**Lactobacillus acidophilus** attenuates downregulation of DRA function and expression in inflammatory models

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Singh V, Kumar A, Raheja G, Anbazhagan AN, Priyamvada S, Saksena S, Jhandier MN, Gill RK, Alrefai WA, Borthakur A, Dudeja PK. Lactobacillus acidophilus attenuates downregulation of DRA function and expression in inflammatory models. Am J Physiol Gastrointest Liver Physiol 307: G623–G631, 2014. First published July 24, 2014; doi:10.1152/ajpgi.00104.2014.—Probiotics, including Lactobacilli, are commensal bacteria that have been used in clinical trials and experimental models for the prevention and treatment of diarrhea disorders. Our previous studies have shown that Lactobacillus acidophilus (LA) and its culture supernatant (CS) stimulated Cl−/HCO3− exchange activity, acutely via an increase in the surface levels of downregulated in adenoma (DRA, SLC26A3) and in long-term treatments via increasing its expression involving transcriptional mechanisms. However, the role of LA in modulating DRA activity under inflammatory conditions is not known. Current in vitro studies using human intestinal epithelial Caco-2 cells examined the efficacy of LA or its CS in counteracting the inhibitory effects of interferon-γ (IFN-γ) on Cl−/HCO3− exchange activity. Pretreatment of cells with LA or LA-CS for 1 h followed by coinubation with IFN-γ significantly alleviated the inhibitory effects of IFN-γ on Cl−/HCO3− exchange activity. In the in vivo model of dextran sulfate sodium-induced experimental colitis (3% in drinking water for 7 days) in C57BL/6J mice, administration of live LA (3 × 109 colony-forming units) via oral gavage attenuated colonic inflammation. LA administration also counteracted the colitis-induced decrease in DRA mRNA and protein levels. Efficacy of LA or its secreted soluble factors in alleviating inflammation and inflammation-associated dysregulation of DRA activity could justify their therapeutic potential in inflammatory diarrheal diseases.

probiotics; Caco-2; diarrhea; interferon-γ; dextran sulfate sodium; downregulated in adenoma

THE MOVEMENT OF WATER and ions (especially Na+ and Cl−) across the luminal membrane is an important function of colonic epithelial cells in physiological conditions. This movement is impaired in most forms of diarrheal diseases, including those associated with inflammatory bowel diseases. Two members of the SLC26 gene family, SLC26A3 or downregulated in adenoma (DRA) and SLC26A6 or putative anion transporter-1 (PAT-1), are known to play important roles in the apical Cl−/HCO3− exchange process in intestine. Decreased DRA expression, and its functional impairment have been shown to be associated with various diarrheal disorders (3, 16, 24, 38). Also, DRA mRNA expression has been shown to be significantly decreased by proinflammatory cytokines IL-1β, interferon-γ (IFN-γ), as well as in patients with ulcerative colitis (31, 38).

Probiotics are live microorganisms that when administered in adequate amounts confer health benefits to the host. Probiotics have been previously demonstrated to modulate cytokine production, strengthen mucosal barrier, and reduce inflammation in in vitro and experimental colitis models (20, 22, 30). All of these mechanisms may contribute to their therapeutic efficacy in colitis; however, the specificity of individual probiotic strains and their precise mechanisms of action are poorly understood. In this regard, earlier studies have shown probiotic Lactobacillus-mediated alleviation of inflammation-associated chloride secretion in response to cytokines or enteropathogen infection (30). We have previously shown that the probiotic L. plantarum or their secreted soluble factors counteract TNF-α-induced downregulation of expression and function of SMCT-1, a transporter for short-chain fatty acids, in rat intestinal IEC-6 cells (4). Also, we have previously demonstrated Lactobacillus acidophilus (LA) to stimulate Cl−/HCO3− exchange activity via an increase in the surface levels (6) and expression of the Cl−/HCO3− exchanger DRA in both in vitro and in vivo models (6, 28). However, there are no studies on the modulation of DRA activity by probiotics under inflammatory conditions.

