Gastrin stimulates MMP-1 expression in gastric epithelial cells: putative role in gastric epithelial cell migration

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Submitted 13 March 2015; accepted in final form 6 May 2015

Am J Physiol Gastrointest Liver Physiol 309: G78–G86, 2015. First published May 14, 2015; doi:10.1152/ajpgi.00084.2015.—The pyloric antral hormone gastrin is a primary regulator of gastric luminal environment by virtue of its action in stimulating acid secretion from parietal cells (11). Although these cells express the CCK-2 receptor at which gastrin acts, it is generally thought that gastrin increases acid secretion largely via indirect effects mediated by histamine release from enterochromaffin-like (ECL) cells which also express CCK-2 receptors (4, 13). Thus gastrin stimulates histamine release from these cells, which in turn stimulates acid secretion, and it also increases histamine synthesis and storage via induction of histamine decarboxylase (HDC) and vesicular monoamine transporter-2 (6–8). In addition, it is now well recognized that gastrin plays a role in regulating the organization of the gastric mucosa (29, 36). These effects include increased ECL cell numbers (19), stimulation of cell proliferation, mucosal thickness (25), epithelial cell migration (15, 23), and branching morphogenesis (26). The actions of gastrin in regulating mucosal organization are mediated, at least partly, by induction of genes encoding proteins involved in remodeling of the extracellular matrix, including both proteases, notably matrix metalloproteinases (MMPs), and urokinase plasminogen activator, and their inhibitors, notably tissue inhibitors of MMPs (TIMPs) and plasminogen activator inhibitor-1 and -2 (PAI-1, PAI-2) (5, 32, 33, 40).

The MMPs are a family of ~25 members that degrade extracellular matrix and other extracellular proteins (22, 30). They may be either soluble or membrane bound and are typically produced as inactive precursors that are converted to their active form after delivery to the cell surface. The expression of MMPs in stromal cells is common, and with the exception of MMP-7, they are less commonly expressed in normal epithelial cells. However, gastric mucosa may be an exception in this regard since there is reported to be expression of MMP-1, -2, and -9 in parietal cells (31). In addition, there is evidence that MMP-1, -2, and -9 are expressed in gastric cancer in both tumor cells and stroma (14, 21), and infection with the oncogenic bacterium *H. pylori* is associated with induction of MMP-1 (17, 27, 41). In contrast, rather less is known of the factors that might regulate MMP-1 expression in normal gastric mucosa in the absence of *H. pylori*, inflammation, or cancer. In view of the expression in parietal cells of both MMP-1 and the CCK-2 receptor, we have now examined the hypothesis that gastrin regulates gastric MMP-1 expression. We report here an association between serum gastrin concentrations and MMP-1 transcript abundance in the gastric corpus mucosa of healthy subjects, and we show that gastrin increases MMP-1 expression in gastric epithelial AGS-G8 cells via protein kinase C (PKC) and p42/44 MAP kinase activity. Gastrin-stimulated AGS-G8 cell migration in both scratch wound and Boyden chamber assays was inhibited by MMP-1 immunoneutralization. We conclude that MMP-1 expression is a target of gastrin implicated in mucosal remodeling.

**THE PYLORIC ANTRAL HORMONE** gastrin is a primary regulator of the gastric luminal environment by virtue of its action in stimulating acid secretion from parietal cells (11). Although these cells express the CCK-2 receptor at which gastrin acts, it is generally thought that gastrin increases acid secretion largely via indirect effects mediated by histamine release from enterochromaffin-like (ECL) cells which also express CCK-2 receptors (4, 13). Thus gastrin stimulates histamine release from these cells, which in turn stimulates acid secretion, and it also increases histamine synthesis and storage via induction of histamine decarboxylase (HDC) and vesicular monoamine transporter-2 (6–8). In addition, it is now well recognized that gastrin plays a role in regulating the organization of the gastric mucosa (29, 36). These effects include increased ECL cell numbers (19), stimulation of cell proliferation, mucosal thickness (25), epithelial cell migration (15, 23), and branching morphogenesis (26). The actions of gastrin in regulating mucosal organization are mediated, at least partly, by induction of genes encoding proteins involved in remodeling of the extracellular matrix, including both proteases, notably matrix metalloproteinases (MMPs), and urokinase plasminogen activator, and their inhibitors, notably tissue inhibitors of MMPs (TIMPs) and plasminogen activator inhibitor-1 and -2 (PAI-1, PAI-2) (5, 32, 33, 40).

