Paxinos and Watson (35): midline, 12.8 mm caudal to bregma, 7.5 mm deep, with the incisor bar set 3.3 mm below the interaural line. The cannula was secured with cranialfacial cement. A sterile internal cannula was placed to maintain patency and prevent infection. On postprocedure day 3, cannula position was tested by 4V injection of 210 μg of 5-thio-d-glucose in 3 μL of PBS. A ≥100% rise in blood glucose in response to 5-thio-d-glucose was considered confirmation of correct cannula position. Experiments were conducted beginning on postprocedure day 7, allowing adequate recovery from the procedure and confirmation of cannula position.

Dark-phase food intake. To establish a nesfatin-1 dose sufficient to inhibit food intake for use in the remainder of the studies, chronically cannulated rats were injected via the 4V with nesfatin-1 in 2 μL of PBS at 6 PM, the onset of the dark phase. The nesfatin-1 dose was varied from 0 to 2 μg on the basis of previous dose-response studies of third ventricular nesfatin injection (33). Rats were singly caged in familiar conditions with free access to food and water. Food intake was monitored for up to 24 h and calculated as grams of food consumed per 300 g body wt.

Gastric acid output. The effects of nesfatin-1 on in vivo gastric acid secretion were studied using an acute gastric fistula method previously described (29). Rats with chronically implanted 4V cannulas were fasted for 16 h and then anesthetized with urethane (1.5 g/kg ip). Venous access was achieved via the jugular vein. Through a small midline abdominal incision, the stomach and duodenum were exposed. The esophagus was ligated at the gastroesophageal junction, with care taken to preserve the vagal trunks. A double-lumen cannula was inserted into the stomach via the duodenum before ligation of the pylorus. The stomach was flushed with 50–100 ml of saline to remove any solid content. After collection of gastric outputs in 5 ml of perfusate every 15 min for 30 min to establish baseline acid secretion, test substances were administered via the 4V, intravenously, or subcutaneously. The gastric lumen was subsequently rinsed with 5 ml of saline every 15 min for 120 min. Acid production was measured by titration of gastric rinse solution with 0.1 N NaOH to neutrality and expressed as microequivalents of HCl per 15 min.

Brain sections and c-Fos immunofluorescence. To investigate the activation of brain stem vagal neurons by nesfatin-1, chronically cannulated and freely fed rats were treated with 4V nesfatin-1 (0.5 μg in 2 μL of PBS, n = 3) or vehicle (2 μL of PBS, n = 3) during the dark phase. After 90 min, the rats were deeply anesthetized and euthanized prior to transcardial perfusion with PBS and 4% paraformaldehyde. Brains were removed, postfixed in 4% paraformaldehyde, transferred to a solution of 20% sucrose in PBS for 5 days, and finally embedded with 20% sucrose-OCT compound (2:1) on dry ice. A Leica cryostat was used to section tissue blocks in 40-μm slices through the dorsal motor nucleus of the vagus (DMNV). Each section was stored in 300 μL of PBS with 0.02% sodium azide in each well of the 24-well plate.

Free-floating slices were first rinsed with PBS with 0.5% Triton for 15 min and incubated with a blocking buffer (10% normal goat serum, 3% BSA, 0.4% Triton X-100, and 1% glycine, pH 7.4) for 1 h at room temperature. Sections were then treated with rabbit anti-c-Fos antibody (1:10,000 dilution) at 4°C for 40 h, washed with PBS, and treated with FITC-conjugated goat anti-rabbit IgG (1:400 dilution) for 1 h at room temperature in the dark. After incubation with the secondary antibody, sections were washed in PBS for 15 min and incubated with 4,6-diamidino-2-phenylindole (1:6,000 dilution) for 3 min. Finally, sections were dehydrated to slides, dried for 1 h, mounted with ProLong Gold antifade reagent, and covered with coverslips. All antibodies were diluted in 5% goat serum blocking buffer. Slides were viewed using a fluorescence microscope (Nikon Eclipse Ti-U). The average numbers of c-Fos-positive (green) neurons per section per brain nucleus were assessed.