In the current study, we have examined the role of LA in reversing the inflammatory cytokine-induced downregulation of DRA in human intestinal epithelial Caco-2 cells and in a dextran sulfate sodium (DSS) colitis model in mice. For this, we first measured the Cl−/HCO3− exchange activity in Caco-2 cells treated with IFN-γ (30 ng/ml) in the presence or absence of LA or culture supernatant (CS) for 24 h. Interestingly, pretreatment of LA or LA-CS significantly alleviated the inhibitory effects of IFN-γ on Cl−/HCO3− exchange activity. In a mouse model of DSS-induced experimental colitis, oral gavage of live LA showed alleviation of colitis-associated weight loss, diarrheal phenotype as well as downregulation of DRA mRNA and protein levels. In conclusion, our study shows the efficacy of LA and their secreted soluble factors in counteracting the proinflammatory effects of cytokines and justifies the potential therapeutic role of LA or LA-derived bioactive factors in inflammatory disorders of the intestine.

MATERIALS AND METHODS

Materials

Caco-2 cells were procured from ATCC. Radiumucleotide 125I was obtained from PerkinElmer (specific activity 17.0 Ci/mg). RNeasy kits for RNA extraction were obtained from Qiagen (Frederick, MD), and the real-time qRT PCR kit was from Stratagene (La Jolla, CA).
4,4′-Disothiocyanate-stilbene-2,2′-disulfonic acid (DIDS) was procured from Sigma-Aldrich (St. Louis, MO). Human recombinant IFN-γ was obtained from Sigma. Common reagents for SDS-PAGE such as ammonium per sulfate, acrylamide, and bis-acrylamide were from Fisher Scientific (Pittsburgh, PA).

**Cell Culture**

Caco-2 cells were grown in T-75 cm² culture flasks at 37°C in a 5% CO₂ atmosphere in Medium Essential Medium containing 20% FBS, 20 mM HEPES, 100 IU/ml penicillin, and 100 mg/ml streptomycin. Cells (2 × 10⁶) seeded per well in 12-well Transwell inserts were used between passages 25 and 45. Fully differentiated confluent monolayers were used for the experiments (10–12 days postplating).

**Bacterial Culture**

The following Lactobacilli species, with ATCC strain numbers given in parentheses, were used in this study: LA (4357), L. rhamnosus (53103), L. plantarum (14917), and L. casei (393). These species were grown overnight in MRS broth (Difco, Detroit, MI) at 37°C without shaking. Bacteria were spun down by centrifuging at 3,000 rpm for 10 min. For in vitro studies, CS was separated from bacteria by using a 0.22-μm filter, and diluted in cell culture media (1:10) for further use. For treating the cell monolayers, the bacterial pellet was washed with DMEM-F-12 media (Invitrogen Life Technologies, Carlsbad, CA) containing 5 mg/l mannos and resuspended in the same media (6). For in vivo studies, 3 × 10⁹ colony-forming units (CFU) of bacteria in 200 μl PBS, as reported earlier from our laboratory (28), were gavaged per animal for the first 2 days of DSS treatments in LA or LA + DSS groups.

**Treatment of Caco-2 Cells**

Bacterial suspensions in DMEM-F-12 media were diluted to OD₆₀₀ₙ₅₀ = 0.2 in the same media and applied to the apical surface of cell monolayers at a multiplicity of infection of 50 for indicated time periods (6). For Cl⁻/HCO₃⁻ (OH⁻) exchange activity, cells were pretreated for 1 h from the apical side with LA bacterial suspension described above, or with the bacteria-free CS diluted in a ratio of 1:10 in serum-reduced medium (1% FBS) and then coincubated with IFN-γ from the basolateral side (30 ng/ml) for an additional 24 h. Similarly for promoter studies, 24 h posttransfection, cells were pretreated with LA-CS for 1 h followed by coincubation of IFN-γ (30 ng/ml) for an additional 24 h or incubated with IFN-γ (30 ng/ml) or LA-CS for 24 h alone in serum-reduced medium (1% FBS) (34).