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**MATERIALS AND METHODS**

**Cells and reagents.** AGS-G8 cells were maintained as previously described (35). HGT-1 cells were kindly donated by Dr. C. Laboisse (INSERM U239, Hôpital Bichat, Paris, France), (18) and RGM-1 cells were obtained from Riken BioResource Centre (Tsukuba, Ibaraki, Japan) (16); both had been transfected with the CCK-2 receptor and subcloned to yield HGT1-G8 and RGM1-G8 cells, as described (38). Unsulfated human heptadecapeptide gastrin (hG17ns) was obtained from Bachem (St Helens, Merseyside, UK). Phorbol 12-myristate 13-acetate (PMA) was obtained from Calbiochem (Darmstadt, Germany); brefeldin A (BFA) was obtained from Cambio.
(Dry Drayton, Camb, UK). 125I-LG17 was purchased from Perkin Elmer (Cambridge,Cambs, UK). Patients. Subjects were selected from a cohort of ~1,400 patients, aged 18 and over, who had clinical indications for undergoing diagnostic upper gastrointestinal endoscopy. Subjects were selected for the current investigation if they were H. pylori negative and showed no endoscopic or histological evidence of upper gastrointestinal neoplasia or preneoplastic pathology (atrophic gastritis, gastric intestinal metaplasia, or Barrett’s esophagus). Further exclusion criteria included diabetes mellitus, coma or hemodynamic instability, being moribund or having terminal malignancy, cirrhosis (Child B or C), abnormal clotting or bleeding diathesis, inability to give informed consent, contraindication to endoscopy, pregnancy, HIV, hepatitis B or C infections. Subjects underwent diagnostic gastroscopy in the Gastroenterology Unit at the Royal Liverpool University Hospital. Endoscopic pinch biopsies of gastric corpus and antrum (2–4 of each) were obtained for histology; H. pylori status was determined on the basis of serrality, antral urease test (Pronto Dry; Medical Instrument, Solothurn, Switzerland), and antral and corpus histology. An additional 8 corpus biopsies were taken for RNA extraction and real-time PCR analysis. The study groups consisted of controls and patients taking PPIs (n = 33), omeprazole 20–40 mg; n = 4, esomeprazole 20–40 mg; n = 41, lansoprazole 15–30 mg; n = 2, pantoprazole 20 mg; n = 4, rabeprazole 20 mg). The study was approved by the Liverpool Local Research Ethics Committee and by the Royal Liverpool and Broadgreen University Hospitals NHS Trust, and all patients gave written, informed consent.

**INS-gas mice.** INS-Gas mice or FVB/N wild-type controls were maintained in an appropriately controlled environment with a 12:12-h light/dark cycle and were fed a commercial pellet diet with water ad libitum as previously described (37). Animals were killed by increasing CO2 concentration. Gastric corpus extracts were prepared from unfasted animals in RIPA buffer as previously described (20). All animal experiments were approved by the University of Liverpool Animal Welfare Committee, and were conducted in compliance with Home Office requirements and the UK Animals (Scientific Procedures) Act 1986.