In vitro culture of DMNV neurons. Neurons of the DMNV were harvested from neonatal Sprague-Dawley rats, as previously described (49). After the animals were euthanized, the brain was rapidly removed and placed into a chilled electrolyte solution. The brain stem medulla was then sectioned transversely into a 1-mm slice. The DMNV was identified under a dissecting microscope as the area immediately ventral to the nucleus of the solitary tract (NTS) and dorsal to the XII nucleus. DMNV tissue was excised and digested with trypsin type I (0.4 mg/ml) at 30°C for 20 min. Tissue was then gently dissociated by pipette trituration and plated onto polystyrene-coated 25-mm chamber slides in 35-mm culture dishes. Neurons were maintained in serum-free culture medium containing Neurobasal A medium, 2% B27 supplement, 2 mM glutamine, 1% penicillin, 1% streptomycin, and 5 ng/ml β-FGF. One-half of the medium was replaced on day 1 and every 3rd day thereafter. Cells were utilized between day 5 and day 10.

Intracellular Ca²⁺ concentration measurements. Intracellular Ca²⁺ concentration ([Ca²⁺]i) was measured using dual-wavelength fura 2-AM microfluorometry and imaging, as previously reported (2, 49). DMNV neurons plated on coverslips were loaded with 1 μM fura 2-AM and incubated for 60 min at 37°C. Coverslips were placed into a perfusion chamber mounted on the stage of an inverted fluorescence microscope (Nikon) and perfused with HBSS containing CaCl₂. Superfusion of buffer and reagents was kept constant at 1 ml/min at 30°C. For Ca²⁺-free conditions, HBSS without CaCl₂ was used. After 60 s of exposure to buffer, cells were perfused with one of the following reagents: nesfatin-1, NiCl₂, verapamil, ω-conotoxin GVIA, and mibebradil dihydrochloride. At the conclusion of each experiment, cells were exposed to 55 mM KCl; cells that were unresponsive to KCl were excluded. [Ca²⁺]i was measured using a Nikon inverted fluorescence microscope (Mager Scientific, Dexter, MI) with ×40 oil-immersion objective and TILLvision digital imaging system (TILL Photonics, Pleasanton, CA). [Ca²⁺]i was determined for each individual cell from the ratio of fluorescence intensity at 550 nm following excitation at 340 nm to that following excitation at 380 nm. Measurements were corrected for background fluorescence intensity. The change in fluorescence ratio (ΔF) in response to a stimulus was calculated for each cell and divided by that cell’s basal fluorescence ratio (F₀) to yield percent change in intracellular Ca²⁺ compared with baseline (ΔF/F₀). Cells with ΔF/F₀ >10% in response to a given stimulus were considered responsive.

Data analysis. Data were analyzed with GraphPad Prism software (GraphPad Software, San Diego, CA) using ANOVA or Student’s t-test as appropriate. Values are means ± SE. Significance was accepted at P < 0.05.
gastric acid output. Peak stimulated acid production was reduced by 61% and 26% in response to 2 and 0.5 μg of nesfatin-1, respectively (Fig. 1 C).

Effect of nesfatin-1 on c-Fos expression in DMNV neurons. Given the ability of nesfatin-1 to inhibit vagally mediated gastric acid production, c-Fos immunofluorescence was used to determine whether 4V nesfatin-1 injection caused activation of vagal neurons. In controls treated with PBS (2 μl) via the 4V (n = 3), levels of c-Fos positivity in DMNV neurons were low (Fig. 2A). In animals treated with 0.5 μg of nesfatin-1 via the 4V (n = 3), we observed a 16-fold increase in the mean c-Fos-positive neurons per section per brain nucleus were significantly increased by nesfatin-1 exposure: *P < 0.01.
number of c-Fos-positive neurons per DMNV compared with controls \((P < 0.01; \text{Fig. 2, B and C})\). Neurons in the adjacent NTS did not exhibit changes in c-Fos positivity associated with nesfatin-1 injection. These observations suggest that 4V administration of nesfatin-1 results in stimulation of a population of DMNV neurons.