**Cl⁻/HCO₃⁻ Exchange Activity**

Cl⁻/HCO₃⁻ exchange activity was assessed as ¹²⁵I⁻ uptake (in place of ³⁶Cl⁻ uptake) carried out in well-differentiated polarized Caco-2 cells as described previously (16, 32). Briefly, after treatment, the cell monolayers were incubated in base medium containing 20 mM HEPES, pH 8.5, at room temperature. After 30 min, base medium was removed, and the cells were rapidly washed with 1 ml tracer-free mannitol uptake buffer containing 260 mM mannitol, 20 mM Tris/2-(N-morpholino)ethanesulfonic acid, pH 7.0. This was followed by incubation with or without 600 μM DIDS in uptake buffer for 5 min, since this time period is within the linear range of ¹²⁵I⁻ uptake in cells. The radioactive ¹²⁵I⁻ in sodium iodide (3 mM) was added to the uptake buffer at a concentration of 1.0 μCi/ml. Uptake was stopped by removing the radioactivity-containing buffer and washing the cells rapidly two times with ice-cold PBS, pH 7.2. Furthermore, the cells were solubilized with 0.5 N NaOH for 4 h, and protein concentration was measured by the Bradford method (8). Radioactivity was measured by a Packard tri-Carb 1600 TR Liquid Scintillation Analyzer (Packard Instruments; PerkinElmer, Waltham, MA). The Cl⁻/HCO₃⁻ (OH⁻) exchange activity was then calculated as DIDS-sensitive ¹²⁵I⁻ uptake, and the specific activity is expressed as nanomoles per milligram protein per 5 min.

**Transfections**

Caco-2 cells were transfected with DRA promoter (p-1,183/+114), cloned upstream of the luciferase reporter gene in pGL2-Basic and β-galactosidase expression vector by electroporation using an Amassa Nucleofector System as described previously (34). Activities of Firefly Luciferase and β-galactosidase were measured according to the manufacturer’s instructions (Promega, Madison, WI). DRA promoter activity was expressed in terms of relative luciferase activity normalized to β-galactosidase activity.

**In Vivo Studies**

All of the animal studies were approved by the Animal Care Committee of the University of Illinois at Chicago and Jesse Brown Veteran Affairs Medical Center. C57BL/6j mice (male, 8 wk old) were obtained from Jackson Laboratories (Bar Harbor, ME). Four groups of mice (8 animals/group) were used in this study for DSS induction of mild colitis as described previously (21, 33). In two of the groups 3% (wt/vol) DSS (mol mass 36–50 kDa; MP Biomedicals, Solon, OH) was given orally in drinking water for 7 days, whereas the control groups received drinking water only. In one of the two DSS groups, LA suspended in sterile PBS was administered by oral gavage (3 × 10⁹ CFUs) along with DSS one time per day for the first 2 days (LA + DSS), whereas the other group received sterile PBS with no bacteria. Mice were daily monitored for weight change. On day 8, mice were euthanized, the entire colon was removed, and the length and weight were recorded. Mucosa was scraped from the distal colon for RNA and protein extraction.

**Myeloperoxidase Activity**

Myeloperoxidase (MPO) activity in the distal colon was assessed using the method of Krawisz et al. with minor modifications (18). MPO activity was normalized to the amount of protein in supernatant as measured by Bradford’s method (8) and calculated as units per gram protein and expressed relative to control (considered as 100).

**Real-Time PCR**

RNA was extracted from mouse colonic mucosal samples using Qiagen RNeasy kits. RNA was reverse transcribed and amplified using a Brilliant SYBR Green qRT-PCR Master Mix kit (Stratagene). Mouse DRA was amplified with gene-specific primers. Mouse histone or GAPDH was amplified as an internal control (28). Relative levels of DRA mRNA were expressed as percent of control normalized to histone or GAPDH.