**Real-time PCR.** Corpus biopsies were collected in RNA Later (Life Technologies LTD, Paisley, Scotland, UK) and RNA extracted in 1.0 ml Trizol. (Invitrogen, Seraing, Belgium) according to the manufacturer’s instructions. RNA pellets were resuspended in 30 μl of nuclease free water and 2 μg of RNA reverse transcribed with avian myeloblastosis virus reverse transcriptase and oligo(dT) primers (Promega, Southampton, Hampshire, UK). Real-time PCR was carried out using an ABI7500 platform (Applied Biosystems, Warrington, Lancashire, UK) using TaqMan primer/probe sets (human MMP-1, MMP-3, MMP-7, GAPDH), Precision 2x real time PCR master mix (Primer Design, Southampton, UK), and 5'-FAM, 3'-TAMRA double dye probes (Eurogentec, Southampton, Hampshire, UK). All values were standardized to GAPDH. Assays included a no template control (NTC) and 3 quality controls and were only accepted if they met the following criteria: the quality controls within 15% of their anticipated mean quantity, PCR amplification efficiency between 90–110%, and the correlation coefficient of the slope of the standard curve greater than 0.97. Primers and probes were designed using Primer Express v3.0 (Applied Biosystems) and were purchased from Eurogentec (Seraing, Belgium). Probes for detection of human MMP-1, MMP-3, MMP-7, and GAPDH cDNA were intron-spanning and were as follows: MMP-1, 5’-TGA AGG TGT AGG TAG GGT ACA TCA AA-3’ (forward), 5’-CCA ACA ATT TCA GAG AGT AGA ACT TAC AT-3’ (reverse); MMP-3, 5’-TTG CTG CTC ATG AAA TTG GCC ACT CC-3’ (probe), 5’-ACA AAG GAT ACA GGG ACC AA-3’ (forward); 5’-TAG AGT GGG ACT ATC AAA GCT TCA GT-3’ (reverse); MMP-7, 5’-CCT GTA TGC AAC TCA TGA ACT TGG C-3’ (probe), 5’-GGA TGT TAG CAC TT GAG GTT AAT CT-3’ (forward), 5’-GGA ATG TCC CAT ACC CAA AG-3’ (reverse); GAPDH, 5’-CGT CGC CAG CCG ACC AAC A-3’ (probe), 5’-GCT CCT GTT CGA CAG TCA-3’ (forward), 5’-ACC TTC CCC ATG GTG TCT GA-3’ (reverse).

**Gastrin radioimmunoassay.** Serum samples were assayed for total amidated gastrin concentrations by radioimmunoassay using antibody L2 (which reacts with G-17 and G-34 but not pregastrin or Gly-gastrins) and 125I-G-17 as previously described (12). The upper limit of the reference range for fasting serum gastrin in this assay is 30 pm.

**Immunohistochecmistry.** Tissue sections from gastric corpus biopsies fixed in 10% neutral-buffered formalin and paraffin-embedded were processed for immunohistochemical detection of MMP-1 using mouse monoclonal antibody to MMP-1 (R and D Systems, Minneapolis, MN), which reacts with both pro- and active MMP-1, and En Vision FLEX/HRP (Dako, Carpenteria, CA) as secondary antibody; antigen retrieval was performed by incubating at pH 9.0, 93°C, for 15 min as previously described (33).

**AGS-G6 cell secretion of MMP-1.** AGS-G6 cells that express the CCK-2 receptor (35) were incubated in serum-free medium with or without ghG17ns (1–10 nM) for up to 24 h. In different experiments, cells were treated with PMA (100 nM), Ro320432 (2 μM), U0126 (10 μM), or BFA (10 μg/ml). Media was concentrated using Strataclean resins (Agilent Technologies, Santa Clara, CA), and cell extracts prepared in RIPA buffer containing protease and phosphatase inhibitors (Calbiochem) were resolved by SDS-PAGE electrophoresis.

**Western blots.** Proteins were resolved by SDS-PAGE electrophoresis, transferred to nitrocellulose (Amersham Pharmacia Biotech, Buckinghamshire, UK), and incubated with antibodies to MMP-1 (R and D Systems) followed by horseradish peroxidase-conjugated secondary antibody and detection by incubation with Clarity Western ECL Blotting Substrate (Bio-Rad Laboratories, Hertfordshire, UK) and exposure to HyperFilm (Amersham Pharmacia Biotech). Samples of cell extracts were reprobed for GAPDH (Biodisc, Saco, ME) to normalize for protein loading.