**Nesfatin-1 induces \([\text{Ca}^{2+}]_i\) increments in DMNV neurons.** Nesfatin-1-induced \([\text{Ca}^{2+}]_i\) transients were evaluated in primary cultured DMNV neurons to confirm DMNV neuron response to nesfatin-1. Superfusion of fura 2-loaded DMNV neurons with \(10^{-6}\) M nesfatin-1 for 300 s induced \([\text{Ca}^{2+}]_i\) increments in 95% of cells \((n = 102)\). The increment in \([\text{Ca}^{2+}]_i\) reached a peak \(\Delta F/F_0 (102 \pm 6\%)\) within 30 s, maintained a plateau for the duration of nesfatin-1 administration, and subsequently declined to baseline (Fig. 3A). All neurons responded simultaneously, indicating that the effects of nesfatin-1 were direct, rather than the result of propagation through gap junctions or synapses. DMNV neurons responded to nesfatin-1 in a dose-dependent fashion over a range of concentrations from \(10^{-10}\) to \(10^{-6}\) M \((n = 443, P < 0.05)\). Increasing doses of nesfatin-1 resulted in \([\text{Ca}^{2+}]_i\) increments of greater magnitude (Fig. 3B), as well as a greater proportion of neurons reaching the response threshold (Fig. 3C). For the remaining experiments, \(10^{-7}\) M was chosen as a working concentration for nesfatin-1. Repetitive exposure to nesfatin-1 produced progressive decrements in peak \(\Delta F/F_0\) responses (Fig. 4A). The average \(\Delta F/F_0\) values for the three exposures were 45 ± 3%, 40 ± 2%, and 26 ± 2%, respectively \((n = 76, P < 0.05\) for peak 3 compared with peak 1; Fig. 4B).

**Mechanism of nesfatin-1-induced \([\text{Ca}^{2+}]_i\) increments in DMNV neurons.** The mechanism by which nesfatin-1 causes increased cytoplasmic \([\text{Ca}^{2+}]_i\) concentrations in DMNV neurons was evaluated by nesfatin-1 exposure under a variety of conditions. First, the involvement of extracellular \([\text{Ca}^{2+}]_i\) in nesfatin-induced \([\text{Ca}^{2+}]_i\) transients was examined. In the absence of extracellular \([\text{Ca}^{2+}]_i\), nesfatin-1 failed to increase \([\text{Ca}^{2+}]_i\) in any of the cells (Fig. 5). This observation suggests that nesfatin-1-induced \([\text{Ca}^{2+}]_i\) transients are the result of \([\text{Ca}^{2+}]_i\) influx across the plasma membrane.

To determine which subtype of \([\text{Ca}^{2+}]_i\) channel is involved in influx of \([\text{Ca}^{2+}]_i\), DMNV neurons were exposed to nesfatin-1 in
the presence of various Ca^2+ channel blockers. NiCl₂ (3 mM), a nonspecific Ca^2+ channel blocker, abolished nesfatin-1-induced intracellular Ca^2+ transients in all neurons (n = 93; Fig. 6). Antagonists specific to the three major Ca^2+ channel subtypes were then utilized. Superfusion with the L-type Ca^2+ channel blocker verapamil (1 μM) or the N-type Ca^2+ channel blocker ω-conotoxin GVIA (1 μM) did not significantly alter nesfatin-1-induced peak ΔF/F₀ (n = 73 and 86, respectively; Fig. 7A). Selective blockade of T-type Ca^2+ channels with 1, 10, and 20 μM mibebradil caused dose-dependent decreases in average peak ΔF/F₀ (33.5%, 18.0%, and 10.2%, respectively; Fig. 7, B–D). These observations suggest that nesfatin-1 increases cytoplasmic Ca^2+ concentrations in DMNV neurons by influx of extracellular Ca^2+, at least in part via a T-type Ca^2+ channel.

**DISCUSSION**

The present study represents the first report that nesfatin-1 inhibits gastric acid secretion stimulated by a central vagal mechanism. This conclusion is supported by the following observations: 1) intracerebroventricular nesfatin-1 decreased 2-DG-stimulated gastric acid output in a dose-dependent manner in urethane-anesthetized rats, with no effect on basal or pentagastrin-stimulated gastric acid production; 2) in conscious rats, 4V nesfatin-1 activates DMNV neurons; and 3) in primary cultured DMNV neurons, nesfatin-1 causes dose-dependent increments in cytosolic Ca^2+ that can be abolished by the absence of extracellular Ca^2+ or blockade of T-type Ca^2+ channels. Taken together, these observations indicate that nesfatin-1 is able to activate vagal neurons in vitro and in vivo and that central exposure to nesfatin-1 inhibits vagally stimulated gastric acid secretion.

Nesfatin-1 was first described by Oh et al. (33) as a hypothalamic neuropeptide able to reduce dark-phase food intake. Subsequent studies, including this one, provide concordant evidence that nesfatin-1 injected into the brain elicits an anorexigenic response in rodents (3, 18, 26, 40, 41, 48). Nesfatin-1 immunoreactivity has been identified in areas of the brain relevant to feeding behavior, including the paraventricular nucleus, supraoptic nucleus, lateral hypothalamic area, arcuate nucleus, dorsomedial hypothalamic nucleus, periventricular nucleus, Edinger-Westphal nucleus, NTS, and DMNV (7, 15–17, 21, 24). As with other anorexigenic neuropeptides, nesfatin-1 has been demonstrated to affect digestive processes as well as feeding behavior. Centrally administered nesfatin-1 delays gastric emptying in rats (41) and mice (17) and also suppresses gastroduodenal motility (3). This inhibition of gastrointestinal motility by nesfatin-1 is hypothesized to contribute to the induction of satiety.