**Western Blotting**

Tissue lysates were prepared from the scraped colonic mucosa using cell lysis buffer (Cell Signaling, Danvers, MA). Lysates were run on an 8% gel and then transferred onto nitrocellulose membrane. Immunoblotting was carried out with anti-DRA affinity-purified antibody as previously described (6). Bands were visualized with enhanced chemiluminescence detection reagents.

**Immunofluorescence Staining in Mouse Colonic Tissues**

Sections of colonic tissues from different mice groups were snap-frozen in optimal cutting temperature embedding medium. For immunostaining, 5-μm frozen sections were fixed with 1% paraformaldehyde in PBS for 10 min at room temperature. Fixed sections were washed in PBS, permeabilized with 5% Nonidet P-40 for 5 min, and blocked with 5% normal goat serum (NGS) for 30 min. Tissues were incubated with DRA antibody (1:100) in PBS with 1% NGS for 90
Fig. 1. Long-term treatment with *Lactobacillus acidophilus* (LA)-culture supernatant (CS) stimulates Cl⁻/HCO₃⁻ exchange activity in Caco-2 cells. Overnight serum-starved postconfluent Caco-2 cells were treated with 1:10 dilution of CS of LA, *L. rhamnosus* (LR), *L. plantarum* (LP), and *L. casei* (LC) for 24 h, and apical Cl⁻/HCO₃⁻ exchange activity [4,4'-diisothiocyanate-stilbene-2, 2'-disulfonic acid (DIDS)-sensitive ¹²⁵I uptake] was measured. Values are means ± SE; *n* = 3 mice. *P* < 0.05 compared with control.

Statistical Analysis

Data are presented as means ± SE of 3–8 independent experiments. Differences between control vs. treated were analyzed using one-way ANOVA with Tukey’s test. Differences were considered significant at *P* ≤ 0.05.

RESULTS

**Long-Term Treatment with LA-CS Stimulates Cl⁻/HCO₃⁻ Exchange Activity in Caco-2 Cells**

Our earlier studies showed that short-term treatments with LA and *L. rhamnosus* were effective in enhancing Cl⁻/HCO₃⁻ (OH⁻) exchange activity (5). In the current studies, the effects of long-term treatments with various *Lactobacillus* species on Cl⁻/HCO₃⁻ (OH⁻) exchange activity were evaluated. Postconfluent Caco-2 monolayers were treated with the CS (diluted 1:10 in DMEM) from different species of *Lactobacillus* for 24 h, and Cl⁻/HCO₃⁻ exchange activity was measured as DIDS-sensitive ¹²⁵I uptake. As shown in Fig. 1, LA-CS significantly enhanced Cl⁻/HCO₃⁻ exchange activity (~2-fold) in Caco-2 cells after 24 h. The CS of *L. rhamnosus*, *L. plantarum*, and *L. casei* showed no significant effect. Therefore, all further in vitro or in vivo studies were carried out only with LA.

**Live LA or CS blocks IFN-γ-Induced Inhibition of Cl⁻/HCO₃⁻ Exchange Activity in Caco-2 Cells**

We next examined whether LA can counteract the inhibitory effects of IFN-γ on DRA function. Postconfluent Caco-2 monolayers were pretreated with live bacteria or CS (LA-CS) from the apical surface for 1 h, which was continued for another 24 h with and without IFN-γ (30 ng/ml) added basolaterally. The concentrations of IFN-γ used in this study are based upon our previous studies showing inhibition of NHE3 (2) and DRA (34) gene expression by IFN-γ in Caco-2 cells as well as other studies (15). Live LA or LA-CS (1:10) significantly stimulated activity, whereas IFN-γ treatment significantly (*P* < 0.05 vs. control) inhibited the Cl⁻/HCO₃⁻ exchange activity in Caco-2 cells. Furthermore, both live LA (Fig. 2A) and LA-CS (Fig. 2B) completely blocked the IFN-γ-induced inhibition of Cl⁻/HCO₃⁻ exchange. These results show that either live bacteria or their secreted products in CS are enough to block the inhibition of Cl⁻/HCO₃⁻ exchange activity in response to IFN-γ treatment.