**Enzyme assays.** Fluorogenic assays for enzyme activity were performed using the MMP-1 selective substrate DNP-Pro-Cha-Abu-Cys(Me)-His-Ala-Lys(Ne-Me-Abz)-NH2 (Cha, β-cyclohexylalanyl; Abz, 1-α-aminobutyryl; Abz, 2-aminobenzoyl) (Calbiochem) at a concentration of 12 μM with equal volumes of assay buffer and media from 3 × 106 AGS-G6 cells as previously described (14).

**Migration assays.** The action of neutralizing MMP-1 antibodies (2.5 μg/ml; MAB901, R and D Systems) on the stimulation of AGS-G6 cell migration by ghG17ns (10 nM) was studied using scratch-wound assays and Boyden chambers as previously described (23, 24).

**Statistics.** Results are expressed as means ± SE. The association between MMP-1 and serum gastrin was examined using Spearman rank correlation. Other comparisons were made using t-tests or ANOVA as appropriate.

**RESULTS**

**Increased corpus MMP1 mRNA is associated with elevated serum gastrin.** In human subjects with normal gastric histology and negative for H. pylori, the relative abundance of proMMP-1 transcripts in corpus biopsies was ~2-fold higher in those receiving PPIs compared with those not (P < 0.05) (Fig. 1A). As expected there was also a significant difference in serum gastrin concentrations. To dissociate differences associated with PPI usage per se from those associated with serum gastrin, we first compared proMMP-1 transcript abundance in patients on PPIs divided into normal (<30 pM) and elevated (>30 pM) serum gastrin concentrations. There was again an ~2-fold difference in MMP-1 transcripts in the two groups (P < 0.05) (Fig. 1B). We then compared proMMP-1 transcript abundance in control subjects and those PPI-treated subjects in whom serum gastrin concentrations were within the reference
biopsies revealed MMP-1 immunoreactivity in glandular metaplasia.

**Immunohistochemistry of normal corpus mucosal biopsies within the reference range (Fig. 1).**

When the patients as a whole were divided into quartiles based on serum gastrin, it was clear that the increase in MMP-1 was associated with the third and fourth quartiles of serum gastrin (Fig. 1D). The correlation between serum gastrin and MMP-1 transcript abundance was statistically significant (Spearman rho, 0.255; degrees of freedom, 160; \( P = 0.00105 \)). There was also a significant correlation between serum gastrin and MMP-1 transcript abundance in the PPI group alone (Spearman rho, 0.257; degrees of freedom, 83; \( P = 0.0175 \)), but there was no significant correlation in the control group. The data therefore point to an association between serum gastrin in concentrations above those in normal fasting individuals and MMP-1 transcript abundance.

The specificity of the association between circulating gastrin and increased gastric mucosal proMMP-1 transcript abundance is indicated by the fact that proMMP-3 transcripts did not differ significantly in subjects with high and low serum gastrin concentrations (Fig. 1A and B). However, previous studies have shown stimulation of MMP-7 expression by gastrin (33), and consistent with this we found a 2.3-fold elevation in proMMP-7 transcript abundance in subjects on PPIs compared with controls (\( P < 0.05 \)), and a 2.6-fold elevation in patients on PPIs with serum gastrin above the reference range compared with normal serum gastrin (\( P < 0.05 \)) (Fig. 1, A and B); there was no significant difference between control subjects and those on PPIs in whom serum gastrin concentrations were within the reference range (Fig. 1C).

**Immunohistochemical localization of MMP-1 in hypergastrinemia.** Immunohistochemistry of normal corpus mucosal biopsies revealed MMP-1 immunoreactivity in glandular epithelial cells (Fig. 2). The localization was consistent with expression in both parietal and chief cells. Generally mucus neck and surface epithelial cells were either lightly stained or were negative. Staining in the stroma was also either very light or negative. There was no obvious difference in intensity of staining between patients in the different groups (Fig. 2).