In this study, we microinjected nesfatin-1 into the 4V to investigate its effects on gastrointestinal secretory function. Our results confirm that nesfatin-1 causes dose-dependent, short-term inhibition of dark-phase food intake. At doses sufficient to inhibit food intake, nesfatin-1 exhibited no appreciable effect on basal or pentagastrin-stimulated acid secretion. In contrast, 2-DG-stimulated acid production was inhibited by nesfatin-1 in a dose-dependent manner. As 2-DG is known to trigger gastric acid secretion by activation of the vagal center (10, 19, 32), our study indicates that nesfatin-1 suppresses vagally stimulated gastric acid production.

The selective inhibition of vagally mediated gastric acid production is rare, but not unique. Nitric oxide donors, for example, have been observed to inhibit vagally stimulated gastric acid secretion without affecting basal or pentagastrin- or histamine-stimulated acid production (4). Nitric oxide is hypothesized to serve as an intermediary in the gastric acid suppression induced by endotoxin, cytokines, or stress (13, 14, 27). Interestingly, while several orexigenic hormones have been implicated in vagal stimulation of gastric acid production, nesfatin is the only anorexigenic hormone confirmed to inhibit vagally mediated gastric acid production (11, 28, 46). Vagovagal circuits relayed through the DVC regulate a variety of gastric functions, including the secretion of gastric acid (8, 9). The DVC is a paired structure located in the dorsal caudal medulla alongside the central canal, which is contiguous with the 4V. It comprises two adjacent and functionally integrated nuclei: the NTS and the DMNV. Afferent vagal fibers from the gastrointestinal tract synapse on interneurons located in the NTS. In turn, these neurons synapse on efferent vagal motor neurons in the DMNV. These preganglionic, efferent vagal...
fibers project to ganglia in the upper gastrointestinal tract (5, 44, 50), with the highest density terminating in the stomach (6). The majority of the associated postganglionic neurons project onto interstitial cells of Cajal, providing parasympathetic control of gastrointestinal motility (9, 44). A minority of these postganglionic neurons directly innervate parietal cells and gastric paracrine cells, resulting in cholinergic stimulation of gastric acid production.

Given our observation that nesfatin-1 administered in close proximity to the DVC inhibited vagally stimulated gastric acid production, we hypothesized that nesfatin-1 might have a direct effect on neurons involved in vagovagal circuits. In our in vivo study, DMNV, but not NTS, neurons were activated by 4V nesfatin-1, as evidenced by c-Fos immunofluorescence. Confirmation that DMNV neurons can be stimulated by nesfatin-1 was obtained through the examination of intracellular Ca\(^{2+}\) signaling in vitro. Nesfatin-1 inhibited the capacity to evoke Ca\(^{2+}\) transients in 95% of primary cultured rat DMNV neurons. Ca\(^{2+}\) ions are critically important to many functions of the nervous system, from neurotransmitter release to intracellular signal transduction. The large gradient between intracellular and extracellular Ca\(^{2+}\) concentration highlights the importance of the mechanisms controlling its influx and efflux (12, 37). Nesfatin-1 has been reported to evoke Ca\(^{2+}\) signaling in isolated vagal afferent neurons via N-type Ca\(^{2+}\) channels (22) and in cultured hypothalamic neurons via L- and P/Q-type Ca\(^{2+}\) channels (7). Our study indicates that nesfatin-1 evokes Ca\(^{2+}\) transients in vagal neurons via modulation of T-type Ca\(^{2+}\) channels (20, 37). The significance of T-type Ca\(^{2+}\) channel activation in vagal neuron response to nesfatin-1 and the ultimate impact of this particular signaling pathway on gastric acid secretion are unknown.