**LA-CS Increases DRA mRNA Levels and Blocks IFN-γ-Induced Decrease in DRA**

We next examined whether LA counteracts the IFN-γ-mediated decrease in Cl⁻/HCO₃⁻ exchange activity via alterations in DRA mRNA levels. For these studies, cells were pretreated with LA-CS (1:10) from the apical surface for 1 h, which was continued for an additional 24 h, with and without IFN-γ (30 ng/ml) added basolaterally in serum-reduced medium (1% FBS). As reported previously by us (28) DRA mRNA levels were significantly increased (~2.2-fold) in re-
LA-CS Attenuates IFN-γ-Induced Downregulation of DRA (Diaminoribose) mRNA Levels in Caco-2 Cells

We have previously shown that the activity of DRA full-length promoter construct p-1183/+114 exhibited (~50%) inhibition in response to IFN-γ treatment compared with untreated controls (1, 34). In an attempt to examine whether LA-CS can reverse the effects of IFN-γ on DRA promoter, Caco-2 cells were transfected with DRA full-length promoter construct p-1183/+114. Twenty-four hours posttransfection, cells were treated with LA-CS (1:10 dilution) apically or IFN-γ basolaterally or coincubated with LA-CS and IFN-γ for 24 h, and DRA promoter activity was assessed. DRA promoter activity was markedly increased (~1.5-fold) in response to LA-CS; IFN-γ treatment exhibited a 50% decrease in the activity, whereas the inhibitory effects of IFN-γ on DRA promoter activity were blocked in cells pretreated with LA-CS alone.

LA-CS Attenuates IFN-γ-Induced Inhibition of DRA Promoter Activity in Caco-2 Cells

In Vivo Effects of LA on DSS-Induced Colitis in Mice

Our in vitro studies in Caco-2 cells clearly demonstrated that out of four Lactobacillus species tested, long-term treatment with LA showed maximum increase in Cl−/HCO3− exchange activity by increasing DRA expression. Also LA-CS was able to block the inhibition of Cl−/HCO3− exchange activity and repression of DRA promoter caused by IFN-γ in Caco-2 cells. Therefore, we get a comprehensive idea of the effect of this probiotic strain in a complex physiological setting, we next investigated the effects of LA on DRA expression in an in vivo mouse model of DSS-induced colitis.

LA did not attenuate DSS-induced loss of body weight. The DSS-induced colitis murine model is commonly used as a chemical injury model to address the pathogenesis of inflammatory bowel disease (14, 37). To test the efficacy of LA in ameliorating colitis, C57BL/6 mice were given 3% (wt/vol) DSS in drinking water for 7 days. As shown in Fig. 5A, significant weight loss was evident in the DSS-treated group compared with the control or LA-treated group. Whereas animals in the control group gained weight over a 1-wk period of observation, the mice in the DSS group showed significant reduction in weight over the same time, which was evident starting after the 5th day of DSS treatment and persisted until the 6th and 7th day compared with control. However, this weight loss was not attenuated in the LA + DSS mice group (Fig. 5A).

LA reduces MPO activity in acute DSS-treated mice colon. Because the DSS-induced colitis mostly affects the distal part of the colon (26), a part (2 cm) of distal colon was used for determination of MPO activity. Colonic MPO level, an index of neutrophil accumulation, was significantly increased in DSS mice compared with control or LA mice (P < 0.01 vs. control or LA) (Fig. 5B). This increase in MPO levels was attenuated by LA treatment in the LA + DSS group (Fig. 5B). These results suggested that LA exerts its protective effects in the colon of mice with colitis by decreasing inflammation caused by DSS.

