microRNA-192 suppresses the expression of the farnesoid X receptor

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Krattinger R, Boström A, Schiöth HB, Thasler WE, Mwinyi J, Kullak-Ublick GA. microRNA-192 suppresses the expression of the farnesoid X receptor. Am J Physiol Gastrointest Liver Physiol 310: G1044–G1051, 2016. First published April 14, 2016; doi:10.1152/ajpgi.00297.2015.—Farnesoid X receptor (FXR, NR1H4) plays an important role in the regulation of bile acid homeostasis in liver and intestine and may exert protective effects against certain forms of cancer such as colon carcinoma. However, the role of FXR in cell growth regulation, apoptosis, and carcinogenesis is still controversial. Similar to FXR, microRNA-192 (miR-192) is mainly expressed in the liver and colon and plays an important role in the pathogenesis of colon carcinoma. In this study, we investigated the extent to which FXR is regulated by miR-192. Two in silico-predicted binding sites for miR-192-3p within the NR1H4-3’ untranslated region (UTR) were examined in vitro by luciferase reporter assays. Wild-type and mutated forms of the NR1H4-3’UTR were subcloned into a pmirGLO vector and cotransfected into Huh-7 cells with miR-192-3p. To study the effects of miR-192 on the expression of FXR, FXR target genes and cell proliferation, Huh-7 and Caco-2 cells were transfected with miR-192-5p and -3p mimics or antagomirs. In addition, the correlation between FXR and miR-192 expression was studied by linear regression analyses in colonic adenocarcinoma tissue from 27 patients. MiR-192-3p bound specifically to the NR1H4 mRNA and protein levels as well as the mRNA levels of the FXR-inducible bile acid transporters OSTα-OSTβ and OATP1B3. Significant inverse correlations were detected in colonic adenocarcinoma between NR1H4 mRNA and miR-192-3p expression. In summary, microRNA-192 suppresses the expression of FXR and FXR target genes in vitro and in vivo.

miR-192; farnesoid X receptor; bile-acid transporters; drug-induced liver injury; colonic adenocarcinoma

NUCLEAR FARNESOID X RECEPTOR (FXR, NR1H4) is a ligand-activated transcription factor that plays a crucial role in the regulation of bile acid, cholesterol, lipid, and glucose homeostasis. It is mainly expressed in the liver, intestines, kidney, and adrenal glands (34). FXR regulates key genes involved in human bile acid synthesis and metabolism, including bile acid transporters (4). Studies on FXR knockout mice have shown that FXR exerts hepatoprotective effects. Diminished FXR expression has been linked to an increase in inflammatory responses and neoplastic transformation in mice (7, 16). Mice lacking FXR expression show elevations in serum and hepatic bile acid levels and a higher incidence of hepatocellular carcinoma. In this study, we investigated the extent to which FXR is regulated by miR-192.
(HCC), Lian et al. (19) showed a significantly suppressed expression of miR-192 in HCC tissue compared with nontumorous tissue. An important role of miR-192 targeting ZEB2 mRNA has also been found in HCC (13). In contrast, certain forms of cancer, such as cholangiocarcinoma and esophageal cancer, show increased miR-192 expression during carcinogenesis (21, 26).

In this study, we investigated to what extent miR-192 modulates the expression of FXR and thereby affects the expression of FXR target genes in liver and colon cancer-derived cell lines.

**MATERIALS AND METHODS**

**Bioinformatics.** An in silico search for possible miRNA-binding sites in the 3’ untranslated region (UTR) of the NR1H4 gene was performed by using miRANDA (Memorial Sloan-Kettering Cancer Center, New York, NY), DIANA-microT-CDS (Biomedical Science Research Center Alexander Fleming, Athens, Greece) and miRBase (Faculty of Life Science, University of Manchester, Manchester, UK). mRNA and miR expression data from 27 colon adenocarcinoma tissue samples [E-GEOD-29623, Affymetrix GeneChip Human Genome U133 Plus 2.0 and NIH TaqMan microRNA Array v2.0 (5)] were retrieved from the openly accessible platform ArrayExpress (EMBL-European Bioinformatics Institute, Wellcome Trust Genome Campus, Hinxton, UK).