**Western blot analysis of MMP-1 in corpus mucosa.** Gastric corpus biopsies selected from individuals with either low or high gastrin were processed for detection of MMP-1 by Western blot. In both cases, there was a major band corresponding to proMMP-1 (Fig. 3A). There was clear evidence of an association between proMMP-1 abundance and serum gastrin (\( P < 0.05 \)) (Fig. 3, A and B). We then examined MMP-1 by Western blot of extracts of gastric corpus from an animal model of hypergastrinemia, the INS-Gas mouse (37). Again, there was a major band corresponding to proMMP-1 and a significant association between proMMP-1 abundance and serum gastrin (\( P < 0.05 \)) (Fig. 3, C and D).

**Stimulation of MMP-1 expression in AGS-G\(_R\) cells: role of CCK-2 receptors, PKC, and p42/44 MAP kinase.** To study the cellular action of gastrin on MMP-1 expression we examined three different cell lines expressing CCK-2 receptors. In Western blots, MMP-1 immunoreactivity was virtually undetectable in unstimulated cells but in response to 10 nM hG17ns there was stimulation in media and cell extracts of all three cell lines of a band of \( \sim 52 \) kDa corresponding to proMMP-1 (Fig. 4A). The response was greater in AGS-G\(_R\) cells than either RGM1-G\(_R\) or HGT1-G\(_R\); moreover AGS-G\(_R\) cells have been intensively used for studies of the effects of gastrin in vitro, thereby justifying their use for more detailed studies. Western blots of media revealed a concentration-related increase of...
proMMP-1 over the range 1–10 nM hG17ns in AGS-GR cells (Fig. 4B); there was also a faint band of ~42 kDa corresponding to the active form. The action of G17 was mediated by CCK-2 receptors because 1) a CCK-2 receptor antagonist, L740,093, inhibited the effect of G17 in AGS-GR cells but had no effect on PMA used as a positive control (Fig. 4C), and 2) G17 had no effect on MMP-1 abundance in the media of AGS cells which do not express the receptor while again PMA was a good stimulant (Fig. 4D).

Stimulation of proMMP-1 expression by G17 was reversed by the PKC inhibitor, Ro320432. Moreover, as expected, PMA-stimulated proMMP-1 expression was also inhibited by Ro320432. Evidence that PKC acts via a mechanism involving activation of p42/44 MAP kinase was provided by the observation that the effects of both G17 and PMA were inhibited by the MEK inhibitor, U0126 (Fig. 4E).

Activation of proMMP-1 by AGS-GR cells. Although the primary secretory product of AGS-GR cells treated with G17 was proMMP-1, at high exposures Western blots revealed minor bands corresponding to the active enzyme. To determine the capacity of AGS-GR cells for proMMP-1 activation, we performed stop-flow experiments making use of the ability of BFA to arrest secretion by blocking transport through the early secretory pathway. In cells treated with BFA, proMMP-1 accumulated in cell extracts and was virtually undetectable in media (Fig. 5A). By treating AGS-GR cells with G17 followed by delayed addition of BFA after 16 h, we were able to examine the metabolism in media of the previously secreted protein. In these experiments, there was little further conversion over the period of 16–24 h of proMMP-1 to smaller bands; moreover, relatively abundant proMMP-1 was found in media even after 8 h of BFA treatment so that the precursor is relatively stable in AGS-GR cell media. Although Western blots indicated that bands corresponding to the active enzyme were minor compared with proMMP-1, enzyme activity assays for MMP-1 using the selective substrate DNP-Pro-Cha-Abu-Cys(Me)-His-Ala-Lys(N-Me-Abz)-NH₂ (Cha, β-cyclohexylalanyl; Abu, L-α-amino butyryl; Abz, 2-aminobenzoyl) suggested that gastrin increased enzyme activity 2-fold (Fig. 5B); BFA inhibited secretion of the active enzyme but in delayed addition experiments had no effect on the activity of previously secreted enzyme (Fig. 5B), again confirming that in the bulk phase of the cell media there is little or no conversion of proMMP-1 to active enzyme.
Stimulation of cell migration by G17 is partially mediated by MMP-1. Gastrin stimulates AGS-GR cell migration (23) and to determine the role of MMP-1 we examined the effect of immunoneutralization of MMP-1 in scratch-wound migration assays (Fig. 6, A and B) and in Boyden chamber chemotaxis assays (Fig. 6 C). As expected, gastrin stimulated AGS-GR cell migration in both types of assay. Immunoneutralization reduced the response by about 35% ($P < 0.05$) in both cases (Fig. 6, A–C).