Changes in colon weight-to-length ratio. Colon weight and length were recorded for each mouse immediately after death (8th day since the start of DSS treatment) (Fig. 5C). The length of the colon was significantly shortened (P < 0.05) in mice in the DSS group (Table 1). Although the colon length was greater in the LA group, compared with control, this was not statistically significant (P > 0.05 vs. control).

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A treatment groups

P sulfat sodium. Full colonic length and weight (excluding cecum) were

Weight, g 0.38

Length, cm 6.48

Table 1. Colonic weight and colon length of different
treatment groups

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>LA</th>
<th>DSS</th>
<th>LA + DSS</th>
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<tr>
<td>Length, cm</td>
<td>6.48 ± 0.18</td>
<td>6.75 ± 0.40</td>
<td>5.04 ± 0.09*</td>
<td>5.75 ± 0.24</td>
</tr>
<tr>
<td>Weight, g</td>
<td>0.38 ± 0.03</td>
<td>0.33 ± 0.02</td>
<td>0.41 ± 0.03</td>
<td>0.39 ± 0.03</td>
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Values are means ± SE. LA, Lactobacillus acidophilus; DSS, dextran sulfate sodium. Full colonic length and weight (excluding cecum) were measured immediately after death. *P < 0.05 vs. control.

LA Blocks DSS-Induced Decrease in DRA mRNA and Protein Expression in Distal Colon

Our previous studies have already shown that DRA mRNA and protein expression in mice colonic tissue increases in response to live LA gavaged for 24 h (28). To examine whether LA could restore DSS-induced downregulation of DRA mRNA and protein expression, we examined DRA mRNA and protein levels in distal colonic mucosa of control, DSS, LA, and LA + DSS mice groups using real-time PCR and immuno-blotting. As shown in Fig. 6, DRA mRNA levels in distal colon were significantly increased (~1.5-fold) in response to LA administration. DSS caused a significant decrease in DRA mRNA and protein expression (~50–60%) in distal colon. On the other hand, LA treatment to DSS mice significantly attenuated this decrease in the levels of DRA mRNA (Fig. 6) and protein (Fig. 7) in LA + DSS mice. LA-mediated reversal of DSS effects on DRA expression was also measured by immuno-fluorescence staining of colonic sections. As shown in Fig. 8, the apical membrane level of DRA was substantially reduced in response to DSS treatment. On the other hand, LA administration not only increased apical membrane DRA level compared with control but also almost completely blocked the inhibitory effects of DSS on DRA expression (Fig. 8). These results suggest that modulation of DRA protein expression by LA might be a major contributor to the observed anti-diarrheal effects of LA in DSS colitis.
have potential therapeutic value in treating diarrhea due to their Cl−/HCO3− exchange activity (35). We have previously shown that acute stimulation in DRA activity by LA and its impor-
tivity. Parallel to function, IFN-γ-mediated inhibitory effects on apical Cl−/HCO3− exchange (12). Its impor-
tants are localized to basolateral membranes of polarized Caco-2 cells, it is unlikely that live LA or its supernatant-
derived soluble factors counteracted IFN-γ effects via blocking its binding to the receptors. In addition, LA could also act via an independent pathway to counteract inhibition of promoter activity by IFN-γ. For example, involvement of p38, ERK1/2 MAPK, and phosphatidylinositol 3-kinase (PI 3-kinase) have been previously reported in LA-induced modulation of Cl− secretion in Caco-2 and HT29/cl.19a cells (30). We previously showed that acute stimulation in DRA activity by LA and increase in DRA levels on the plasma membrane involved PI 3-kinase (but not MEK1, MEK2, and p38 MAPK pathway)(6). It is possible that the PI 3-kinase-mediated pathway may be involved in observed effects of LA in counteracting the effects of cytokines. Future studies may focus on delineating exact signaling mechanisms underlying the effects of LA.