**Cell culture.** The human hepatoma-derived Huh-7 and colon carcinoma-derived Caco-2 cell lines (American Type Culture Collection, Molsheim, France) were cultured in RPMI-1640 and Dulbecco’s modified Eagle’s medium (Life Technologies, Carlsbad, CA), respectively, supplemented with 10% fetal bovine serum (Sigma-Aldrich, St. Louis, MO), 100 U/ml penicillin, and 100 µg/ml streptomycin (Invitrogen, Carlsbad, CA). Primary human hepatocytes (PHHs) obtained from three patients suffering from primary or secondary liver carcinoma (ethical approval by the Local Ethical committee of the University of Munich, Germany, and the Ethics Committee of the Canton of Zurich, Switzerland) were isolated from the cancer-adjacent normal tissue and cultured as described (17). PHHs were kept in maintenance medium including ultraglutamine for 5 h before further procedures. Cell cultures were incubated at 37°C in a humidified atmosphere with 5% CO2.

**Transient transfection with miR-192.** To investigate the effect of miR-192 on NR1H4 mRNA and protein expression, Huh-7 and Caco-2 cells were seeded in 12-well plates (8×104 and 4×105 cells/ml, respectively). After 24 h, cells were transfected with 100 nM hsa-miRvANA miRNA mimics or anti-miR miRNA inhibitors (hsa-miR-192-3p/-5p; Life Technologies). RT-PCR was performed by using 1:5 diluted cDNA and 9.3 µl RT-PCR Universal Fast Master Mix (Applied Biosystems) including primers. U6snRNA was used as an internal control. Mean miR expression in three passages of cells was analyzed per cell line. All measurements were performed in triplicate.

**Western blot analysis.** Protein samples (17–30 µg) were diluted 1:5 with loading buffer, denatured, and separated by 8% SDS-polyacrylamide gel electrophoresis. The separated proteins were transferred to polyvinylidene fluoride membranes and preincubated for 1 h in blocking buffer and then for 16 h with the primary antibodies (anti-FXR, Santa Cruz Biotechnology, Dallas, TX; anti-β-actin, Abcam, Cambridge, UK) at 4°C. After four washing steps, the membranes were incubated with a horseradish peroxidase-conjugated secondary antibody (goat-anti-rabbit, Thermo Scientific, Waltham, MA) for 1 h at room temperature. Protein bands were visualized with SuperSignal West Femto Maximum Sensitivity Substrate (Thermo Scientific) and Fusion FX7 (Vilber Lourmat, Eberhardzell, Germany). β-Actin was used as a housekeeping gene.

**Transient cotransfection with miR-192-3p (miR-192*) and NR1H4 3’UTR plasmid constructs and luciferase reporter assays.** To examine the NR1H4 3’UTR as a target of miR-192-3p in vitro, luciferase assays were performed with a pmirGLO Dual-Luciferase miRNA Target Expression Vector (Promega, Fitchburg, WI), containing both the coding sequences of firefly luciferase and Renilla luciferase (internal control). The NR1H4 3’UTR (accession number NG_029843.1) was cloned into the pmirGLO vector system by using specific primers (Table 1). Plasmids were verified by Sanger sequencing. Huh-7 cells were seeded in 48-well plates (1.6×105 cells/ml). After 24 h, the cells were transfected with 50 nM hsa-miRvANA miRNA mimics or a negative control (hsa-miR-192-3p, Negative Control no. 1; Life Technologies) and 100 ng/well of plasmid DNA with use of Lipofectamine 2000 (Invitrogen). The activities of firefly and Renilla luciferases were measured at 24 h after cotransfection using the Dual-Luciferase Reporter 1000 Assay System (Promega) according to the manufacturer’s protocol. Hsa-miR-21-5p and its effect on the miR-21 target sequence were used as a positive control. Analysis was performed using the GloMax Multi Detection System (Promega). An empty pmirGLO vector was used as a background control. QuikChange Multi Site-Directed Mutagenesis and QuikChange II XL Site-Directed Mutagenesis Kits (Agilent Technologies, Santa Clara, CA) were used to introduce mutations into the miR-192-3p-binding sites using specific primers (Table 1). Three independent luciferase reporter assays were performed including wild-type and mutated target sequences.