**DISCUSSION**

The present data show an association between serum gastrin concentrations and the abundance of MMP-1 transcripts in biopsies from normal human stomach. In gastric epithelial cells expressing CCK-2 receptors, gastrin stimulates proMMP-1 expression through a mechanism involving PKC and p42/44 MAP kinase, and increases MMP-1 enzyme activity in the media. Moreover, MMP-1 is at least partly involved in mediating gastrin-stimulated cell migration. Thus MMP-1 is a candidate for mediating the actions of gastrin in the remodeling of gastric epithelia in both health and disease.

The observation that serum gastrin concentrations are linked to increased abundance of MMP-1 transcripts in corpus biopsies from normal human stomach. In gastric epithelial cells expressing CCK-2 receptors, gastrin stimulates proMMP-1 expression through a mechanism involving PKC and p42/44 MAP kinase, and increases MMP-1 enzyme activity in the media. Moreover, MMP-1 is at least partly involved in mediating gastrin-stimulated cell migration. Thus MMP-1 is a candidate for mediating the actions of gastrin in the remodeling of gastric epithelia in both health and disease.

**Fig. 3.** Increased proMMP-1 in corpus biopsies from hypergastrinemic humans and mice, detected by Western blot. A: Western blot showing proMMP-1 in gastric corpus biopsies from four subjects with serum gastrin < 30 pm (Hi-gas, −) and four subjects with hypergastrinemia (Hi-gas, +). B: means ± SE of serum gastrin and abundance of MMP-1 estimated by densitometry of Western blots of the data shown in A. C: Western blot showing proMMP-1 in gastric corpus biopsies from four wild-type mice and four INS-Gas hypergastrinemic mice. D: means ± SE of serum gastrin and abundance of MMP-1 estimated by densitometry of Western blots of data shown in C.

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strongest response in AGS-GR cells. We made use of the latter for more detailed studies since they are a relatively well studied model of gastrin-stimulated gene expression (1, 32, 33, 39) that has also been used for studies of \textit{H. pylori}-induced MMP-1 expression (17, 27). The data suggest that gastrin increases proMMP-1 appearance in the media of these cells via stimulation of CCK-2 receptors. Other studies have previously reported that Gly-extended gastrins (which have low or no affinity for CCK-2 receptors) may increase MMP-1 expression in LoVo and HT29 colon cancer cells (3). If so, however, this is likely to be by a different mechanism to that reported here. Our data indicate that gastrin increases MMP-1 via activation of PKC and of p42/44 MAP kinase. Previously, a promoter element in the \textit{MMP-1} gene required for induction by PMA has been defined (2). Moreover, in AGS cells there are p42/44 MAP kinase-mediated increases in MMP-1 stimulated by inflammatory signals and \textit{H. pylori} (27, 28). The present data suggest that a noninflammatory endocrine stimulus, gastrin, also activates this pathway via CCK-2 and PKC. In the same cells, gastrin increases \textit{PAI-1} and \textit{PAI-2} gene expression by PKC and p42/44 MAP kinase activation. Interestingly \textit{PAI-1} and \textit{PAI-2} are inhibitors of extracellular proteases and suppress migration. Thus activation of a common signaling pathway leads not just to release of proinvasive MMP-1 but also to putative restraining mechanisms.