We extended our in vitro studies showing efficacy of LA in reversing the effects of inflammatory cytokine on DRA activity using an in vivo model of DSS colitis. Based on our previous study (33), we used 3% DSS for 7 days to induce mild colitis in mice, since DSS at high concentrations (4–5%) is known to cause ulcerations and erosion of the epithelium (23). Two doses of LA were chosen based on our previous study showing that, under similar conditions, LA attenuated inflammation and reversed the reduced expression of MDR-1 transporter in DSS-treated mice. Also, multiple studies suggest that probiotic bacteria remain colonized in the gut for at least 7 days to exert beneficial effects (27, 33). In DSS-induced colitis in mice, the most affected segment of the gastrointestinal tract is known to be the distal colon showing inflammation, shortening of length and loose fecal pellet, compared with control mice (29). Our promoter studies have shown that LA-CS enhanced the activity of the full-length (p-1183/+114) DRA promoter. We have previously shown that IFN-γ inhibition of DRA promoter activity involved the JAK/STAT1 pathway (34). Hence, it can be speculated that LA-secreted soluble factors in the CS could reverse the effects of inflammatory cytokines either by blocking the phosphorylation of STAT-1 signaling pathways induced by IFN-γ or via blocking the binding of cytokine to its receptor. However, because live LA or LA-CS were added from the apical side of cell monolayers, whereas IFN-γ receptors are localized to basolateral membranes of polarized Caco-2 cells, it is unlikely that live LA or its supernatant-
derived soluble factors counteracted IFN-γ effects via blocking its binding to the receptors. In addition, LA could also act via an independent pathway to counteract inhibition of promoter activity by IFN-γ. For example, involvement of p38, ERK1/2 MAPK, and phosphatidylinositol 3-kinase (PI 3-kinase)

Diarrhea associated with intestinal inflammatory diseases, including colitis, is the result of a complex interaction of multiple inflammatory mediators and their effects on the intes-
nal epithelium. Diarrhea can result partly in response to enhanced secretion of high levels of proinflammatory cyto-
kines such as IFN-γ and TNF-α (7, 13, 17). Studies have shown that Caco-2 cells treated with proinflammatory cyto-
kines (IFN-γ, IL-1β) have diminished expression of DRA, and the same is true for patients suffering from ulcerative colitis (34, 36). SLC26A3 or DRA is the main candidate gene for luminal human intestinal Cl−/HCO3− exchange (12). Its impor-
tance is further emphasized by DRA knockout mice, which exhibit diarrhea phenotype owing to loss of luminal membrane Cl−/HCO3− exchange activity (35). We have previously shown in in vitro and in vivo models that probiotics such as LA may have potential therapeutic value in treating diarrhea due to their efficacy in increasing Cl−/HCO3− exchange activity and DRA expression involving both short-term trafficking and long-term transcriptional changes, respectively (6, 28). Our group (4, 5) as well as others (30) have previously demonstrated that probiotics reverse the effects of inflammatory cytokines in human intestinal epithelial cells and inflammation in a mouse model (33). Experimental studies have implicated DRA repression as one of the potential events during intestinal inflamma-
tion (19, 38). However, the functional consequences of this repression have not been studied in detail. Also, nothing is known about the potential beneficial effects of probiotics on DRA (Cl−/HCO3− exchange) function and expression under inflammatory states such as colitis.

In the current report, we have shown that pretreatment with LA or bacteria-free CS of LA counteracts the inhibitory effects of IFN-γ on DIDS-sensitive apical Cl−/HCO3− exchange activity. Parallel to function, IFN-γ-mediated inhibitory effects on DRA mRNA levels and promoter activity were abrogated by LA-CS. Consistent with our data, earlier reports have shown that pretreatment with live probiotics reversed the effects of TNF-α or IFN-γ on cystic fibrosis transmembrane conductance regulator (30). These results suggest that LA-derived soluble factors exert distinct effects to alleviate/block the cytokine effects on DRA expression and function. Our functional studies are limited by lack of DRA-specific inhibitor; however, given that LA-CS treatment does not affect PAT-1 expression (28) and our functional read out is parallel to changes in DRA mRNA and protein, it is highly likely that LA-mediated effects on apical Cl−/HCO3− exchange activity are mediated by DRA.