**Cell proliferation and invasion assays.** Cell proliferation was determined by using the alamarBlue (Thermo Scientific) cell viability reagent according to the manufacturer’s instructions. Briefly, Huh-7 and Caco-2 cells were seeded in 96-well plates (7.5×104 and 2×104 cells/ml, respectively). After 24 h, cells were transfected with 100 nM miR-192 mimics, FXR siRNA, or the corresponding negative controls. After 48, 72, and 96 h 10 µl of alamarBlue was added to each well and cells were incubated for 4 h at 37°C. The absorbance was
Table 1. Primer sequences used for subcloning and mutagenesis

<table>
<thead>
<tr>
<th>Oligonucleotide</th>
<th>Sequence (5'-3')</th>
<th>Purpose</th>
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<tbody>
<tr>
<td>FXR_3UTR_FW</td>
<td>aggagtcttgaggtagattcagg</td>
<td>Subcloning of NR1H4 3' end into pmirGlo</td>
</tr>
<tr>
<td>FXR_3UTR_RV</td>
<td>agggtgcccaagttgggacactc</td>
<td>Subcloning of NR1H4 3' end into pmirGlo</td>
</tr>
<tr>
<td>FXRmut_miR192*-FW1</td>
<td>ggacattccttgattctgctgacagccctgtttgcctaatatttcgccg</td>
<td>Multisite-directed mutagenesis</td>
</tr>
<tr>
<td>FXRmut_miR192*-FW2</td>
<td>ggaggtagttccttggagccgaccccctgcttcattccgggct</td>
<td>Multisite-directed mutagenesis</td>
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<tr>
<td>FXRwt_miR192*-FW</td>
<td>gggtgattccttggagccgaccccctgcttcattccgggct</td>
<td>Site-directed mutagenesis</td>
</tr>
<tr>
<td>FXRwt_miR192*-RV</td>
<td>cggcgcggattggatgtaaatcactcttg</td>
<td>Site-directed mutagenesis</td>
</tr>
</tbody>
</table>

Recognition sites for the restriction enzymes are underlined in the primers used for subcloning. In silico-predicted binding sites for hsa-miR-192-3p are underlined in the mutagenesis primers, whereas mutations inserted into these sites are indicated in bold. FXRwt_miR192* primers were used to optimize the second binding site for miR-192-3p.

determined at 560/600 nm by using the GloMax Multi Detection System.

Cell invasion was determined with extracellular matrix-coated invasion chambers (QCM 24-well cell invasion assay, Millipore, Billerica, MA) according to the manufacturer’s instructions. Huh-7 and Caco-2 cells were harvested and resuspended in serum-free medium after pretreatment for 48 h with miR-192 mimics or negative control (cell density for transfection: 1 × 10^5 and 4 × 10^5 cells/ml, respectively). Then, 1 × 10^5 cells were plated into the invasion chamber, whereas the bottom well of the chamber contained 500 μl of the corresponding medium supplemented with 10% FBS. After 48 h of incubation, the invaded cells on the underside of the membrane were detached, lysed, and stained with CyQuant GR Dye (Millipore). Fluorescence was measured by use of a 490/510 – 570 filter set and GloMax Multi Detection System.

Statistical analysis. Paired one-sample t-tests were performed to compare the effects of miR-192 mimics, antagonirs, FXR siRNA and the corresponding negative controls on FXR mRNA/protein expression levels and on cell proliferation/invasion. Luciferase activities were compared between miR-192-3p- and mock-transfected cells by one-way analysis of variance. All data obtained from transfection experiments were compared with negative control mimics or inhibitors, in which expression levels in mock-transfected cells were defined as 1 (except for cell proliferation assays, where values were normalized to the 48-h time point of a particular condition). In general, the negative controls were not expected to bind to the NR1H4 3' UTR or to block the activity of endogenously expressed miR-192. Only experiments with verified positive controls were included in statistical analyses. The association between FXR and miR-192 expression levels was investigated in 27 colonic adenocarcinoma tissue samples by linear regression analyses excluding subjects that had undergone chemotherapy [E-GEOD-29623 (5)]. Values are shown as averages ± standard deviation. A P value of less than 0.05 was considered to be statistically significant. Statistical analyses were performed with R Software (version 2.15.2) and GraphPad Prism (version 5.04).

RESULTS

The mature forms of pre-miR-192 derived from the 5' and 3' strands of the precursor are detectable in Huh-7 and Caco-2 cells. The online database miRBase lists two mature sequences for the human miRNA precursor pre-miR-192, miR-192-5p, and miR-192-3p. Quantitative measurement of the endogenous expression levels showed the presence of both strands at considerable amounts in Huh-7, PHH, and Caco-2 cells. Mir#-3p was expressed 10-fold less in Huh-7 cells, 32-fold less in Caco-2 cells, and 39-fold less in PHH cells compared with miR#-5p. Furthermore, miR-192-3p showed a 9- to 10-fold higher expression in Caco-2 cells than in the hepatoma cell line or the primary human hepatocytes (data not shown).