Enzyme assays confirmed increases in MMP-1 activity in response to gastrin. Western blots of media from gastrin-stimulated cells showed a major band corresponding to proMMP-1 and a relatively minor band corresponding to the active form. Thus AGS-GR cells have some capacity to activate

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**Fig. 4. Gastrin stimulates proMMP-1 expression by AGS-G\textsubscript{R} cells revealed by Western blot.**

**A:** Western blots of HGT1-G\textsubscript{R}, RGM1-G\textsubscript{R}, and AGS-G\textsubscript{R} cells reveal stimulation of proMMP-1 by hG17ns (10 nM) in media (top) and in cell extracts (middle) while there is no difference in cellular GAPDH (bottom).

**B:** concentration-dependent increases in proMMP-1 (52 kDa) in response to hG17ns (1–10 nM).

**C:** stimulation of proMMP-1 by hG17ns (10 nM) is inhibited by the CCK-2 receptor antagonist L740,093 (0.2 \textmu M), while responses to PMA (100 nM) are not; the changes in proMMP-1 can be seen both in media and cell extracts but there is no change in GAPDH abundance in cell extracts.

**D:** hG17ns and PMA increase proMMP-1 abundance in AGS-GR media, but only PMA also stimulates AGS cells which lack the CCK-2 receptor.

**E:** the effects of both gastrin and PMA on proMMP-1 in media are inhibited by a PKC inhibitor, Ro320432 (2.0 \textmu M), and an inhibitor of activation of p42/44 MAP kinase, U0126 (10 \textmu M).
MMP-1, but this remains relatively modest, and following secretion proMMP-1 appears to be relatively stable in the media of these cells. A range of proteases may activate MMP-1, and it seems possible that in vivo activation is achieved by sub-epithelial cells. In this context it is worth noting that proteomic studies have demonstrated increased MMP-1 activation by myofibroblasts derived from gastric tumors compared with myofibroblasts from adjacent control tissue (14). While epithelial expression of proMMP-1 is regulated by gastrin, therefore, the functional significance of this depends on activation, which in turn reflects the cellular microenvironment which differs in normal, inflammatory, premalignant, and malignant conditions.

A role for extracellular proteases in mediating cell migration is well recognized (30). These are involved in facilitating the progressive remodeling of cell-matrix interactions at both leading and trailing edges of migrating cells. Migration of gastric epithelial cells normally occurs over a period of days as cells move from the proliferative zone to the base of gastric glands (15), but there is also migration in response to wound healing, and in cancer migration may reflect an epithelial-mesenchymal transition. In scratch-wound assays in vitro, there is migration of individual AGS-GR cells, rather than an advancing sheet of cells, and both in this model and in Boyden chamber migration experiments gastrin is a good stimulant of AGS-GR cell migration. There is presently a need for the development of effective specific inhibitors of MMP-1, but immunoneutralization provides an alternative for present purposes (14). The present study revealed a role for MMP-1 in mediating the effects of gastrin migration. Multiple proteases are likely to be involved in the remodeling of cell-matrix attachments in migration, which may well account for the fact that there was only partial inhibition of the effects of gastrin by neutralizing antibody.

It is now clear that gastrin stimulates expression of a number of genes involved in mucosal organization and protection (9, 10). Although MMP-1 expression is not normally associated with epithelial cells, the epithelial cells of the acid-secreting mucosa of the stomach are an exception. In addition to possible roles for gastric epithelial MMP-1 in response to inflammation and infection, our data suggest that it is now appropriate to consider a role for this enzyme in mediating the effects of gastrin in the healthy stomach. Moreover, while chromogranin A and HDC are useful biomarkers of gastrin effects on ECL cells, we suggest that MMP-1 may prove to be a useful biomarker of gastrin actions on other glandular cells.

ACKNOWLEDGMENTS

We are grateful to J. Kozma and B. Balogh for help with immunohistochemistry, to A. Alqahtani for help with Western blotting, and to the staff of the endoscopy suite of the Gastrointestinal Unit, Royal Liverpool University Hospital.
GASTRIN STIMULATION OF MMP-1 EXPRESSION

G85

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