In addition to vagal, endocrine, and paracrine influences, gastric acid secretion is influenced by a variety of factors, including blood pressure and local blood flow (1, 31, 43). Additionally, all common general anesthetics have some impact on gastrointestinal secretory function (31). In these studies, care was taken to curtail the impact of these other factors. Urethane is the anesthetic agent classically utilized in physiology studies of gastric acid secretion. It has depressive effects on gastric acid production, with augmentation of somatostatin secretion (31, 47). While urethane anesthesia does influence gastric acid secretion, the effects are well-characterized in the literature. Do not preclude stimulation of acid secretion with 2-DG, and are assumed to be similar in the treatment and control groups. Moreover, a single intraperitoneal urethane injection can produce >6 h of general anesthesia, thus minimizing fluctuations in blood pressure associated with varying depth of sedation and analgesia. Care was taken to preserve the

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Fig. 7. Dose-dependent inhibition of nesfatin-1 induced Ca\(^{2+}\) transients by mibefradil, but not by verapamil or ω-conotoxin GVIA. DMNV cells were superfused with 10\(^{-7}\) M nesfatin-1 for 30 s and allowed to return to baseline. Cells were pretreated with a Ca\(^{2+}\) channel blocker for 180 s and then superfused for 30 s with nesfatin-1 in the presence of that Ca\(^{2+}\) channel blocker. A: blockade of T-type Ca\(^{2+}\) channels with 1 \(\mu\)M mibefradil diminishes cytoplasmic Ca\(^{2+}\) response to nesfatin-1 (n = 58). *P < 0.05. Blockade of L- and N-type Ca\(^{2+}\) channels with 1 \(\mu\)M verapamil and 1 \(\mu\)M ω-conotoxin GVIA, respectively, did not significantly inhibit DMNV neuron response to nesfatin-1 (n = 73, P = 0.85 for verapamil; n = 86, P = 0.22 for ω-conotoxin GVIA). B: representative trace of a single neuron demonstrating inhibition of nesfatin-1-induced Ca\(^{2+}\) transients in the presence of 10 \(\mu\)M mibefradil. C and D: dose-inhibition relationship. As concentration of mibefradil was increased, average ΔF/F₀ decreased and inhibition of nesfatin-1-induced [Ca\(^{2+}\)] increased. *P < 0.05 vs. lower dose.
neurovascular supply to the gastrointestinal tract and to maintain normothermia.

The studies presented here provide further evidence to support the role of nesfatin-1 as an anorexigenic neuropeptide involved in the central regulation of feeding behavior. Nesfatin-1 has been previously reported to influence digestion via inhibition of gastrointestinal motility; this study represents the first report that nesfatin-1 affects signaling in efferent vagal neurons and inhibits vagally stimulated gastric acid production.

REFERENCES


AJP-Gastrointest Liver Physiol ▪ doi:10.1152/ajpgi.00178.2012 ▪ www.ajpgi.org
37. Ross WN. Understanding calcium waves and sparks in central neurons. 
38. Schubert ML. Gastric secretion. *Curr Opin Gastroenterol* 26: 598–603, 
   2010.
39. Schubert ML, Peura DA. Control of gastric acid secretion in health and 
   Lambrecht NW, Tache Y. Central nesfatin-1 reduces dark-phase food 
   intake and gastric emptying in rats: differential role of corticotropin-
42. Stengel A, Tache Y. Nesfatin-1—an emerging new player in the brain-
43. Takeuchi K, Ohuchi T, Miyake H, Niki S, Okabe S. Effects of nitric 
   oxide synthase inhibitors on duodenal alkaline secretion in anesthetized 
44. Travagli RA, Hermann GE, Browning KN, Rogers RC. Brainstem 
45. Wren AM, Bloom SR. Gut hormones and appetite control. *Gastroenterology* 
46. Yamada H, Takahashi N, Tanno S, Nagamine M, Takakusaki K, 
   Okumura T. A selective orexin-1 receptor antagonist, SB334867, blocks 
   2-DG-induced gastric acid secretion in rats. *Neurosci Lett* 376: 137–142, 
   2005.
   antibody immunoneutralization increases gastric and gastric acid secretion 
48. Yosten GL, Samson WK. The anorexigenic and hypertensive effects of 
   nesfatin-1 are reversed by pretreatment with an oxytocin receptor antag-
49. Zhang W, Hu Y, Newman EA, Mulholland MW. Serum-free culture of 
   rat postnatal neurons derived from the dorsal motor nucleus of the vagus. 
50. Zheng H, Berthoud HR. Functional vagal input to gastric myenteric 
   plexus as assessed by vagal stimulation-induced Fos expression. *Am J 
   Physiol Gastrointest Liver Physiol* 279: G73–G81, 2000.