Confirmation of the NR1H4 3' UTR as a target of miR-192-3p by luciferase reporter assays. The software tools miRANDA and DIANA-microT-CDS predicted binding of miR-192-3p to NR1H4 transcript positions 199–227 and 324–
(position relative to translational stop codon, NG_029843.1, Fig. 1A). No binding site within the NR1H4 3′UTR was predicted for miR-192-5p. Cotransfection of miR-192-3p mimics and the NR1H4 3′UTR target sequence into Huh-7 cells resulted in a 30% decrease in luciferase activity compared with the empty vector control (P < 0.01). In contrast, luciferase activity remained unaffected by cotransfection of the NR1H4 3′UTR target sequence carrying the miR-192 binding sites in the mutated form compared with the wild-type construct (Fig. 1B), indicating a negative interaction of miR-192-3p with the predicted binding sites in the NR1H4 3′UTR.

MiR-192 attenuates endogenous NR1H4 mRNA levels in Huh-7 and Caco-2 cells. To investigate the effect of miR-192 on endogenous FXR expression levels, Caco-2 and Huh-7 cells were transfected with 100 nM miR-192-3p or -5p mimics for 24 or 48 h. As shown in Fig. 2A, NR1H4 mRNA levels were repressed to 75 and 65% by miR-192-3p and -5p mimics, respectively, in Huh-7 cells (P < 0.05). In Caco-2 cells, a decrease by 15 and 28% in NR1H4 mRNA expression was detected after transfection with miR-192-3p and -5p mimics for 24 h (P < 0.05). To examine whether transfection of an antagomir could reverse the endogenous miR-192-dependent inhibitory effect on FXR expression, Caco-2 and Huh-7 cells were transfected with 100 nM anti-miR-192 inhibitors for 24 or 48 h, respectively. As shown in Fig. 2B, NR1H4 mRNA levels were increased significantly by anti-miR-192-3p inhibitors in Caco-2 cells as well as anti-miR-192-5p inhibitors in Huh-7 cells compared with the anti-miR miRNA inhibitor negative control (P < 0.05). Thus the miR-192-dependent effect on FXR regulation appeared to be stronger in Huh-7 cells compared with Caco-2 cells.

Fig. 2. Effect of miR-192-5p and -3p mimics (A) and inhibitors (B) on NR1H4 mRNA expression in Huh-7 and Caco-2 cells at 48 h and 24 h, respectively, after transfection. NR1H4 mRNA expression relative (rel.) to β-actin was determined by real-time PCR. The miR mimic/inhibitor negative control (NC) was used for normalization. Experiments were repeated 3 times. *P < 0.05; ns, not significant.

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Fig. 3. Effect of miR-192-5p and -3p mimics on FXR protein expression in Huh-7 cells at 72 h after transfection and in Caco-2 cells at 48 h after transfection (A). Anti-miRNA-dependent effect on FXR protein levels (B). Caco-2 and Huh-7 cells were treated with anti-miR-192-5p and -3p inhibitors for 48 and 72 h. FXR protein expression relative to β-actin was determined by Western blotting and densitometry. Bands of 1 representative blot are shown for each condition and cell line. Experiments were repeated 3 times. **P < 0.01, *P < 0.05; ns, not significant.
MIR-192 suppresses FXR protein translation in Huh-7 and Caco-2 cells. Consistent with the miRNA-dependent effects on NR1H4 mRNA expression, a 63% decrease in FXR protein expression was seen after transfection with miR-192-3p mimic in Huh-7 cells (P < 0.05). Weaker downregulation by 14% was observed after transfection of miR-192-5p. A decrease by 14 and 43% in FXR protein expression was observed in Caco-2 cells after transfection with miR-192-3p and -5p, respectively (Fig. 3A). As shown in Fig. 3B, the endogenous miR-192-dependent suppressive effect on FXR protein expression was reversed upon transfection with anti-miR-192-3p or -5p inhibitors. In Huh-7 cells, an increase in FXR protein levels was not seen following transfection of the miR-192-3p antagonist, which could be explained by the lower expression of the 3′ in relation to the 5′ strand in Huh-7 compared with Caco-2 cells.