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We extended our in vitro studies showing efficacy of LA in reversing the effects of inflammatory cytokine on DRA activity using an in vivo model of DSS colitis. Based on our previous study (33), we used 3% DSS for 7 days to induce mild colitis in mice, since DSS at high concentrations (4–5%) is known to cause ulcerations and erosion of the epithelium (23). Two doses of LA were chosen based on our previous study showing that, under similar conditions, LA attenuated inflammation and reversed the reduced expression of MDR-1 transporter in DSS-treated mice. Also, multiple studies suggest that probiotic bacteria remain colonized in the gut for at least 7 days to exert beneficial effects (27, 33). In DSS-induced colitis in mice, the most affected segment of the gastrointestinal tract is known to be the distal colon showing inflammation, shortening of length and loose fecal pellet, compared with control mice (29). Our studies also showed that mice administered DSS showed significant weight loss and shortening of the colon. However, these effects were not significantly reversed by administration of live LA. Furthermore, weight loss, MPO activity, and colon weight-to-length ratio (a potential readout for diarrheal pheno-
type) were significantly greater in the DSS group compared
with all other groups. Occurrence of solid fecal pellet in LA + DSS mice indicated potential beneficial effects of LA on diarrheal phenotype and on colonic inflammation caused by DSS. Also, the inhibition of enhanced colonic MPO activity in DSS mice by LA reflects a potent anti-inflammatory effect of LA against tissue injury. The anti-inflammatory/immunomodulatory properties of various *Lactobacillus* species have previously been described by our group and others in experimental models of colitis (10, 25, 33). Studies have shown that fecal excretion of $\text{HCO}_3^-$ is reduced in patients with active ulcerative colitis, which is caused by impairment of the colonic anion exchange process induced by inflammation (9). This is mainly due to a major decrease in the $\text{Cl}^-/\text{HCO}_3^-$ exchange activity in the luminal membrane, associated with a downregulation of the anion exchanger DRA (SLC26A3) in colitis (36). As expected, levels of DRA mRNA and protein expression were dramatically reduced in the distal colon of DSS-treated mice compared with control, whereas the other group where live LA was administrated along with DSS showed significantly less reduction in DRA mRNA and protein levels compared with the DSS-treated group alone. Our results are consistent with previous reports showing partial reversal of the effects of DSS in response to pretreatments with LA (11, 25). However, the mechanism of LA-mediated reduction in gut inflammation is not entirely clear. This could involve attenuation of effect of cytokines, beneficial effects on epithelial integrity, and improvement in barrier function (29).

Overall, our results demonstrated that the soluble factor(s) present in the CS of LA attenuate the IFN-γ-induced decrease in DRA function, mRNA and promoter activity. Also, live LA administration in vivo showed its effects by reversing the DSS-induced decrease in DRA mRNA and protein levels and appeared to have potential antidiarrheal effects in an experimental model of DSS colitis. These results are of critical

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importance in increasing our understanding of the molecular basis of the beneficial effects of L.A and L.A-derived molecules. Additionally, efficacy of LA supernatant to counteract inflammation-induced downregulation of DRA expression and function highlights the novel therapeutic potentials of L.A-secreted soluble factors for diarrheal disorders associated with gut inflammation where live bacteria may be counterindicated for therapeutic purposes due to compromised barrier function of the intestine. In future studies, it will also be important to examine efficacy of LA as a therapeutic (after DSS treatment) rather than preventive agent in reversing inflammation and DRA expression.

REFERENCES