MIR-192 suppresses the expression of key FXR target genes. The bile acid transporters organic-anion transporting polypeptide 1B3 (OATP1B3, SLCO1B3) and organic solute transporters α/β (OST alpha/beta, SLC51A/SLC51B) were chosen as model genes to examine the effect of miR-192 on FXR-regulated expression of transport proteins. Huh-7 cells were transfected with 100 nM miR-192-3p or -5p for 48 h. At 24 h after transfection, cells were treated with 50 μM CDCA for 24 h to activate FXR and compared with the miRNA mimic negative control. As shown in Fig. 4, SLCO1B3, SLC51A, and SLC51B mRNA levels were decreased by either one or both strands of miR-192, suggesting a reduction of gene transcription secondary to reduced expression of the transcriptional activator FXR. This was confirmed by simultaneous FXR knockdown and miRNA transfection experiment (data not shown).

Linear regression analyses reveal an inverse association between the expression levels of miR-192 and FXR in colon adenocarcinoma. To investigate whether the in vitro effects of miR-192 on FXR expression were reproducible in vivo, we analyzed 27 tissue samples from chemotherapy-untreated patients with primary colonic adenocarcinoma (Table 2). As shown in Table 3 and Fig. 5, a significant inverse association in expression was observed for the NR1H4 mRNA transcript coding for FXRα2(+) and hsa-miR-192-3p. FXRα2 isoforms represent the most abundantly expressed FXR protein forms in colonic tissue, whereas the isomer FXRα1 is predominantly expressed in liver (30). No significant associations were found for NR1H4 mRNA and hsa-miR-192-5p expression, showing that the 5′ strand does not confer pronounced NR1H4 mRNA degradation in vivo. These findings support the regulatory effects of miR-192 on FXR expression in hepatoma and colon cancer-derived cell lines observed in vitro.

MIR-192 exhibits suppressive effects on proliferation of Huh-7 and Caco-2 cells. To investigate the functional significance of the observed miR-192/FXR interaction, we performed cell proliferation assays using alamarBlue. As shown in Fig. 6, transfection with miR-192-3p mimic significantly reduces proliferation of Huh-7 and Caco-2 cells, whereby a stronger effect could be observed in the hepatoma cell line. These findings confirm the previously described suppressive effects of miR-192 on proliferation of the colon cancer cell lines HT-29, RKO, and HCT116 (27). Knockdown of FXR expression causes similar antiproliferative effects in Huh-7 cells. In contrast, reduction in FXR expression does not seem to have any influence on the proliferative potential of Caco-2 cells. By performing cell invasion assays, we saw a trend toward a suppressive effect for both strands of the miR-192 precursor molecule on cell invasion of Huh-7 cells; however, the results were not significant (data not shown). In a previous work, Lian et al. (19) showed that miR-192 could significantly downregulate cell invasion of Huh-7 cells.

DISCUSSION

The aim of this study was to investigate whether FXR is a target of miR-dependent, posttranscriptional gene regulation. Regulation of FXR expression by miR-421 as an oncogenic miR in biliary tract cancer has already been postulated (36). Our results show an additional, as yet uncharacterized role of miR-192 in regulating FXR expression.

### Table 2. Clinical characteristics of 27 patients with primary colonic adenocarcinoma

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<tr>
<td></td>
<td>female</td>
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<td>AJCC stage</td>
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<td></td>
<td>pT2</td>
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<td></td>
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<td></td>
<td>pT4</td>
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<td>Tumor grade</td>
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</tr>
<tr>
<td></td>
<td>2</td>
<td>20 (74.1)</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>4 (14.8)</td>
</tr>
<tr>
<td>Treatments</td>
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<td>27 (100)</td>
</tr>
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</table>

Clinical characteristics of the 27 patients showing primary colonic adenocarcinoma who were included into linear regression analysis. AJCC, American Joint Committee on Cancer.

### Table 3. Linear regression analysis of miR-192-3p expression with FXRα2(+) \( \times \) FXRα1(−), and miR-192-5p expression in primary colonic adenocarcinoma

<table>
<thead>
<tr>
<th>Transcript</th>
<th>Coefficient</th>
<th>Standard Error</th>
<th>P Value</th>
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<tr>
<td>FXRα2(+)</td>
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<td>0.66</td>
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</tr>
<tr>
<td>FXRα1(−)</td>
<td>1.87</td>
<td>0.70</td>
<td>1.90E-02</td>
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<tr>
<td>hsa-miR-192-3p</td>
<td>0.31</td>
<td>0.14</td>
<td>4.13E-02</td>
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Linear regression analysis of hsa-miR-192-3p with the NR1H4 mRNA transcripts coding for FXRα2(+) and FXRα1(−), and miR-192-5p expression in primary colonic adenocarcinoma tissue obtained from 27 patients. miR-192-3p inversely correlates with FXRα2(+) expression. Adjusted R²: 0.348.
Our in vitro experiments elucidated the inhibitory effect of miR-192 on FXR expression by transfecting colon and liver cell lines with miR-192 mimics and inhibitors. In the case of the 3’ strand, we found a significant miR-192-dependent inhibitory effect on NR1H4 mRNA and protein expression in Huh-7 cells, whereas in Caco-2 cells only transfection with the anti-miR-192-3p inhibitor showed a relevant effect. A possible explanation for the observed weaker inhibitory effect of the miR-192-3p mimic in Caco-2 cells, compared with that in Huh-7 cells, may be the more abundant expression of miR-192 in the colon cell line. The observed miR-192*-dependent effects on NR1H4 mRNA levels in both cell lines and our in vivo findings that showed a strong inverse miR-192* and FXR\(_{\alpha2}(+)\) correlation in colonic adenocarcinoma support our hypothesis of a strong endogenous effect of miR-192 on FXR expression. Degradation of NR1H4 mRNA transcripts can be explained by perfect complementary interference of the miR-192-3p seed sequence at the two in silico-predicted binding positions of the NR1H4-3’UTR. The 5’ strand of miR-192 also appeared to repress FXR gene expression in vitro, albeit through a different mechanism. Transfections with the corresponding antagomirs support our findings with both mimics in our model cell lines.

According to the miRBase database, about 80 different human miRNA precursors can yield two abundant mature miRNAs, i.e., the 5’ strand (miR-#-5p) and 3’ strand (miR-#-3p) with different seed sequences and mRNAs as binding targets. There is increasing evidence for interplay between the 5’ and 3’ strands of the same precursor molecule targeting the same group of genes, thereby reinforcing a certain phenotype (10), which can be supported by our data.

Hsa-miR-192 has been shown to play a crucial role in the pathogenesis of colon carcinoma, the third most common cancer in Western countries (6). Because of its cancer stage-dependent decline in expression, miR-192 has been suggested to be a potential biomarker to predict metastasis in colon
target genes and plays a role in the pathogenesis of liver and FXR regulation, which could affect the expression of FXR and OATP1B3. We show a new miR-dependent mechanism of significantly decreasing the expression of FXR target genes OSTα/β in a FXR-dependent manner.

It remains unclear whether restoration of miR-192 expression and the consequently diminished expression of FXR target genes would be beneficial for cancer patients, especially considering that miR-192 has been shown to possibly act as an oncogenic miRNA by downregulation of Smad interacting protein 1 in other inflammation-related cancers (21, 26, 35). Therefore, it can be speculated that diminished expression of the transport protein OSTα/OSTβ, a heterodimer-forming bile acid transporter important for the excretion of bile acids from hepatocytes and enterocytes, may be a mechanism of intracellular bile acid accumulation promoting inflammation and/or cancer development in certain cases.

By performing cell proliferation and invasion assays, we were able to confirm the previously described suppressive role of miR-192 in liver and colon cancer progression. We observed a miR-192-associated antiproliferative effect in Huh-7 cells that is stronger than in Caco-2 cells, where no noteworthy effects of the 3’ strand arm could be seen on FXR protein levels. Furthermore, knockdown of FXR did not show any influence on Caco-2 cell proliferation. These observations support the hypothesis that the antiproliferative effect of miR-192-3p may be to some extent FXR-dependent. Future studies have to elucidate to what extent the miR-192/FXR interplay supports or inhibits tumor pathogenesis. Our results are in line with reports showing that enhanced FXR expression and the associated altered expression of FXR-regulated drug-uptake transporters is related to chemoresistance in cancer patients. Treatment of the colorectal adenocarcinoma cell line LS174T with cisplatin leads to a cisplatin-resistant phenotype that is accompanied by a 350-fold increase in FXR expression (22). Furthermore, increased expression of OATP1B3 has been suggested to confer antiapoptotic resistance to paclitaxel by altering p53-dependent pathways (18).

It may be of value to confirm the detected association between miR-192 and FXR expression in larger cohorts. We cannot exclude the contribution of other miRs or epigenetic effects of the 3’ UTR.

In conclusion, miR-192-5p and -3p negatively regulate the expression of FXR in a synergistic manner, thereby significantly decreasing the expression of FXR target genes OSTα/β and OATP1B3. We show a new miR-dependent mechanism of FXR regulation, which could affect the expression of FXR target genes and plays a role in the pathogenesis of liver and colon cancers and their response to anticancer therapies.

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GRANTS

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

AUTHOR CONTRIBUTIONS


REFERENCES


